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Corresponding Author: David S. Hibbett,

Corresponding Author's Institution: Clark University

First Author: David S Hibbett, PhD

Order of Authors: David S Hibbett, PhD; David S. Hibbett

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Abstract: A comprehensive phylogenetic classification of the kingdom Fungi is proposed, with reference to recent molecular phylogenetic analyses, and with input from diverse members of the fungal taxonomic community. The classification includes 195 taxa, down to the level of order, of which 19 are described or validated here: Dikarya subkingdom nov.; Chytridiomycota, Neocallimastigomycota phyla nov.; Agaricomycetes, Dacrymycetes, Monoblepharidomycetes, Neocallimastigomycetes, Tremellomycetes class. nov.; Eurotiomycetidae, Lecanoromycetidae, Mycocaliciomycetidae subclass. nov.; Acarosporales, Corticiales, Baeomycetales, Candelariales, Gloeophyllales, Melanosporales, Trechisporales, Umbilicariales ords. nov. The clade containing Ascomycota and Basidiomycota is classified as subkingdom Dikarya, reflecting the putative synapomorphy of dikaryotic hyphae. The most dramatic shifts in the classification relative to previous works concern the groups that have traditionally been included in the Chytridiomycota and Zygomycota. The Chytridiomycota is retained in a restricted sense, with Blastocladiomycota and Neocallimastigomycota representing segregate phyla of flagellated Fungi. Taxa traditionally placed in Zygomycota are distributed among Glomeromycota and several subphyla incertae sedis, including

Mucoromycotina, Entomophthoromycotina, Kickxellomycotina, and Zoopagomycotina. Microsporidia are included in the Fungi, but no further subdivision of the group is proposed. Several genera of "basal" Fungi of uncertain position are not placed in any higher taxa, including Basidiobolus, Caulochytrium, Olpidium, and Rozella.



Department of Biology
950 Main Street
Worcester, MA 01610-1477

508-793-7173 Phone
508-793-8861 Fax
www.clarku.edu

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Dr. David L. Hawksworth
Chief Executive Editor
Mycological Research

Dear David:

Please find enclosed the revised version of our manuscript titled "A Higher-Level Phylogenetic Classification of the *Fungi*", intended for *Mycological Research*. Thank you very much for moving ahead so promptly with the review of our original submission, and also for your careful editing. Thanks to your work, and that of many other coauthors, there have been numerous minor corrections, updates to the literature, etc. Below, I describe the major changes that have appeared since you last handled the ms:

- Table 1 has been separated into three separate tables.
- Figure 1 has been separated into three separate figures. I did not replace these with color versions, but if you think that would improve clarity, then I will be happy to do so.
- MycoBank numbers have been provided for all new taxa (thanks to Joost Stalpers).
- John Taylor and Valerie Hofstetter decided they should not be listed as coauthors.
- Claude Roux, Jolanta Miadlikowska, Valerie Reeb, Cecile Gueidan, and Josph Ironside have been added as coauthors.
- It was discovered that *Agaricomycetidae* was described by Parmasto (1986), so that is no longer listed as a new taxon, and the abstract and main text have been modified accordingly. I inserted some comments regarding the different delimitations of *Agaricomycetidae* in the present work vs. Parmasto (1986).
- The citation for *Chytridiomycetes* was revised to Caval.-Sm. (1998). This is not an easy problem. I consulted the de Bary (1863) volume that was cited and it does not contain *Chytridiomycetes* (I looked at every page and checked the index). The text cites the usage of this name in Serbinow (1907), Cejp (1957), Sparrow (1958), and Alexopoulos *et al.* (1996). I also included a note directing readers to David (2002), which contains more information about the history of this name.
- Bibliographic information has been filled in for Fischer ("1891"), Schröter (1889), and Winter (1880). However, I was not able to find the Corner citation for *Thelephorales*. The Oberwinkler citation should be adequate, I think.
- References to *Mycologia* 98(6) have been changed from 2006 to 2007, including the tables. However, I have not included page numbers for this issue, because it is quite possible that repagination will be needed after corrections of errors that remained in the proofs. I will add page numbers for this issue at the proof stage.

- I hope you will not mind, but I deleted the text formerly on p. 10 that explained that taxa such as Hyphomycetes and Coelomycetes are no longer needed. Instead, I simply noted that the classification is restricted to *Fungi*...including sexual and asexual forms." I think that the text as worded could have been interpreted (justly or otherwise) as implying that all names based on asexual forms at any rank are superfluous, and that is a swamp that I do want to wade into at this time.
- I gather that you are seeing boxes around the references for Ekman and Tønsberg (2002) and Suh *et al.* (2007). I'm afraid that I don't see them, and therefore cannot remove them. Perhaps this is a Mac-PC compatibility issue in MS Word?
- Your queries inserted in the text have been removed.

I hope that these revisions are adequate. Again, thank you very much for considering this ms for publication in *Mycological Research*.

Sincerely,



David S. Hibbett

Associate Professor

Tel.: (508) 793-7332

Fax: (508) 793-7174

E-mail: dhibbett@clarku.edu

Lab homepage: <http://www.clarku.edu/faculty/dhibbett/index.html>

A higher-level phylogenetic classification of the *Fungi*

David S. HIBBETT¹, Manfred BINDER¹, Joseph F. BISCHOFF², Meredith BLACKWELL³,
Paul F. CANNON⁴, Ove E. ERIKSSON⁵, Sabine HUHNDORF⁶, Timothy JAMES⁷, Paul M.
KIRK⁴, Robert LÜCKING⁶, Thorsten LUMBSCH⁶, François LUTZONI⁷, P. Brandon
MATHENY¹, David J. MCLAUGHLIN⁸, Martha J. POWELL⁹, Scott REDHEAD¹⁰, Conrad L.
SCHOCH¹¹, Joseph W. SPATAFORA¹¹, Joost A. STALPERS¹², Rytas VILGALYS⁷, M.
Catherine AIME¹³, André APTROOT¹⁴, Robert BAUER¹⁵, Dominik BEGEROW¹⁶, Gerald L.
BENNY¹⁷, Lisa A. CASTLEBURY¹³, Pedro W. CROUS¹², Yu-Cheng DAI¹⁸, Walter GAMS¹²,
David M. GEISER¹⁹, Gareth W. GRIFFITH²⁰, Cécile GUEIDAN⁷, David L.
HAWKSWORTH²¹, Geir HESTMARK²², Kentaro HOSAKA²³, Richard A. HUMBER²⁴, Kevin
HYDE²⁵, Joseph E. IRONSIDE²⁰, Urmas KÕLJALG²⁶, Cletus P. KURTZMAN²⁷, Karl-Henrik
LARSSON²⁸, Robert LICHTWARDT²⁹, Joyce LONGCORE³⁰, Jolanta MIADLIKOWSKA⁷,
Andrew MILLER³¹, Jean-Marc MONCALVO³², Sharon MOZLEY-STANDRIDGE³³, Franz
OBERWINKLER¹⁵, Erast PARMASTO³⁴, Valérie REEB⁷, Jack D. ROGERS³⁵, Claude ROUX³⁶,
Leif RYVARDEN³⁷, José Paulo SAMPAIO³⁸, Arthur SCHUESSLER³⁹, Junta SUGIYAMA⁴⁰,
R. Greg THORN⁴¹, Leif TIBELL⁴², Wendy A. UNTEREINER⁴³, Christopher WALKER⁴⁴,
Zheng WANG¹, Alex WEIR⁴⁵, Michael WEISS¹⁵, Merlin M. WHITE⁴⁶, Katarina WINKA⁵, Yi-
Jian YAO⁴⁷, Ning ZHANG⁴⁸

¹Biology Department, Clark University, Worcester, MA 01610, USA

²National Library of Medicine, National Center for Biotechnology Information, 45 Center
Drive, Bethesda, MD 20892-6510, USA

³Department of Biological Sciences, Louisiana State University, Baton Rouge, LA 70803, USA

⁴CABI UK, Bakeham Lane, Egham, Surrey TW20 9TY, UK

⁵Department of Ecology and Environmental Science, Umeå University, SE-901 87 Umeå, Sweden

⁶Department of Botany, The Field Museum of Natural History, 1400 South Lake Shore Drive, Chicago, IL 60605-2496, USA

⁷Department of Biology, Duke University, Box 90338, Durham, NC 27708, USA⁸Department of Plant Biology, University of Minnesota, 1445 Gortner Avenue, St Paul, MN 55108-1095, USA

⁹Department of Biological Sciences, Box 870344/319 Biology, University of Alabama, Tuscaloosa, AL 35487-0344, USA

¹⁰ECORC, Agriculture and Agri-Food Canada, CEF, Neatby Building, Ottawa, ON K1A 0C6, Canada

¹¹Department of Botany and Plant Pathology, Oregon State University, 2082 Cordley Hall, Corvallis, OR 97331, USA

¹²Centraalbureau voor Schimmelcultures, Uppsalalaan 8, NL-3584 CT Utrecht, The Netherlands

¹³USDA ARS Systematic Botany and Mycology Laboratory, Bldg 011A Rm 319 BARC-WEST, 10300 Baltimore Ave, Beltsville, MD 20705 USA

¹⁴ABL Herbarium, Gerrit van der Veenstraat 107, NL-3762 XK Soest, The Netherlands

¹⁵Botanical Institute, University of Tübingen, Auf der Morgenstelle 1, D-72076 Tübingen, Germany

¹⁶Max-Planck-Institute for Terrestrial Microbiology, Karl-von-Frisch-Strasse, D-35043 Marburg, Germany

¹⁷Department of Plant Pathology, 1453 Fifield Hall, Hull Road, P.O. Box 110680, University of Florida, Gainesville FL 32611-0680, USA

¹⁸*Institute of Applied Ecology, Chinese Academy of Sciences, Shenyang 110016, China*

¹⁹*Department of Plant Pathology, Pennsylvania State University, University Park, PA 16802, USA*

²⁰*Institute of Biological Sciences, University of Wales, Aberystwyth SY23 3DA, UK*

²¹*Departamento de Biología Vegetale II, Facultad de Farmacia, Universidad Complutense de Madrid, Plaza Ramón y Cajal, Ciudad Universitaria, E-28040 Madrid, Spain*

²²*Department of Biology, University of Oslo, P.O. Box 1066, Blindern, N-0316 Oslo, Norway*

²³*Department of Botany, The Field Museum, 1400 S Lake Shore Dr., Chicago, IL 60605, USA,*

²⁴*USDA-ARS Plant Protection Research Unit, US Plant, Soil and Nutrition Laboratory, Tower Road, Ithaca, NY 14853-2901, USA*

²⁵*Centre for Research in Fungal Diversity, Department of Ecology & Biodiversity, University of Hong Kong, Pokfulam Road, Hong Kong SAR, China*

²⁶*Institute of Botany and Ecology, Tartu University, 40 Lai Street, Tartu 51005, Estonia*

²⁷*Microbial Properties Research, NCAUR, ARS, USDA, 1815 N.University Street, Peoria, IL 61604-3999, USA*

²⁸*Botanical Institute, Göteborg University, Box 461, SE-405 30 Göteborg, Sweden*

²⁹*Department of Ecology and Evolutionary Biology, University of Kansas, Lawrence, KS 66045-7534, USA*

³⁰*Department of Biological Sciences, University of Maine, 5722 Deering Hall, Orono, ME 04469-5722, USA*

³¹*Center for Biodiversity, Illinois Natural History Survey, 1816 South Oak Street, Champaign, IL 61820-6970, USA*

³²*Royal Ontario Museum, Centre for Biodiversity and Conservation Biology, 100 Queen's Park, Toronto, ON M5S 2C6, Canada*

³³*Division of Natural Sciences, Mathematics, and Engineering, Middle Georgia College, 1100 Second Street, SE, Cochran, GA 31014-1599, USA*

³⁴*Institute of Agricultural and Environmental Sciences, Estonian University of Life Sciences, 181 Riia Street Tartu, 51014, Estonia*

³⁵*Department of Plant Pathology, Washington State University, P.O. Box 646430, Pullman, WA 99164-6430, USA*

³⁶*Chemin des Vignes vieilles, 84120, Mirabeau, France*

³⁷*Botanical Institute, University of Oslo, P.O. Box 1066, Blindern, N-0316 Oslo, Norway*

³⁸*CREM, SABB, Faculdade de Ciências e Tecnologia, Universidade Nova de Lisboa, 2829-516 Caparica, Portugal*

³⁹*Genetics Institute, Department Biology I, Maria-Ward-Strasse 1a, D-80638 Munich, Germany*

⁴⁰*TechnoSuruga Co. Ltd, Tokyo Office, Ogawamachi Kita Building 4F, 1-8-3, Kanda Ogawamachi, Chiyoda-ku, Tokyo 101-0052, Japan*

⁴¹*Department of Biology, University of Western Ontario, 213 Biological and Geological Sciences Building, 1151 Richmond Street North, London, ON, N6A 5B7, Canada*

⁴²*Department of Systematic Botany, Evolutionary Biology Centre, Uppsala University, Norbyvägen 18 D, SE-752 36, Uppsala, Sweden*

⁴³*Botany Department, Brandon University, 270-18th Street, Brandon, MB R7A 6A9, Canada*

⁴⁴*Biological Research and Imaging Laboratory, 2 Penny Hedge, New Milton, Hampshire BH25 7TB, UK*

⁴⁵*Faculty of Environmental and Forest Biology, SUNY College, 242 Illick Hall, 1 Forestry Drive, Syracuse, NY 13210, USA*

⁴⁶*Department of Biology, Boise State University, 1910 University Drive, S/N Building, Room 210, Boise, ID 83725-1515, USA*

⁴⁷*Systematic Mycology and Lichenology Laboratory, Institute of Microbiology, Chinese Academy of Sciences, Beijing 100080, China*

⁴⁸*Department of Plant Pathology, Cornell University, Geneva, NY 14456, USA*

*Corresponding author.

E-mail address: dhibbett@clarku.edu

Telephone: 1-508-793-7332

Fax: 1-508-793-7174

ABSTRACT

A comprehensive phylogenetic classification of the kingdom *Fungi* is proposed, with reference to recent molecular phylogenetic analyses, and with input from diverse members of the fungal taxonomic community. The classification includes 195 taxa, down to the level of order, of which 19 are described or validated here: *Dikarya* subkingdom nov.; *Chytridiomycota*, *Neocallimastigomycota* phyla nov.; *Agaricomycetes*, *Dacrymycetes*, *Monoblepharidomycetes*, *Neocallimastigomycetes*, *Tremellomycetes* class. nov.; *Eurotiomycetidae*, *Lecanoromycetidae*, *Mycocaliciomycetidae* subclass. nov.; *Acarosporales*, *Corticiales*, *Baeomycetales*, *Candelariales*, *Gloeophyllales*, *Melanosporales*, *Trechisporales*, *Umbilicariales* ords. nov. The clade containing *Ascomycota* and *Basidiomycota* is classified as subkingdom *Dikarya*, reflecting the putative synapomorphy of dikaryotic hyphae. The most dramatic shifts in the classification relative to previous works concern the groups that have traditionally been included in the *Chytridiomycota* and *Zygomycota*. The *Chytridiomycota* is retained in a restricted sense, with *Blastocladiomycota* and *Neocallimastigomycota* representing segregate phyla of flagellated *Fungi*. Taxa

traditionally placed in *Zygomycota* are distributed among *Glomeromycota* and several subphyla *incertae sedis*, including *Mucoromycotina*, *Entomophthoromycotina*, *Kickxellomycotina*, and *Zoopagomycotina*. *Microsporidia* are included in the *Fungi*, but no further subdivision of the group is proposed. Several genera of “basal” *Fungi* of uncertain position are not placed in any higher taxa, including *Basidiobolus*, *Caulochytrium*, *Olpidium*, and *Rozella*.

Keywords: AFTOL, *Eumycota*, Lichens, Molecular phylogenetics, *Mycota*, Nomenclature, Systematics

Introduction

The molecular revolution in fungal taxonomy commenced in the early 1990s, with analyses of PCR-amplified ribosomal RNA genes (White *et al.* 1990). Today, fungal molecular systematics is a mature discipline in which multi-locus datasets, extensive taxon sampling, and rigorous analytical approaches are standard. To gain an overview of the current state of the science it is only necessary to survey the recent “Deep Hypha” issue of *Mycologia* (98(6), 2007 [“2006”]), which contains 21 phylogenetic studies, all of which employ multiple genes to some extent (in some cases, multiple rRNA genes) and that address broad relationships in every major group of *Fungi* (except *Microsporidia*). Another recent milestone is the kingdom-level study of James *et al.* (2006), which used a dataset of six genes (nuc-ssu, -lsu, and 5.8S rRNA, *rpb1*, *rpb2* and *tef1*) sampled in nearly 200 species from every major clade of *Fungi* (including *Microsporidia*).

As the broad outlines of fungal phylogeny have come into focus, there have been repeated attempts to summarize the state of knowledge and to restructure higher-level

classifications. Two important works that have influenced fungal taxonomy in the 21st century are *Ainsworth & Bisby's Dictionary of the Fungi* (9th edn: Kirk *et al.* 2001), which contains a comprehensive kingdom-wide classification down to the level of genus, and *The Mycota VII* (McLaughlin *et al.* 2001a, b), an edited volume with chapters on all major groups of *Fungi*. These publications represented major advances toward a phylogenetic classification of *Fungi*, but they are already out of date. In the five years since the last edition of the *Dictionary* and the *Mycota VII* appeared, more than 360 articles with the key word "phylogen*" were published in *Mycologia* and *Mycological Research* alone, and approximately 80 % of the more than 100 000 fungal rRNA gene sequences now in GenBank were deposited (some by molecular ecologists). Recent publications that survey the entire fungal kingdom based on molecular phylogenies include the chapter by Taylor *et al.* in *Assembling the Tree of Life* (Cracraft & Donoghue 2004), the "New Higher Level Classification of Eukaryotes" (Adl *et al.* 2005), and the first large collaborative analysis of the Assembling the Fungal Tree of Life (AFTOL) project (Lutzoni *et al.* 2004). Taxonomic studies on individual groups of *Fungi* are too numerous to list. Two notable highlights include proposals to recognize the phylum *Glomeromycota* (Schüßler *et al.* 2001) and to include the *Microsporidia* within the *Fungi* (Keeling *et al.* 2000).

On-line fungal taxonomies are also proliferating. One of the most important on-line general classifications of *Fungi* is that of GenBank (www.ncbi.nlm.nih.gov/Taxonomy), which serves a diverse community of researchers, including ecologists and molecular biologists. Another highly visible on-line classification is that of the Tree of Life Web Project (tolweb.org/tree), which is widely used by teachers and students. The classification of *Ascomycota* is being updated regularly *via* the on-line Myconet series (www.fieldmuseum.org/myconet), and this has

been the basis for recent revisions at GenBank, but there is no comparable on-line resource for other major groups of *Fungi*. It is likely that on-line taxonomies will take on even greater prominence in the future, especially as they become integrated with databases of taxonomic names, particularly Index Fungorum (www.indexfungorum.org), MycoBank (www.mycobank.org), and other global biodiversity informatics resources (e.g. Global Biodiversity Information Facility, www.gbif.org).

While there is broad agreement regarding the composition of the major clades of *Fungi*, there is considerable variation in the names that have been applied to these groups. For example, the clade that is called *Basidiomycetes* in the latest edition of the *Dictionary* is called *Hymenomycetes* at GenBank. Similarly, the clade that is called *Ascomycetes* in the *Dictionary of the Fungi* is called *Pezizomycotina* in Myconet. Such inconsistencies create confusion, especially for students and non-specialists, and they hamper efforts to develop taxonomic databases.

There is consequently a pressing need for the fungal systematics community to adopt a consensus higher-level classification for the *Fungi* that is based on well-supported monophyletic groups, and which can be recommended for general use. This is an opportune moment to create such a classification. With the new multi-locus analyses, many nodes that were not previously resolved are now supported with confidence. The timing is also good because there are multiple projects in progress that seek to create or update broad classifications of the *Fungi*. In particular, a tenth edition of the *Dictionary* is in preparation, as is a fourth edition of an influential textbook of mycology (Alexopoulos *et al.* 1996). The classifications used by GenBank, the Tree of Life Web Project, and Myconet are being revised continuously. If the classifications employed by these and other major taxonomic resources could be unified, it would

promote communication and awareness of fungal phylogeny, and provide a framework for future revisions at all taxonomic levels.

This article presents a higher-level classification for all groups of *Fungi*, with reference to recent molecular phylogenetic studies. The authors represent diverse fungal taxonomy projects, including *Ainsworth & Bisby's Dictionary of the Fungi* (Cannon, Kirk, Stalpers), GenBank (Bischoff), Myconet (Eriksson, Lumbsch, Huhndorf), and Alexopoulos' mycology text (Blackwell, Spatafora). Many of the authors are contributors to the *Fungi* pages in the Tree of Life Web Project. Discussions leading to this classification began in 2004, under the auspices of the AFTOL project and the Deep Hypha Research Coordination Network (Blackwell *et al.* 2006), which were supported by the US National Science Foundation. Throughout the development of this classification, every effort has been made to work in a transparent, consultative manner. The first draft classification was presented at the 2005 Deep Hypha meeting (Tucson, AZ) and subsequently was distributed to a group of 100 fungal systematists for comment. The classification was revised based on comments received and was posted on the AFTOL classification project web site (www.clarku.edu/faculty/dhibbett/AFTOL/AFTOL.htm). Additional modifications were made following the 2006 Deep Hypha meeting (Baton Rouge, LA). For example, the classification of the *Pucciniomycotina* was revised to reflect the classification of Bauer *et al.* (2006). The present paper represents a first attempt at a broad-based consensus classification of the *Fungi*. However, the first 20 authors have exercised editorial control and are therefore to be held accountable for errors.

Structure and Principles

This classification is restricted to organisms that belong in the monophyletic kingdom *Fungi*, including sexual and asexual forms. It does not consider other organisms formerly included in the kingdom but which are now known not to belong there, even if still studied by mycologists, such as the oomycetes and slime moulds.

The classification adopted here uses a Linnean hierarchy as modified by the *International Code of Botanical Nomenclature* [Code] (McNeill *et al.* 2006), and uses seven ranks, including: order (suffix: *-ales*), subclass (*-mycetidae*), class (*-mycetes*), subphylum (*-mycotina*), phylum (*-mycota*; except *Microsporidia*), subkingdom, and kingdom. The rankings of taxa reflect the preferences and past practices of various authors, as well as the need to keep the nested hierarchies of clades and Linnean categories parallel. Taxa placed at the same rank are not necessarily equivalent in age (except sister taxa), number of species, or degree of morphological divergence.

The classification is limited to taxa down to the level of order. In many orders, especially those representing larger groups, such as *Agaricales*, there is still not enough resolution or taxon sampling to structure a comprehensive family-level classification. The challenge of creating family-level classifications is made even more difficult by the Code (McNeill *et al.* 2006), which requires that names of taxa at the rank of family or lower follow the principle of priority (which does not apply to higher ranks). Ideally, construction of consensus classifications within many of the orders recognized here will involve the coordinated efforts of groups of taxonomic specialists. It is hoped that the present classification will facilitate those endeavors.

The taxa included here are all supported as monophyletic by at least one published phylogenetic analysis (not applicable to monotypic taxa), with the exception of the *Lahmiales* and *Triblidiales* (*Pezizomycotina*) and *Asellariales* (*Kickxellomycotina*), for which molecular data are not available. Support for the monophyly of each group is

summarized in Tables 1-3, which list selected phylogenetic studies, the type of data that were analyzed, the number of OTUs sampled, and bootstrap frequencies and Bayesian posterior probabilities. No attempt has been made to cite all of the relevant studies for each group. The analyses chosen for inclusion in the Tables are those that have the greatest numbers of loci or taxa, and that provide the strongest support for monophyly of the clades in question. To supplement the information in the tables, brief comments on synonyms, phylogenetic relationships, and composition are provided below for some taxa, along with bibliographic citations for all taxon names. However, it is beyond the scope of this article to discuss each taxon in detail. For additional literature on the phylogeny and taxonomy of individual taxa, readers should consult the studies listed in Tables 1-3 and below, and the references therein.

The classification is also presented as a set of tree diagrams (Figs 1-3). Taxa of uncertain position are listed as *incertae sedis*, and have been placed at the least inclusive level in the hierarchy where they can be assigned with confidence. There are several nodes resolved in the tree figure that are not reflected in the classification (Figs 1-3). These unnamed clades, for which there is strong to moderate support in recent studies, include the *Dacrymycetes* plus *Agaricomycetes* (*Basidiomycota*) (Matheny *et al.* 2006, 2007a), *Saccharomycotina* plus *Pezizomycotina* (*Ascomycota*) (James *et al.* 2006; Spatafora *et al.* 2007), and the inoperculate euascomycetes (*Ascomycota*) (e.g. Lumbsch *et al.* 2002). The inoperculate euascomycetes has been recognized as a superclass, the *Leotiomyceta* (Eriksson & Winka 1997; Lumbsch *et al.* 2002), which is a rank that is not employed here, while the *Dacrymycetes* plus *Agaricomycetes* correspond to the subclass *Hymenomycetidae* of Swann & Taylor (1995). The absence of these groups from the present classification should not be interpreted as a judgment on their monophyly. Rather, it reflects a desire to keep the classification simple, and to minimize the number

of intercalary ranks (as per the directives of Art. 4.3 of the *Code*). Future revisions to this classification will have to consider how to incorporate additional deep nodes, including those that will be resolved with the application of genome-scale datasets (Galagan *et al.* 2005; Kuramae *et al.* 2006; Robbertse *et al.* 2006). One possibility is to employ an unranked category (with or without a uniform suffix) that could be inserted at any level in the taxonomic hierarchy (Hibbett & Donoghue 1998). For example, an unranked classification was adopted in part by Adl *et al.* (2005).

Overview of the Classification

The classification accepts one kingdom, one subkingdom, seven phyla, ten subphyla, 35 classes, 12 subclasses, and 129 orders. Taxa that are described or validated here include *Chytridiomycota*, *Monoblepharidomycetes*, *Neocallimastigomycota*, *Neocallimastigomycetes*, *Dikarya*, *Acarosporales*, *Baeomycetales*, *Candelariales*, *Umbilicariales*, *Lecanoromycetidae*, *Eurotiomycetidae*, *Mycocaliciomycetidae*, *Melanosporales*, *Tremellomycetes*, *Dacrymycetes*, *Agaricomycetes*, *Corticiales*, *Gloeophyllales* and *Trechisporales*. Thus, 90 % of the 195 taxon names employed in the present classification have been validly published previously. The clade containing the *Ascomycota* and *Basidiomycota* is classified as the subkingdom *Dikarya* (as used in James *et al.* 2006), reflecting the putative synapomorphy of dikaryotic hyphae (Tehler 1988). All of the other new names are based on automatically typified teleomorphic names. The classification of *Ascomycota* largely parallels that of the Myconet classification, including recent changes that will be adopted in the forthcoming 2007 "Outline of the *Ascomycota*". In *Basidiomycota*, the clades formerly called *Basidiomycetes*, *Urediniomycetes*, and *Ustilaginomycetes* in the last edition of *Ainsworth & Bisby's Dictionary of the Fungi* are called the *Agaricomycotina*,

Pucciniomycotina, and *Ustilaginomycotina*, respectively, as in Bauer *et al.* (2006). This is done to minimize confusion between taxon names and informal terms (basidiomycetes is a commonly used informal term for all *Basidiomycota*) and to refer to the included genera *Agaricus* (including the cultivated button mushroom) and *Puccinia* (which includes barberry-wheat rust). Another significant change in the *Basidiomycota* classification is the inclusion of the *Wallemiomycetes* and *Entorrhizomycetes* as classes *incertae sedis* within the phylum, reflecting ambiguity about their higher-level placements (Matheny *et al.* 2007b).

The most dramatic changes in the classification concern the “basal fungal lineages”, which include the taxa that have traditionally been placed in the *Zygomycota* and *Chytridiomycota*. These groups have long been recognized to be polyphyletic, based on analyses of rRNA, *tef1*, and *rpb1* (James *et al.* 2000; Nagahama *et al.* 1995; Tanabe *et al.* 2004, 2005). The recent multilocus analyses of James *et al.* (2006) and others now provide the sampling, resolution, and support necessary to structure new classifications of these early-diverging groups, although significant questions remain. The *Chytridiomycota* is retained in a highly restricted sense, including *Chytridiomycetes* and *Monoblepharidomycetes*. The *Blastocladales*, a traditional member of the *Chytridiomycota*, is here treated as a phylum, the *Blastocladiomycota*, as in James *et al.* (2007). The *Neocallimastigales*, whose distinctiveness from other chytrids has long been recognized, is also elevated to phylum, based on both morphology and molecular phylogeny. The genera *Caulochytrium*, *Olpidium*, and *Rozella*, which have traditionally been placed in the *Chytridiomycota*, and *Basidiobolus*, which has been classified in the *Zygomycota* (*Entomophthorales*), are not included in any higher taxa in this classification, pending more definitive resolutions of their placements.

The phylum *Zygomycota* is not accepted in this classification, pending resolution of relationships among the clades that have traditionally been placed in the *Zygomycota* (see discussion under *Mucoromycotina*). The traditional *Zygomycota* are here distributed among the phylum *Glomeromycota* and four subphyla *incertae sedis*, including *Mucoromycotina*, *Kickxellomycotina*, *Zoopagomycotina* and *Entomophthoromycotina*. A clade containing the *Glomeromycota* and the *Dikarya* was resolved previously based on ribosomal RNA genes and was classified as the *Symbiomycotina* (Tehler *et al.* 2003). That taxon is not included here, because there was not strong support for the clade in the analyses of James *et al.* (2006) or Liu *et al.* (2006). If the *Symbiomycotina* is added to this classification, it will need to be assigned a rank between kingdom and subkingdom, or perhaps be classified as an unranked taxon.

Microsporidia, unicellular parasites of animals and protists with highly reduced mitochondria (Germot *et al.* 1997; Hirt *et al.* 1997; Peyretailade *et al.* 1998), are included here as a phylum of the *Fungi*, based on analyses by Keeling *et al.* (2000), Gill & Fast (2006), James *et al.* (2006), and Liu *et al.* (2006). The latter study concluded that *Microsporidia* are the sister group of the rest of the *Fungi* and should not be classified as true *Fungi*, but that topology does not conflict with the delimitation of the monophyletic *Fungi* as proposed here. The analysis of James *et al.* (2006) suggested that *Rozella*, which was not sampled by Liu *et al.* (2006), is the sister group of the *Microsporidia*. No subdivision of the *Microsporidia* is proposed, owing to a lack of well-sampled multilocus analyses of this group (but see Vossbrinck & Debrunner-Vossbrinck 2005, for an analysis using SSU rRNA genes).

Phylogenetic classification of *Fungi*

Many of the citations and authorities in the list below were obtained from the Index Fungorum databases www.indexfungorum.org. A brief list of exemplar genera, including the type for automatically typified names, is given for each order (for small orders, all included genera are listed). A number of the genera listed are used in a modern, restricted sense, and readers are urged to consult the primary literature cited below and in Tables 1-3 for information about current generic concepts. Comprehensive lists of genera and families included in each order will be forthcoming in the *Dictionary of the Fungi* (10th edn; listing on-line at www.indexfungorum.org) and in the next revision of Myconet (for *Ascomycota*). Further information on the names of fungi (not only kingdom *Fungi*) above the rank of order and their places of publication may be found in the preliminary catalogue compiled by David (2002).

In accordance with the practice in recent editions of the *Code*, all scientific names regardless of rank are placed in *italic* type here except in the first line of the treatment of each accepted taxon where they are given in **bold** Roman type to make them stand out. When these names are used by other mycologists in their own publications, we wish to encourage the practice of the use of italics as recommended in the Preface to the current *Code* (McNeill *et al.* 2006).

Kingdom: **Fungi** T. L. Jahn & F. F. Jahn ex R. T. Moore, *Bot. Mar.* **23**: 371 (1980).

Synonym: *Fungi* T. L. Jahn & F. F. Jahn, *How to Know the Protozoa*: 7 (1949), *nomen nudum*.

The concept of the *Fungi* as one of six kingdoms of life was introduced by Jahn & Jahn (1949), and a five kingdom system was advanced by Whittaker (1959), but neither of these works included a Latin diagnosis and the name was therefore invalid under the *Code* until the required Latin was provided by Moore (1980). Although Moore did not

make a specific reference to Jahn & Jahn's book, he was well aware that the name was in widespread use in the rank of kingdom, and it is therefore fitting to include the authors who first did this in the citation.

Phylum: **Chytridiomycota** M. J. Powell, **phylum nov.**

MycoBank no.: MB 501278

Synonyms: *Archemycota* Caval.-Sm., *Biol. Rev.* **73**: 246 (1998), *pro parte*.

Thallus monocentricus vel polycentricus vel filamentosus; propagatio asexualis zoosporis, flagello retrorsum inserto, kinetosomate et centriolo supervacaneo praeditis, 9 munimentis flagelli, et complexu "microbody-corpore lipideo" descriptis; propagatio sexualis meiosi post copulationem perfecta; apparatus Golgi e cisternis superimpositis constans; tegumentum nuclei mitosi procedente circum polos fenestratum.

Typus: Chytridium A. Braun 1851.

Thallus monocentric, polycentric, or filamentous; asexual reproduction by zoospores with a single posteriorly-directed flagellum, both a kinetosome and non-functional centriole, nine flagellar props, and a microbody-lipid globule complex; sexual reproduction with zygotic meiosis where known; Golgi apparatus with stacked cisternae; nuclear envelope fenestrated at poles during mitosis.

Used as a phylum name without Latin diagnosis or description among others by von Arx (1967) and Margulis *et al.* (1990). Equivalent to euchytrids of James *et al.* 2006, the "core chytrid clade" of James *et al.* (2007), or the "core chytrid clade" plus the *Monoblepharidales* of James *et al.* (2000). Earlier usages are not indicated in the author citation of the name, because the circumscription adopted here differs significantly from that of those authors.

Class: **Chytridiomycetes** Caval.-Sm., *Biol. Rev.* **73**: 246 (1998).

Synonym: *Archimycetes* A. Fisch. (Fischer 1892) *pro parte* (included *Olpidiopsis*, *Hypochytrium*).

Type: Chytridium A. Braun 1851.

Reproducing asexually by zoospores bearing a single posteriorly-directed flagellum; zoospores containing a kinetosome and a non-flagellated centriole; thallus monocentric or rhizomycelial polycentric; sexual reproduction not oogamous.

Cavalier-Smith (1998) provided a brief, four-word, Latin description that was not diagnostic for phyla of uniflagellate fungi, and has been revised above. The name *Chytridiomycetes* was also used by Serbinow (1907), Cejp (1957), Sparrow (1958), and Alexopoulos *et al.* (1996). For further discussion of the nomenclatural history of the name, see David (2002).

Order: **Chytridiales** Cohn, *Jber. schles. Ges. vaterl. Kultur* **57**: 279 (1879).

Emend. Schröter (as '*Chytridineae*') in Engler & Prantl, *Nat. Pflanzenfam.* **1**(1): 64 (1892).

Emend. Barr, *Can. J. Bot.* **58**: 2384 (1980). *Emend.* Letcher & Powell, *Mycol. Res.* **110**: 907 (2006).

Type: Chytridium A. Braun 1851.

Thallus monocentric or polycentric rhizomycelial; zoospores typically with flagellar base containing an electron-opaque plug, microtubules extending from one side of the kinetosome in a parallel array, ribosomes aggregated near the nucleus, kinetosome parallel to non-flagellated centriole and connected to it by fibrous material, nucleus not associated with kinetosome, fenestrated cisterna (rumposome) adjacent to lipid globule.

Exemplar genera: Chytridium A. Braun 1851, *Chytriomycetes* Karling 1945, *Nowakowskiella* J. Schröt. 1893.

An emended description is presented above to conform to the circumscription adopted here. Monophyly of this group, as currently delimited, is not certain; *Polychytrium* Ajello 1942 and its allies and *Chytriomycetes angularis* Longcore 1992 and its allies may eventually be segregated from *Chytridiales* s. str.

Order: **Rhizophydiales** Letcher, in Letcher *et al.*, *Mycol. Res.* **110**: 908 (2006).

Exemplar genera: Rhizophydium Schenk 1858, *Kappamyces* Letcher & M.J. Powell 2005, *Terramyces* Letcher 2006, *Boothiomyces* Letcher 2006; *Batrachochytrium* Longcore, Pessier & D.K. Nichols 1999 is on a long branch in this clade with no near relatives.

Order: **Spizellomycetales** D. J. S. Barr, *Can. J. Bot.* **58**: 2384 (1980).

Exemplar genera: Spizellomyces D.J.S. Barr 1980, *Powellomyces* Longcore, D.J.S. Barr & Désauln. 1995, *Kochiomyces* D.J.S. Barr 1980.

This classification does not include *Caulochytrium*, *Olpidium*, *Rozella*, or the *Rhizophlyctis rosea* clade, which are considered *incertae sedis*.

Class: **Monoblepharidomycetes** J. H. Schaffn., *Ohio Nat.* **9**: 449 (1909), as "*Monoblepharideae*".

Type: Monoblepharis Cornu 1871.

Thallus filamentous, either extensive or a simple unbranched thallus, often with a basal holdfast; asexual reproduction by zoospores or autospores; zoospores containing a kinetosome parallel to a non-flagellated centriole, a striated disk partially extending around the kinetosome, microtubules radiating anteriorly from the striated disk, a

ribosomal aggregation, and rumposome (fenestrated cisterna) adjacent to a microbody; sexual reproduction oogamous by means of posteriorly uniflagellate antherozoids borne in antheridia and nonflagellate female gametes borne in oogonia.

Schaffner (1909) used the name "*Monoblepharideae*" as a class but with the ending of a suborder; this must be changed without change of authorship or date of publication (*Code*, Art. 16.3).

Order: **Monoblepharidales** J. Schröt., in Engler & Prantl, *Nat. Pflanzenfam.* **1**: 106 (1893), as "*Monoblepharidineae*".

Emend. Sparrow, *Aquatic Phycomycetes*: 458 (1943).

Emended description as for *Monoblepharidomycetes*.

Exemplar genera: *Monoblepharis* Cornu 1871, *Harpochytrium* Lagerh. 1890, *Oedogoniomyces* Tak. Kobay. & M. Ôkubo 1954.

Phylum: **Neocallimastigomycota** M. J. Powell, **phylum nov.**

Mycobank no.: MB 501279

Thallus monocentricus vel polycentricus; fungi anaerobici, intra tractum digestivum animalium herbivororum vel fortasse in substratis anaerobicis terrestribus vel limnicis; mitochondriis carentes sed hydrogenosomatibus praediti; zoosporae retrorsum uni- vel multiflagellatae, kinetosoma praesens sed centriolum supervacaneum absens; complexus kinetosomati affixus e radio marginali et annulo circumflagellari compositus; microtubuli e radio entendentem circum nucleum radiantes et flabellum posterius formantes; munimenta flagelli absentia; tegumentum nuclei mitosi procedente integrum remanens.

Typus: *Neocallimastix* Vavra & Joyon ex I. B. Heath 1983.

Thallus monocentric or polycentric; anaerobic, found in digestive system of larger herbivorous mammals and possibly in other terrestrial and aquatic anaerobic environments; lacks mitochondria but contains hydrogenosomes of mitochondrial origin; zoospores posteriorly unflagellate or polyflagellate, kinetosome present but non-functional centriole absent, kinetosome-associated complex composed of a skirt, strut, spur and circumflagellar ring, microtubules extend from spur and radiate around nucleus, forming a posterior fan, flagellar props absent; nuclear envelope remains intact throughout mitosis.

Class: **Neocallimastigomycetes** M. J. Powell, **class. nov.**

Mycobank no.: MB 501280

Diagnosis latina ut in *Neocallimastigomycota* (vide supra).

Typus: Neocallimastix Vavra & Joyon ex I.B. Heath 1983.

Order: **Neocallimastigales** J. L. Li, I. B. Heath & L. Packer, *Can. J. Bot.* **71**: 403 (1993).

Exemplar genera: Neocallimastix Vavra & Joyon ex I.B. Heath 1983, *Caecomycetes* J.J. Gold 1988, *Orpinomyces* D.J.S. Barr, H. Kudo, Jakober & K.J. Cheng 1989.

Phylum: **Blastocladiomycota** T. Y. James, *Mycologia* **98**: XXX (2007) ["2006"].

Synonym: *Allomycota* Caval.-Sm., *BioSystems* **14**: 465 (1981). This phylum was proposed to reflect phylogenetic information from a number of molecular studies (James *et al.* 2007; Liu *et al.* 2006). Equivalent to

Class: **Blastocladiomycetes** T. Y. James, *Mycologia* **98**: XXX (2007) ["2006"].

Synonym: *Allomyces* Caval.-Sm., *Biol. Rev.* **73**: 246 (1998), based on *Allomyces* E. J. Butler 1911.

Cavalier-Smith provided a brief, five-word Latin description for *Allomyces* that is not diagnostic from other uniflagellate fungi. The name *Allomyces* was not taken up, because it is appropriate to have a class name based on the same genus as an included ordinal name, and because Cavalier-Smith's "diagnosis" was vague.

Order: **Blastocladales** H. E. Petersen, in Kanouse, *Am. J. Bot.* **14**: 295 (1927).

Exemplar genera: *Allomyces* E.J. Butler 1911, *Blastocladia* Reinsch 1877, *Coelomyces* Keilin. 1921.

Phylum: **Microsporidia** Balbiani *C. R. Acad. Sci. Paris* **95**: 1168 (1882).

The nomenclatural status of *Microsporidia* is ambiguous. It has been treated as a phylum under the zoological *Code* (International Commission on Zoological Nomenclature 1999), but there is disagreement about the correct author citation (Larsson 2000; Sprague & Becnel 1998), and it is uncertain if the name would be valid under the botanical *Code*. This uncertainty arises as *Microsporidium* Balbiani 1884 appears to be a later synonym of *Nosema* Naegeli 1857. The present work follows the recommendation of Sprague & Becnel (1998) in attributing *Microsporidia* to Balbiani (1882), but this must be regarded as provisional. Before the status of the *Microsporidia* can be resolved, it will be necessary to decide whether the nomenclature of the group as a whole should be governed by the zoological or the botanical *Code* although the latter now allows names of fungi described under the zoological *Code* to be accepted. The final decision will require input from the community of scientists who study *Microsporidia*.

No subdivision of the group is proposed here, owing to the lack of well-sampled multi-gene phylogenies within the group. However, Vossbrinck & Debrunner-Vossbrinck (2005) proposed a class-level classification of microsporidia, based on small-subunit rRNA gene sequences.

Phylum: **Glomeromycota** C. Walker & A. Schüßler, *in* Schüßler *et al.*, *Mycol. Res.* **105**: 1416 (2001).

Class: **Glomeromycetes** Caval.-Sm., *Biol. Rev.* **73**: 246 (1998), as "*Glomomycetes*".

Synonym: *Geomycetes* Caval.-Sm., *Biol. Rev.* **73**: 247 (1998).

Order: **Archaeosporales** C. Walker & A. Schüßler, *in* Schüßler *et al.*, *Mycol. Res.* **105**: 1418 (2001).

Synonym: *Geosiphonales* Caval.-Sm., *Biol. Rev.* **73**: 247 (1998).

Exemplar genera: *Archaeospora* J. B. Morton & D. Redecker 2001, *Geosiphon* F. Wettst. 1915.

Order: **Diversisporales** C. Walker & A. Schüßler, *Mycol. Res.* **108**: 981 (2004).

Exemplar genera: *Acaulospora* Gerd. & Trappe 1974, *Diversispora* C. Walker & A. Schüssler 2004, *Gigaspora* Gerd. & Trappe 1974, *Pacispora* Oehl & Sieverd. 2004.

Order: **Glomerales** J. B. Morton & Benny, *Mycotaxon* **37**: 473 (1990), as "*Glomales*".

Exemplar genus: *Glomus* Tul. & C. Tul. 1845.

Order: **Paraglomerales** C. Walker & A. Schüßler, in Schüßler *et al.*, *Mycol. Res.* **105**: 1418 (2001).

Exemplar genus: Paraglomerus J.B. Morton & D. Redecker 2001.

Subphyla incertae sedis (not assigned to any phylum):

Subphylum: **Mucoromycotina** Benny, **subphylum nov.**

MycoBank no.: MB 501281

Fungi saprotrophici vel raro mycoparasiti facultativi, gallas facientes, haustoriis carentes, raro ectomycorrhizam facientes. Mycelium ramosum, juvene coenocyticum, maturum aliquando septis microporosis divisum. Reproductio asexualis sporangiis vel sporangiolis vel merosporangiis, raro chlamydosporis vel arthrosporibus vel blastosporibus effecta. Reproductio sexualis zygosporibus plus minusve globosis e suspensoribus oppositis vel appositis formatis effecta.

Typus: Mucor Fresen. 1850.

Fungi saprobes, or rarely gall-forming, nonhaustorial, facultative mycoparasites, or forming ectomycorrhiza. Mycelium branched, coenocytic when young, sometimes producing septa that contain micropores at maturity. Asexual reproduction by sporangia, sporangiola, or merosporangia, or rarely by chlamydospores, arthrospores, or blastospores. Sexual reproduction by more or less globose zygosporidia formed on opposed or apposed suspensors.

This group includes the *Mucorales*, which is the core group of the traditional *Zygomycota*. Monophyly of the traditional *Zygomycota* (including *Mucorales*, *Glomerales*, *Entomophthorales* and *Harpellales*) was suggested by a recent study by Liu *et al.* (2006) using *rpb1* and *rpb2*, but that finding conflicts with results of analyses that included

additional loci and taxa, which suggested that the traditional *Zygomycota* is polyphyletic (James *et al.* 2006).

The name *Zygomycota* was first published without a Latin diagnosis by Moreau (1954) and is therefore invalid. At present, this classification does not include *Zygomycota*. When relationships among basal fungal lineages are more clearly resolved, it may be appropriate to resurrect and validate *Zygomycota*, to include *Mucoromycotina* and perhaps other clades.

Order: **Mucorales** Fr., *Syst. Mycol.* **3** (2): 296 (1832).

Exemplar genera: *Mucor* Fresen. 1850 (*pro parte*), *Parasitella* Bainier 1903, *Phycomyces* Kunze 1823, *Pilobolus* Tode 1784, *Rhizopus* Ehrenb. 1821.

Order: **Endogonales** F. Moreau ex R. K. Benj., *in* Kendrick (ed.), *Whole Fungus* **2**: 599 (1979).

Emend.: Morton & Benny, *Mycotaxon* **37**: 473 (1990)

Synonym: *Endogonales* F. Moreau, *Encycl. Mycol.* **23**: 1231 (1953), *nomen nudum*.

Exemplar genera: *Endogone* Link 1809, *Peridiospora* C.G. Wu & S.J. Lin 1997, *Sclerogone* Warcup 1990, *Youngiomyces* Y.J. Yao 1995.

Order: **Mortierellales** Caval.-Sm., *Biol. Rev.* **73**: 246 (1998).

Exemplar genera: *Mortierella* Coem. 1863, *Dissophora* Thaxt. 1914, *Modicella* Kanouse 1936.

Subphylum: **Entomophthoromycotina** Humber, **subphylum nov.**

MycoBank no.: MB 501282

Fungi pathogenici obligate animalibus (praecipue invertebratis) vel plantis cryptogamicis vel saprotrophici, interdum in animalibus vertebratis parasitici. Status somaticus mycelium coenocyticum vel septatum, pariete circumdatum vel protoplasticum, in hospite culturisve saepe corpora hyphalia multinucleata formans; forma protoplastica hyphoidea vel amoeboida forma variabilis; cystidia et rhizoidea in aliquot speciebus arthropodicolis formata. Characteres nuclei, sicut magnitudo, nucleoli magnitudo et locus, praesentia aut absentia heterochromatini intermitotici, familiis distinguendis iuvant. Conidiophora simplicia ramosave. Sporae primariae conidia vera, uninucleatae vel plurinucleatae vel multinucleatae, variis modis vi propulsae vel passively liberatae, conidia secundaria persaepe formata. Sporae perdurantes crassitunicatae, bistratosae velut zygosporae post conjugationem velut azygosporae singulae formatae.

Typus: Entomophthora Fresen. 1856.

Obligate pathogens of animals (primarily arthropods), cryptogamic plants, or saprobes; occasionally facultative parasites of vertebrates. Somatic state consisting of a well-defined mycelium, coenocytic or septate, walled or protoplasmic, which may fragment to form multinucleate hyphal bodies; protoplasts either hyphoid or amoeboid and changeable in shape; cystidia or rhizoids formed by some taxa. Such nuclear characters as overall size, location and comparative size of nucleoli, presence or absence of granular heterochromatin in chemically unfixed interphasic nuclei, and mitotic patterns are important at the family level. Conidiophores branched or unbranched. Primary spores true conidia, uni-, pleuri-, or multinucleate, forcibly discharged by diverse possible means or passively dispersed; secondary conidia often produced. Resting spores with thick bi-layered walls form as zygosporae after conjugations of undifferentiated gametangia from different or the same hyphal bodies or hypha or as azygosporae arising without prior gametangial conjugations.

Order: **Entomophthorales** G. Winter, *Rabenh. Krypt.-Fl.* **1**(1): 74 (1880).

Exemplar genera: Entomophthora Fresen. 1856, *Ballocephala* Drechsler 1951, *Conidiobolus* Bref. 1884, *Entomophaga* Batko 1964, *Neozygites* Witlaczil 1885.

Subphylum: **Zoopagomycotina** Benny, **subphylum nov.**

Mycobank no.: MB 501283

Fungi endo- vel ectoparasitici microanimalium vel fungorum. Corpus vegetativum ex thallo simplici ramoso vel nonramoso vel mycelio nonseptato plus minusve extense ramoso constans. Ectoparasitae haustoria intra hospitem formantes. Reproductio asexualis arthrosporis, chlamydosporis vel sporangiolis uni- vel multisporis perfecta; sporangiosporae sporangiolorum multisporum in catenenis (merosporangiis) simplicibus vel ramosis dispositae. Reproductio sexualis zygosporis paene globosis perfacitur; hyphae sexuales hyphis vegetativis similes vel plus minusve ampliatae.

Typus: Zoopage Drechsler 1935.

Endo- or ectoparasites of microanimals and fungi. Vegetative body consisting of a simple, branched or unbranched thallus or more or less extensively branched mycelium. Ectoparasites forming haustoria inside the host. Asexual reproduction by arthrospores, chlamydospores or uni- or multispored sporangiola; sporangiospores of multispored sporangiola formed in simple or branched chains (merosporangia). Sexual reproduction by nearly globose zygosporis; sexual hyphae similar to the vegetative hyphae or more or less enlarged.

The description of this group is based mostly on the validating description for the *Zoopagales* by Benjamin (1979), except that arthrospores have been added, based on Barron's (1975) report of arthrospores in *Helicocephalum* Thaxt. 1891.

Order: **Zoopagales** Bessey ex R. K. Benj., in Kendrick (ed.), *Whole Fungus* **2**: 590 (1979).

Synonym: *Zoopagales* Bessey, *Morph. Tax, Fungi* : 177 (1950), *nomen nudum*.

Exemplar genera: Cochlonema Drechsler 1935, *Rhopalomyces* Corda 1839, *Piptocephalis* de Bary 1865, *Sigmoideomyces* Thaxt. 1891, *Syncephalis* Tiegh. & G. Le Monn. 1873, *Zoopage* Drechsler 1935.

Subphylum: **Kickxellomycotina** Benny, **subphylum nov.**

MycoBank no.: MB 501284

Fungi saprotrophici vel mycoparasitici vel obligate symbiotici. Thallus in nonnullis generibus e tenaculo fungos alios parasitans et haustoriis penetrans; mycelium septatum, ramosum vel simplex; septa in medio excavata et obturata. Reproductio asexualis merosporangiis uni- vel bisporis vel trichosporis vel arthrosporis effecta. Reproductio sexualis zygosporis globosis, biconicis vel allantoideis circinatis effecta.

Typus: Kickxella Coem. 1862.

Fungi saprobes, mycoparasites, or obligate symbionts. Thallus arising from a holdfast on other fungi as a haustorial parasite, or branched, septate, subaerial hyphae.

Mycelium branched or unbranched, regularly septate. Septa with median, disciform cavities containing plugs. Asexual production by 1- or 2-spored merosporangia, trichospores, or arthrospores. Sexual reproduction by zygosporis that are globose, biconical, or allantoid and coiled.

Order: **Kickxellales** Kreisel ex R. K. Benj., in Kendrick (ed.), *Whole Fungus* 2: 610 (1979).

Synonym: *Kickxellales* Kreisel, *Grundz. nat. Syst. Pilze*: 65 (1969), *nomen nudum*.

Exemplar genera: Kickxella Coem. 1862, *Coemansia* Tiegh. & G. Le Monn. 1873, *Linderina* Raper & Fennell 1952, *Spirodactylon* R.K. Benj. 1959.

Order: **Dimargaritales** R. K. Benj., in Kendrick (ed.), *Whole Fungus 2*: 607 (1979).

Exemplar genera: Dimargaris Tiegh. 1875, *Dispira* Tiegh. 1875, *Tieghemiomyces* R.K. Benj. 1959.

Order: **Harpellales** Lichtw. & Manier, *Mycotaxon 7*: 441 (1978).

The taxa in this order have been referred to as "*Trichomyces*" However, *Trichomyces* is no longer a useful phylogenetic taxon because it describes a polyphyletic group. The use of the term should be restricted to ecological rather than phylogenetic groupings, and not capitalized or italicized, i.e. as "trichomyces".

Exemplar genera: Harpella L. Léger & Duboscq 1929, *Furculomyces* Lichtw. & M. C. Williams 1992, *Legeriomyces* Pouzar 1972, *Smittium* R. Poiss. 1937.

Order: **Asellariales** Manier ex Manier & Lichtw., *Mycotaxon 7*: 442 (1978).

Exemplar genera: Asellaria R. Poiss. 1932, *Orchesellaria* Manier ex Manier & Lichtw. 1968.

Asellariales are retained in the *Fungi* here due to their ultrastructural characteristics (Benny & White 2001; Manier 1973; Moss 1975; Saikawa *et al.* 1997). Unpublished *rpb1* and *rpb2* data also support their placement in the *Kickxellomycotina* (T. Y. James & M. M. White, unpubl.).

Subkingdom: **Dikarya** Hibbett, T. Y. James & Vilgalys, **subregnum nov.**

MycoBank no.: MB 501285

Synonym: *Neomycota* Cavl.-Sm., *Rev. Biol.* **73**: 209 (1998).

Fungi unicellulares vel filamentosi, flagellis carentes, saepe stadium dikaryoticum includentes.

Ascomycota et *Basidiomycota* complectens.

Unicellular or filamentous *Fungi*, lacking flagella, often with a dikaryotic state. The least-inclusive clade that contains *Ascomycota* and *Basidiomycota*.

The name alludes to the putative synapomorphy of dikaryotic hyphae (Tehler 1988) and was applied by James *et al.* (2006) without formal description. Kendrick (1985) and Tehler *et al.* (2003) referred to this group as the *Dikaryomycota*, but the termination "-mycota" denotes the rank of phylum under the *Code*. Cavalier-Smith (1998) referred to this group as *Neomycota*. *Dikarya* is used here, because it is more descriptive and is consistent with recent use (James *et al.* 2006; Tehler *et al.* 2003; Kendrick 1985).

Phylum: **Ascomycota** Bold ex Caval.-Sm., *Biol. Rev.* 73: 247 (1998), as "*Ascomycota* Berk. 1857. stat. nov."

Synonyms: *Ascomycetes* Berk., *Intr. Crypt. Bot.*: 270 (1857), rank uncertain; Whittaker (1959: 220).

Ascomycota Bold, *Morph. Pl.*: 7, 180 (1958), *nomen nudum*; Hawksworth *et al.* (1995: 30), Eriksson & Winka (1997: 4), etc, *nomina nuda*.

Basic type: Peziza Fr. 1822.

Cavalier-Smith was not the first to propose the phylum name *Ascomycota*. It seems it was first used by Bold (1958: 7, 180), but without a Latin diagnosis. The name was in widespread use by that time, and its usage was popularized by its employment in the eighth edition of the *Dictionary*, which is listed in Cavalier-Smith's (1998) bibliography. As in the case of the name *Fungi*, the first author to use phylum status is recognized in this author citation. The Latin diagnosis provided by Cavalier-Smith consisted of only two words: "sporae intracellulares". It is questionable whether this description is

diagnostic for only the ascomycetes, but as a validating diagnosis it is acceptable under the *Code*. No detailed reference to the basionym was given, but is provided here. We also propose a basic type, *Peziza* as we can not be sure that the phylum will not be split in the future when more molecular data and material of ascomycetes and basidiomycetes have been sequenced. Hawksworth *et al.* (1995) and Eriksson & Winka (1997: 4) used the phylum names *Ascomycota* and *Basidiomycota*; the latter authors listed 31 nucleotide signatures in the nSSU rDNA genes in *Basidiomycota*. Since then many more sequences have become available, also from many other genes that support monophyly of *Ascomycota* and *Basidiomycota*.

The subdivision of *Ascomycota* used in the present paper is based on the system of Eriksson & Winka (1997), which differs in many respects from that of Cavalier-Smith (1998).

Subphylum: **Taphrinomycotina** O. E. Erikss. & Winka, *Myconet* **1**: 11 (1997).

Class: **Taphrinomycetes** O. E. Erikss. & Winka, *Myconet* **1**: 11 (1997).

Order: **Taphrinales** Gäum. & C. W. Dodge, *Comp. morph. fun.*: 159 (1928).

Exemplar genera: Taphrina Fr. 1815, *Protomyces* Unger 1832.

Class: **Neoelectomycetes** O. E. Erikss. & Winka, *Myconet* **1**: 8 (1997).

Order: **Neoelectales** Landvik, O. E. Erikss., Gargas & P. Gustafss., *Syst. Ascom.* **11**: 114 (1993).

Exemplar genus: Neoelecta Speg. 1881.

Class: **Pneumocystidomycetes** O. E. Erikss. & Winka, *Myconet* **1**: 9 (1997).

Order: **Pneumocystidales** O. E. Erikss., *Syst. Ascom.* **13**: 170 (1994).

Exemplar genus: Pneumocystis P. Delanoë & Delanoë 1912.

Class: **Schizosaccharomycetes** O. E. Erikss. & Winka, *Myconet* **1**: 10 (1997).

Order: **Schizosaccharomycetales** O. E. Erikss., Svedskog & Landvik, *Syst. Ascom.* **11**: 146 (1993).

Exemplar genus: Schizosaccharomyces Linder 1893.

Subphylum: **Saccharomycotina** O. E. Erikss. & Winka, *Myconet* **1**: 10 (1997).

Class: **Saccharomycetes** O. E. Erikss. & Winka, *Myconet* **1**: 10 (1997).

Order: **Saccharomycetales** Kudrjawzew, *System Hefen*: 270 (1960).

Growth usually by individual yeast cells, often accompanied by pseudohyphae and/or true hyphae. Cell walls predominately of β -glucan. Ascospores formed in asci that often are converted from individual cells or borne on simple ascophores. Mitotic and meiotic nuclear divisions within an intact nuclear membrane. Enveloping membrane system in ascospore delimitation associated independently with postmeiotic nuclei. Asexual reproduction by holoblastic budding, conidia or fission (arthrospores).

Exemplar genera: Saccharomyces Meyen ex E.C. Hansen 1838, *Candida* Berkhout 1923, *Dipodascopsis* L.R. Batra & Millner 1978, *Metschnikowia* T. Kamienski 1899.

Subphylum: **Pezizomycotina** O. E. Erikss. & Winka, *Myconet* **1**: 9 (1997).

Class: **Arthoniomycetes** O. E. Erikss. & Winka, *Myconet* **1**: 4 (1997).

Order: **Arthoniales** Henssen & Jahns ex D. Hawksw. & O. E. Erikss, *Syst. Ascom.* **5**: 177 (1986).

Synonym: *Arthoniales* Henssen & Jahns, *Lichenes*: 123 (1973) ["1974"], *nomen nudum*.

Hawkworth & Eriksson (*loc. cit.*) listed only Henssen, but cited the book by Henssen & Jahns (*loc. cit.*) as place for the original but invalid description so both should be cited although Henssen contributed the taxonomic system to the book.

Exemplar genera: Arthonia Ach. 1806, *Chrysothrix* Mont. 1852, *Dirina* Fr. 1825, *Roccella* DC 1805.

Class: **Dothideomycetes** O. E. Erikss. & Winka, *Myconet* **1**: 5 (1997).

Subclass: **Dothideomycetidae** P. M. Kirk, P. F. Cannon, J. C. David & Stalpers ex Schoch *et al.*, *Mycologia* **98**: XXX (2007) ["2006"].

Order: **Capnodiales** Woron., *Annl's Mycol.* **23**: 177 (1925).

Exemplar genera: Capnodium Mont. 1848, *Scorias* Fr. 1825, *Mycosphaerella* Johanson 1884.

Order: **Dothideales** Lindau, in Engler & Prantl (eds), *Nat. Pflanzenfam.* 1(1): 373 (1897).

Exemplar genera: *Dothidea* Fr. 1818, *Dothiora* Fr. 1849, *Sydowia* Bres. 1895, *Stylodothis* Arx & E. Müll. 1975.

Order: **Myriangiales** Starbäck, *K. svenska Vetensk-Akad. Handl., Bih., Afd. III* 25(1): 37 (1899).

Exemplar genera: *Myriangium* Mont. & Berk. 1845, *Elsinoë* Racib. 1900.

Subclass: **Pleosporomycetidae** C. L. Schoch, Spatafora, Crous & Shoemaker, *Mycologia* 98: XXX (2007) ["2006"].

Order: **Pleosporales** Luttr. ex M. E. Barr, *Prodr. Class Loculoasc.:* 67 (1987).

Synonym: *Pleosporales* Luttr., *Mycologia* 47: 520 (1955), *nomen nudum*.

Exemplar genera: *Pleospora* Rabenh. ex Ces. & De Not. 1863, *Phaeosphaeria* I. Miyake 1909, *Lophiostoma* Ces. & De Not. 1863, *Sporormiella* Ellis & Everh. 1892, *Montagnula* Berl. 1896.

Dothideomycetes *incertae sedis* (not placed in any subclass)

Order: **Botryosphaeriales** C. L. Schoch, Crous & Shoemaker, *Mycologia* 98: XXX (2007) ["2006"].

Exemplar genera: *Botryosphaeria* Ces. & De Not. 1863, *Guignardia* Viala & Ravaz 1892.

Order: **Hysteriales** Lindau in Engler & Prantl (eds), *Nat. Pflanzenfam.* **1**(1): 265 (1896), as "Hysteriinae".

Exemplar genera: Hysterium Pers. 1797, *Hysteropatella* Rehm. 1890.

Order: **Patellariales** D. Hawksw. & O. E. Erikss., *Syst. Ascom.* **5**: 181 (1986).

Exemplar genus: Patellaria Fr. 1822.

Order: **Jahnulales** Ka-Lai Pang, Abdel-Wahab, El-Shar., E. B. G. Jones & Sivichai, in Pang *et al.*, *Mycol. Res.* **106**: 1033 (2002).

Exemplar genera: Aliquandostipite Inderb. 2001, *Jahnula* Kirschst. 1936, *Patescospora* Abdel-Wahab & El-Shar. 2002.

Class: **Eurotiomycetes** O. E. Erikss. & Winka, *Myconet* **1**: 6 (1997).

The circumscription of this class and the classification within the *Eurotiomycetes* presented here are derived from the phylogenetic re-delimitation of this class by Ekman and Tønsberg (2002), Lutzoni *et al.* (2004) and Geiser *et al.* (2007), reflecting the inference of shared ancestry between *Eurotiomycetes*, comprising *Coryneliales*, *Onygenales* and *Eurotiales* and *Chaetothyriomycetes*. Three subclasses, *Chaetothyriomycetidae*, *Eurotiomycetidae*, and *Mycocaliciomycetidae*, are defined to represent the major lineages within *Eurotiomycetes*.

Subclass: **Chaetothyriomycetidae** Lutzoni, Gueidan, Unter. & Geiser, **stat. nov.**

Mycobank no.: MB 501286

Basionym: *Chaetothyriomycetes* O. E. Erikss. & K. Winka. *Syst. Ascom.* **1**: 5. 1997.

Lichenized, parasitic, and saprobic ascomycetes with mostly bitunicate / fissitunicate to evanescent asci, produced in perithecial ascomata arranged superficially or immersed in a thallus. Thalli often produced on the surfaces of rocks, lichens, decaying plant material and other substrata. Ascospores variable, from colourless to pigmented, simple to muriform. Hamathecium, when present, consisting of pseudoparaphyses. Pigments, when present, generally related to melanin. Asexual stages with phialidic and annellidic anamorphs observed in non-lichenized taxa.

Order: **Chaetothyriales** M. E. Barr, *Mycotaxon* **29**: 502 (1987).

Exemplar genera: *Capronia* Sacc. 1883, *Ceramothyrium* Bat. & H. Maia 1956, *Chaetothyrium* Speg. 1888.

Order: **Pyrenulales** Fink ex D. Hawksw. & O. E. Erikss., *Syst. Ascom.* **5**: 182 (1986).

Synonym: *Pyrenulales* Fink, *Ohio St. Univ. Bull.* **19**(28): 107 (1951), *nomen nudum*.

Exemplar genera: *Pyrenula* Ach. 1814, *Pyrgillus* Nyl. 1858.

Order: **Verrucariales** Mattick ex D. Hawksw. & O. E. Erikss., *Syst. Ascom.* **5**: 183 (1986).

Synonym: *Verrucariales* Mattick, in Engler, *Syll. Pflanzenfam.* (12 edn): 208 (1954), *nomen nudum*.

Exemplar genera: *Agonimia* Zahlbr. 1909, *Dermatocarpon* Eschw. 1824, *Polyblastia* A. Massal. 1852, *Verrucaria* Schrad. 1794.

Subclass: **Eurotiomycetidae** Geiser & Lutzoni, **subclass. nov.**

Mycobank no.: MB 501287

Fungi saprotrophici vel parasitici vel mycorrhizales; asci globosi in toto ascomate sparsi, raro hymenium formantes; asci plerumque evanescentes, nonnumquam bitunicati. Ascosporae plerumque unicellulares, lenticulares, nonnumquam globosae vel ellipsoideae. Ascomata, si formata, plerumque cleistothecialia vel gymnothecialia, saepe textura stromatica circumdata. Structurae hamatheciales absentes. Gametangia plerumque indistincta e glomere hyphali constantia. Fungi saepe laete colorati. Anamorphae variabiles, seu phialidicae seu arthroconidiales.

Typus: Eurotium Link 1809.

Saprotrophic, parasitic and mycorrhizal. Ascomata, when present, usually cleistothecial/gymnothecial, globose, often produced in surrounding stromatic tissue and brightly coloured; hamathecial elements lacking; gametangia usually undifferentiated and consisting of hyphal coils. Asci usually evanescent, sometimes bitunicate, scattered throughout the ascoma, rarely from a hymenium. Ascospores usually single-celled, lenticular, sometimes spherical or elliptical. Anamorphs variable, including phialidic and arthroconidial forms.

This name was employed by Lutzoni *et al.* (2004) and Geiser *et al.* (2007), in the same sense as the present classification, but without a formal diagnosis.

Order: **Coryneliales** Seaver & Chardón, *Scient. Surv. P. Rico* **8**(1): 40 (1926).

Exemplar genera: Corynelia Ach. 1823, *Caliciopsis* Peck 1880.

Order: **Eurotiales** G. W. Martin ex Benny & Kimbr., *Mycotaxon* **12**: 23 (1980).

Synonym: *Eurotiales* G. W. Martin, *Std. nat. Hist. Iowa Univ.* **18**(Suppl.): 16 (1941), *nomen nudum*.

Exemplar genera: Eurotium Link 1809, *Emericella* Berk. 1857, *Talaromyces* C. R. Benj. 1955, *Elaphomyces* Nees 1820, *Trichocoma* Jungh. 1838, *Byssochlamys* Westling 1909.

Order: **Onygenales** Cif. ex Benny & Kimbr., *Mycotaxon* **12**: 8 (1980).

Synonym: *Onygenales* Cif., *Atti Ist. Bot. Univ. Pavia, ser. 5*, **14**: 238 (1957), *nomen nudum*.

Emend. Currah *Mycotaxon* **24**: 13 (1985).

Exemplar genera: Onygena Pers. 1799, *Gymnoascus* Baran. 1872, *Arthroderma* Curr. 1860.

Subclass: **Mycocaliciomycetidae** Tibell. **subclass nov.**

Mycobank no.: MB 501288

Parasitae vel commensales in lichenibus vel saprotrophici. Ascomata disciformia, stipitata vel sessilia. Excipulum cupulatum, saltem partim scleroticum hyphis stipitis simile. Dispersio sporarum activa, raro passiva et tum mazedio parce evoluta. Asci unitunicati, cylindrici, vulgo apice distincte incrassato, 8-sporei. Ascosporae pallidae ad atrofuscae, ellipsoidales, non-septatae vel transversaliter 1-7-septatae. Parietis sporae atrofusca, laevis vel ornamento intra plasmalemma formato. Derivata acidi vulpinici in speciebus paucis praesentia. Anamorphae coelomycetum et hyphomycetum variae praesentes.

Typus: Mycocalicium Vain. 1890.

Parasites or commensals on lichens or saprobes. Ascomata disciform, stalked or sessile. Excipulum cupulate, and like the stalk hyphae at least in part sclerotized. Spore dispersal active, more rarely passive and ascomata then with a moderately developed mazaedium. Asci unitunicate, cylindrical, mostly with a distinctly thickened apex, 8-spored. Ascospores pale to blackish brown, ellipsoidal or spherical to cuboid, non-septate or transversely 1-7-septate. Spore wall pigmented, smooth or with an ornamentation formed within the plasmalemma. Vulpinic acid derivatives occur in a few species. A variety of coelomycetous and hyphomycetous anamorphs occur.

Order: **Mycocaliciales** Tibell & Wedin, *Mycologia* **92**: 579 (2000).

Exemplar genera: *Mycocalicium* Vain. 1890, *Chaenothecopsis* Vain. 1927, *Stenocybe* (Nyl.) Körb. 1855, *Sphinctrina* Fr. 1825.

Class: **Laboulbeniomyces** Engl., *Syll. Pflanzenfam.* (2nd edn): 46 (1898).

Order: **Laboulbeniales** Lindau, in Engler & Prantl (eds), *Nat. Pflanzenfam.* **1**(1): 491 (1897), as "*Laboulbeniineae*".

Exemplar genera: *Laboulbenia* Mont. & C.P. Robin 1835, *Rickia* Cavara 1899, *Ceratomyces* Thaxt. 1892.

Order: **Pyxidiophorales** P. F. Cannon, in Kirk *et al.*, *Ainsworth & Bisby's Dict. Fungi* (9th edn): xi (2001).

Exemplar genus: *Pyxidiophora* Bref. & Tavel 1891.

Class: **Lecanoromycetes** O. E. Erikss. & Winka, *Myconet* **1**: 7 (1997).

Subclass: **Acarosporomycetidae** Reeb, Lutzoni & Cl. Roux, *Mol. Phylogen. Evol.* **32**: 1053 (2004).

Order: **Acarosporales** Reeb, Lutzoni & Cl. Roux, **ord. nov.**

Mycobank no.: MB 501289

Ascomycetes lichenisati algas virides thallo continentes. Ascomata immersa vel sessilia, disciformia vel perithecioidea. Excipulum hyalinum, annulatum. Hymenium non-amyloideum. Paraphyses mediocriter

vel infirme ramosae, septatae, mediocriter vel infirme anastomosantes. Asci unitunicati, non-amyloidei vel satis infirme amyloidei, polyspori. Ascosporae hyalinae, non-septatae, non-halonatae.

Typus: Acarospora A. Massal. 1852.

Lichen-forming ascomycetes with chlorococcoid photobiont. Ascomata immersed or sessile, disciform or perithecioid. True exciple hyaline, annulate. Hymenium non-amyloid. Paraphyses moderately to poorly branched, septate, moderately to poorly anastomosing. Asci functionally unitunicate, lecanoralean, non-amyloid or with slightly amyloid tholi, polyspored, generally with more than 100 ascospores per ascus. Ascospores hyaline, small, non-septate, non-halonate.

The members of this order were formerly classified within the *Lecanorales*, but Reeb *et al.* (2004) and Lutzoni *et al.* (2004) demonstrated that the *Acarosporaceae* diverged earlier than the *Lecanoromycetidae* and *Ostropomycetidae*. This early divergence within the *Lecanoromycetes* was confirmed by Wedin *et al.* (2005) and Miądlikowska *et al.* (2006).

Exemplar genera: Acarospora A. Massal. 1852, *Pleopsidium* Körb. 1855, *Sarcogyne* Flot 1851.

Subclass: **Lecanoromycetidae** P. M. Kirk, P. F. Cannon, J. C. David & Stalpers ex Miądl., Lutzoni & Lumbsch, **subclass. nov.**

MycoBank no.: MB 501290

Synonym: *Lecanoromycetidae* P. M. Kirk, P. F. Cannon, J. C. David & Stalpers, *Ainsworth & Bisby's Dict. Fungi* (9th edn): xi (2001), *nomen nudum*..

Ascomycetes lichenisati algas virides vel cyanobacteria thallo continentis. Ascomata immersa, sessilia vel elevata, generaliter disciformia. Excipulum hyalinum vel pigmentatum, annulatum vel cupulatum.

Hymenium amyloideum vel non-amyloideum. Paraphyses simplices vel ramosae, septatae,

anastomosantes vel non-anastomosantes. Asci bitunicati, unitunicati vel prototunicati, non-amyloidei vel amyloidei, generaliter octospori, sed etiam 1- ad multispori. Ascosporae hyalinae vel brunneae, non-septatae, vel septatae usque ad muriformes, halonatae vel non-halonatae.

Typus: Lecanora Ach. 1809.

Lichen-forming ascomycetes with green algal or cyanobacterial photobiont. Ascomata immersed, sessile or stalked, usually disciform. True exciple hyaline or pigmented, annulate or cupulate. Hymenium amyloid or non-amyloid. Paraphyses simple or moderately to richly branched, septate, anastomosing or not. Asci bitunicate, functionally unitunicate, or prototunicate, lecanoralean, non-amyloid or amyloid, mostly 8-spored, but varying from 1- to poly-spored. Ascospores hyaline or brown, non-septate, trans-septate or muriform, halonate or non-halonate.

This subclass includes the bulk of lichenized discomycetes and corresponds to the phylogenetic circumscription of this subclass by Reeb *et al.* (2004), Lutzoni *et al.* (2004) and Miadlikowska *et al.* (2006). It is in agreement with the *Lecanorales* of Lumbsch *et al.* (2004) and Wiklund & Wedin (2003). The orders *Peltigerales* and *Teloschistales* are here accepted at the ordinal level, following Miadlikowska & Lutzoni (2004) and Miadlikowska *et al.* (2007).

Order: **Lecanorales** Nannf., *Nova Acta R. Soc. Scient. Upsal.* ser. 4 **8**(2): 68 (1932).

Exemplar genera: Cladonia Hill. ex P. Browne 1756, *Lecanora* Ach. 1809, *Parmelia* Ach. 1803, *Ramalina* Ach. 1809, *Usnea* Dill. ex Adans. 1763

Order: **Peltigerales** Walt. Watson, *New Phytologist* **28**: 9 (1929).

Exemplar genera: Coccocarpia Pers. 1827, *Collema* F. H. Wigg. 1780, *Nephroma* Ach. 1810, *Pannaria* Del. ex Bory 1828, *Peltigera* Willd. 1787.

Order: **Teloschistales** D. Hawksw. & O. E. Erikss., *Syst. Ascom.* 5: 183 (1986).

Exemplar genera: Caloplaca Th. Fr. 1861, *Teloschistes* Norman 1853, *Xanthoria* (Fr.) Th. Fr. 1860.

Subclass: **Ostropomycetidae** Reeb, Lutzoni & Cl. Roux, *Mol. Phylogen. Evol.* 32: 1055 (2004).

Order: **Agyriales** Clem. & Shear, *Gen. Fungi*: 141 (1931).

Exemplar genera: Agyrium Fr. 1822, *Placopsis* (Nyl.) Linds. 1867, *Trapelia* M. Choisy 1929, *Trapeliopsis* Hertel & Gotth. Schneid. 1980.

Order: **Baeomycetales** Lumbsch, Huhndorf & Lutzoni, **ord. nov.**

Mycobank no.: MB 501291

Ascomycetes lichenisati algas virides thallo continentes. Ascomata elevata vel raro sessilia, disciformia.

Excipulum hyalinum vel pigmentatum, annulatum vel cupulatum. Hymenium non-amyloideum.

Paraphyses ramosae, septatae. Asci unitunicati, non-amyloidei vel satis infirme amyloidei, octospori.

Ascospores hyalinae, non-septatae vel septatae, halonatae vel non-halonatae.

Typus: Baeomyces Pers. 1794.

Lichen-forming ascomycetes with chlorococcoid photobiont. Ascomata sessile or rarely stalked, disciform. True exciple hyaline or pigmented, annulate or cupulate. Hymenium non-amyloid. Paraphyses moderately to richly branched, septate. Asci unitunicate, non-

amyloid or with slightly amyloid tholi, 8-spored. Ascospores hyaline, non-septate or trans-septate, halonate or non-halonate.

Baeomycetales was shown to differ from *Agyriales* by Kauff and Lutzoni (2002) and this was confirmed by Miądlikowska *et al.* (2006) and Lumbsch *et al.* (2007).

Exemplar genera: *Ainoa* Lumbsch & I. Schmitt 2001, *Baeomyces* Pers. 1794, *Phyllobaeis* Gierl & Kalb 1993.

Order: **Ostropales** Nannf., *Nova Acta R. Soc. Scient. Upsal.*, ser. 4 **8**(2): 68 (1932).

This order includes also taxa formerly classified in separate orders, such as *Gomphillales*, *Graphidales*, *Gyalectales* and *Trichotheliales*.

Exemplar genera: *Ostropa* Fr. 1825, *Stictis* Pers. 1799, *Gyalecta* Ach. 1808, *Gomphillus* Nyl. 1855, *Graphis* Adans 1763., *Odontotrema* Nyl. 1858, *Porina* Müll. Arg. 1883, *Thelotrema* Ach. 1803.

Order: **Pertusariales** M. Choisy ex D. Hawksw. & O. E. Erikss., *Syst. Ascom.* **5**: 181 (1986).

Synonym: *Pertusariales* M. Choisy, *Bull. mens. Soc. linn. Lyon* **18**: 12 (1949), *nomen nudum*.

This order may not be monophyletic as currently circumscribed, with *Ochrolechiaceae* and some groups of the heterogeneous *Pertusaria* clustering in a separate clade, but without support. Nonetheless, a cluster of taxa in a 'core' group of *Pertusariales* has been strongly supported as monophyletic in phylogenetic analyses by Miądlikowska *et al.* (2006), Lücking *et al.* (2004), Schmitt *et al.* (2005), Lutzoni *et al.* (2004), and Grube *et al.* (2004).

Exemplar genera: *Coccotrema* Müll. Arg. 1888, *Icmadophila* Trevis. 1853, *Ochrolechia* A. Massal. 1852, *Pertusaria* DC 1805.

Lecanoromycetes *incertae sedis* (not placed in any subclass):

Order: **Candelariales** Miadl., Lutzoni & Lumbsch, **ord. nov.**

MycoBank no.: MB 501292

Ascomycetes lichenisati algas virides thallo continentes. Ascomata sessilia, disciformia. Excipulum hyalinum, annulatum. Hymenium amyloideum. Paraphyses ramosae, septatae. Asci unitunicati, amyloidei, ad typum *Candelariae* dictum pertinentes, octo- vel saepe multispori. Ascosporae hyalinae, non-septatae vel raro 1-septatae.

Typus: Candelaria A. Massal. 1853.

Lichen-forming ascomycetes with chlorococcoid photobiont, predominantly nitrophilous. Thallus of various morphology, yellow to orange (pulvinic acid derivatives). Ascomata apothecial, sessile, with or without a distinct margin, yellow to orange. The ascomatal wall formed from densely septate twisted hyphae. paraphyses mostly simple. Excipulum hyaline, hymenium amyloid. Asci unitunicate of *Candelaria*-type with the amyloid lower part of the apical dome and broad apical cushion, often multispored. Ascospores hyaline, aseptate, rarely 1-septate.

Candelariales was shown to differ from *Lecanorales* by Wedin *et al.* (2005) and this was confirmed by Hofstetter *et al.* (2007) and Miadlikowska *et al.* (2007).

Exemplar genera: Candelaria A. Massal. 1853, *Candelariella* Müll. Arg. 1894.

Order: **Umbilicariales** Lumbsch, Hestmark & Lutzoni, **ord. nov.**

MycoBank no.: MB 501293

Ascomycetes lichenisati algas virides thallo continentes. Ascomata sessilia, raro immersa usque ad paucē elevata, plerumque atra, irregularia, disciformia. Excipulum pigmentatum, annulatum. Hymenium amyloideum. Paraphyses simplices vel paulum ramosae, septatae. Asci unitunicati, tholo inconspicue amyloideo, 1-8-spori. Ascosporae hyalinae vel brunneae, non-septatae usque ad muriformes.

Typus: Umbilicaria Hoffm. 1789.

Lichen-forming ascomycetes with chlorococcoid photobiont. Ascomata sessile, or rarely immersed or stalked, mostly black, irregular, disciform. True exciple pigmented, annulate. Hymenium amyloid. Paraphyses simple or slightly branched, septate, apically thickened. Asci unitunicate, with slightly amyloid tholi, 1-8-spored. Ascospores hyaline or brown, non-septate to muriform.

Exemplar genera: Lasallia Mérat 1821, *Umbilicaria* Hoffm. 1789.

Class: **Leotiomyces** O. E. Erikss. & Winka, *Myconet* 1:7 (1997).

Excluding *Geoglossaceae* (Wang *et al.* 2006).

Order: **Cyttariales** Luttr. ex Gamundí, *Darwiniana* 16: 502 (1971).

Synonym: *Cyttariales* Luttr., *Univ. Miss. Stud.* 24(2): 109 (1951), *nomen nudum*.

Exemplar genus: Cyttaria Berk. 1842.

Order: **Erysiphales** H. Gwynne-Vaughan, *Fungi, Ascom., Ustilag., Ured.*: 78 (1922).

Exemplar genera: Erysiphe R. Hedw. ex DC 1805, *Blumeria* Golovin ex Speer 1975, *Uncinula* Lév. 1851.

Order: **Helotiales** Nannf., *Nova Acta R. Soc. Scient. Upsal. ser. 4* 8(2): 68 (1932).

Based on current character and taxon sampling (Wang *et al.* 2006, 2007; Spatafora *et al.* 2007), the monophyly of *Helotiales s. lat.* is not well supported. There exists a minimum of five helotialean lineages that are intermixed with other leotiomycetan taxa (e.g. *Cyttariales*, *Erysiphales*) resulting in a paraphyletic *Helotiales s. lat.* The interrelationships of these taxa are poorly resolved, however, thus preventing the synthesis of an accurate phylogenetic classification at this time. *Leotiomycetes* represents one of the more undersampled higher taxa among the *Ascomycota*, and it is likely that future sampling will result in a phylogenetic classification of a more restricted *Helotiales* and the recognition of additional orders based on current helotialean families (e.g. *Leotiaceae* or *Helotiaceae*, *Sclerotiniaceae*).

Exemplar genera: Mitrula Fr. 1821, *Hymenoscyphus* Gray 1821, *Ascocoryne* J.W. Groves & D.E. Wilson 1967.

Order: **Rhytismatales** M. E. Barr ex Minter, *in* Hawksworth & Eriksson, *Syst. Ascom.* **5**: 182 (1986).

Synonym: *Rhytismatales* M. E. Barr, *Mem. N. Y. bot. Gdn* **28**: 6 (1976), *nomen nudum*.

Exemplar genera: Rhytisma Fr. 1818, *Lophodermium* Chevall. 1826, *Cudonia* Fr. 1849.

Order: **Thelebolales** P. F. Cannon, *in* Kirk *et al.*, *Ainsworth & Bisby's Dict. Fungi* (9th edn): xi (2001).

Exemplar genera: Thelebolus Tode 1790, *Coprotus* Korf ex Korf & Kimbr. 1967, *Ascozonus* (Renny) E.C. Hansen 1876.

Class: **Lichinomycetes** Reeb, Lutzoni & Cl. Roux., *Mol. Phylogen. Evol.* **32**: 1055 (2004).

Order: **Lichinales** Henssen & Büdel, *in* Hawksworth & Eriksson, *Syst. Ascom.* **5**: 138 (1986).

Exemplar genera: Heppia Nägeli ex A. Massal. 1854, *Lichina* C. Agardh 1817, *Peltula* Nyl. 1853.

Class: **Orbiliomycetes** O. E. Erikss. & Baral, *in* Eriksson *et al.*, *Myconet* **9**: 96 (2003).

Order: **Orbiliales** Baral, O. E. Erikss., G. Marson & E. Weber, *in* Eriksson *et al.*, *Myconet* **9**: 96 (2003).

Exemplar genera: Orbilia Fr. 1849, *Hyalorbilia* Baral & G. Marson 2000.

Class: **Pezizomycetes** O. E. Erikss. & Winka, *Myconet* **1**: 8 (1997).

Order: **Pezizales** J. Schröt *in* Engler & Prantl (eds), *Nat. Pflanzenfam.* **1**(1): 173 (1894), as "*Pezizineae*".

Exemplar genera: Peziza Fr. 1822, *Glaziella* Berk. 1880, *Morchella* Dill. ex Pers. 1794, *Pyronema* Carus 1835, *Tuber* F.H. Wigg. 1780.

Glaziella has been described several times, *inter alia* as a zygomycete. Gibson *et al.* (1986) demonstrated it was an ascomycete and proposed a new family and order close to *Pezizales*, but small subunit rRNA gene sequences show that it should be included in *Pezizales* (Landvik & Eriksson 1994).

Class: **Sordariomycetes** O. E. Erikss. & Winka, *Myconet* **1**: 10 (1997).

Subclass: **Hypocreomycetidae** O. E. Erikss. & Winka, *Myconet* **1**: 6 (1997).

Order: **Coronophorales** Nannf., *Nova Acta R. Soc. Scient. Upsal. ser. 4* **8**(2): 54 (1932).

Exemplar genera: Nitschkia G.H. Otth ex P. Karst. 1873, *Scortechinia* Sacc. 1885, *Bertia* De Not. 1844, *Chaetosphaerella* E. Müll. & C. Booth 1972.

Order: **Hypocreales** Lindau, in Engler & Prantl (eds), *Nat. Pflanzenfam.* **1**(1): 343 (1897).

Exemplar genera: Hypocrea Fr. 1825, *Nectria* (Fr.) Fr. 1849, *Cordyceps* (Fr.) Link 1833, *Claviceps* Tul. 1853, *Niesslia* Auersw. 1869.

Order: **Melanosporales** N. Zhang & M. Blackw., **ord. nov.**

Mycobank no.: MB 501294

Ascomata perithecialia vel nonnumquam ostiolo carentia; peridium ascomatis e basi glomeris ascogonialis oriundum, translucidum; centrum pseudoparenchymaticum, paraphysibus absentibus; asci unitunicati, evanescentes; ascosporeae fuscae, poro germinationis utrinque praeditae; anamorphae hyphomycetales. Fungi saepe mycoparasitici.

Typus: Melanospora Corda 1837.

Ascoma perithecial or secondarily cleistothecial, peridium derived from base of an ascogonial coil, translucent; centrum pseudoparenchymatous, paraphyses absent in development; asci unitunicate, evanescent; ascospores dark, with germ pores at both ends; anamorphs hyphomycetous; often mycoparasitic.

Exemplar genus: Melanospora Corda 1837.

Order: **Microascales** Luttr. ex Benny & Kimbr., *Mycotaxon* **12**: 40 (1980).

Synonym: *Microascales* Luttr., *Univ. Miss. Stud.* **24**(2): 108 (1951), *nomen nudum*.

The group as recognized here includes members of the *Halosphaeriales*. In Zhang *et al.* (2007) and Tang *et al.* (2007), the *Halosphaeriales* were maintained separate from the *Microascales*.

Exemplar genera: Microascus Zukal 1885, *Petriella* Curzi 1930, *Halosphaeria* Linder 1944, *Lignincola* Höhnk 1955, *Nimbospora* J. Koch 1982.

Subclass: **Sordariomycetidae** O. E. Erikss. & Winka, *Myconet* 1: 10 (1997).

Order: **Boliniales** P. F. Cannon, in Kirk *et al.*, *Ainsworth & Bisby's Dict. Fungi* (9th edn): x (2001).

Exemplar genera: Camarops P. Karst. 1873, *Apiocamarops* Samuels & J.D. Rogers 1987.

Order: **Calosphaeriales** M. E. Barr, *Mycologia* 75: 11 (1983).

This order has not been placed in a subclass but the work of Réblová *et al.* (2004a) shows that it may be related to the *Diaporthales*. Members of this group were not included in Zhang *et al.* (2007) or Tang *et al.* (2007).

Exemplar genera: Calosphaeria Tul. & C. Tul. 1863, *Togniniella* Réblová, L. Mostert, W. Gams & Crous 2004, *Pleurostoma* Tul. & C. Tul. 1863.

Order: **Chaetosphaeriales** Huhndorf, A. N. Mill. & F. A. Fernández, *Mycologia* 96: 378 (2004).

Exemplar genera: Chaetosphaeria Tul. & C. Tul. 1863, *Melanochaeta* E. Müll., Harr & Sulm. 1969, *Zignoëlla* Sacc. 1878, *Striatosphaeria* Samuels & E. Müll. 1979.

Order: **Coniochaetales** Huhndorf, A. N. Mill. & F. A. Fernández, *Mycologia* **96**: 378 (2004a).

Exemplar genera: *Coniochaeta* (Sacc.) Cooke 1887, *Coniochaetidium* Malloch & Cain 1971.

Order: **Diaporthales** Nannf., *Nova Acta R. Soc. Scient. upsal., ser. 4* **8**(2): 53 (1932).

Exemplar genera: *Diaporthe* Nitschke 1870, *Gnomonia* Ces. & De Not. 1863, *Cryphonectria* (Sacc.) Sacc. & D. Sacc. 1905, *Valsa* Fr. 1849.

Order: **Ophiostomatales** Benny & Kimbr., *Mycotaxon* **12**: 48 (1980).

Exemplar genera: *Ophiostoma* Syd. & P. Syd. 1919, *Fragosphaeria* Shear 1923.

Order: **Sordariales** Chadeff. ex D. Hawksw. & O. E. Erikss., *Syst. Ascom.* **5**: 182 (1986).

Synonym: *Sordariales* Chadeff., in Chadeffaud & Emberger, *Traité Bot.* **1**: 594 (1960), *nomen nudum*.

Exemplar genera: *Sordaria* Ces. & De Not. 1863, *Podospora* Ces. 1856, *Neurospora* Shear & B.O. Dodge 1927, *Lasiosphaeria* Ces. & De Not. 1863, *Chaetomium* Kunze 1817.

Subclass: **Xylariomycetidae** O. E. Erikss. & Winka, *Myconet* **1**: 12 (1997).

Order: **Xylariales** Nannf., *Nova Acta R. Soc. Scient. Upsal., ser. 4* **8**(2): 66 (1932).

Exemplar genera: *Xylaria* Hill ex Schrank 1789, *Hypoxylon* Bull. 1791, *Anthostomella* Sacc. 1875, *Diatrype* Fr. 1849, *Graphostroma* Piroz 1974.

Sordariomycetes incertae sedis (not placed in any subclass)

Order: **Lulworthiales** Kohlm., Spatafora & Volkm-Kohlm., *Mycologia* **92**: 456 (2000).

This order includes members formerly placed in the *Spathulosporales*.

Exemplar genera: *Lulworthia* G. K. Sutherl. 1916, *Lindra* I.M. Wilson 1956.

Order: **Meliolales** Gäum. ex D. Hawksw. & O. E. Erikss., *Syst. Ascom.* **5**: 180 (1986).

Synonym: *Meliolales* Gäum., *Pilze* (2nd edn): 158 (1964), *nomen nudum*.

Exemplar genus: *Meliola* Fr. 1825.

Order: **Phyllachorales** M. E. Barr, *Mycologia* **75**: 10 (1983).

Exemplar genus: *Phyllachora* Nitschke ex Fuckel 1870.

Order: **Trichosphaeriales** M. E. Barr, *Mycologia* **75**: 11 (1983).

Exemplar genus: *Trichosphaeria* Fuckel 1870.

Pezizomycotina incertae sedis (not placed in any class)

Order: **Lahmiales** O. E. Erikss., *Mycotaxon* **27**: 357 (1986).

Exemplar genus: *Lahmia* Körb. 1861.

Order: **Medeolariales** Korf, in Eriksson *Mycotaxon* **15**: 232 (1982).

Exemplar genus: *Medeolaria* Thaxt. 1922.

Order: **Triblidiales** O. E. Erikss., *Syst. Ascom.* **11**: 9 (1992).

Exemplar genera: Huangshania O. E. Erikss. 1992, *Pseudographis* Nyl. 1855,
Triblidium Rebert. 1804.

Phylum: **Basidiomycota** Bold ex R. T. Moore, *Bot. Mar.* **23**: 371 (1980).

Synonyms: *Basidiomycota* Bold, *Morph. Pl.*: 7, 198 (1958), *nomen nudum*;
Basidiomycetes Whittaker (1959: 220), *nomen nudum*.

In common with the practice adopted here for the names *Ascomycota* and *Fungi*, and recognizing that Moore (1980) was validating a name already used before, Bold is acknowledged in the author citation.

Subphylum: **Pucciniomycotina** R. Bauer, Begerow, J. P. Samp., M. Weiß & Oberw.,
Mycol. Progr. **5**: 45 (2006).

Equivalent to *Urediniomycetes* (Kirk *et al.* 2001; Swann & Taylor 1995; Swann *et al.* 2001).

The classification of *Pucciniomycotina* employed here parallels that of Bauer *et al.* (2006) and Aime *et al.* (2007).

Class: **Pucciniomycetes** R. Bauer, Begerow, J. P. Samp., M. Weiß & Oberw., *Mycol.*
Progr. **5**: 48 (2006).

Equivalent to *Urediniomycetidae* (Swann *et al.* 2001).

Order: **Septobasidiales** Couch ex Donk, *Persoonia* **3**: 243 (1964).

Synonym: *Septobasidiales* Couch, *Gen. Septobasidium*: 65 (1938), *nomen nudum*.

Exemplar genera: Septobasidium Pat. 1892, *Auriculosocypha* D.H. Reid & Manim.
1985.

Order: **Pachnocybales** R. Bauer, Begerow, J. P. Samp., M. Weiß & Oberw., *Mycol. Progr.* **5**: 48 (2006).

Exemplar genus: Pachnocybe Berk. 1836.

Order: **Helicobasidiales** R. Bauer, Begerow, J. P. Samp., M. Weiß & Oberw., *Mycol. Progr.* **5**: 48 (2006).

Exemplar genera: Helicobasidium Pat. 1885, *Tuberculina* Tode ex Sacc. 1880.

Order: **Platyglloeales** R. T. Moore, *Mycotaxon* **39**: 247 (1990).

Equivalent to *Platyglloeales s. str.* (Swann *et al.* 2001).

Exemplar genera: Platyglloea J. Schröt. 1887 *s. str.*, *Eocronartium* G.F. Atk. 1902.

Order: **Pucciniales** Clem. & Shear, *Gen Fungi* (2nd edn): 147 (1931).

Equivalent to *Uredinales* .

Exemplar genera: Puccinia Pers. 1801, *Uromyces* (Link) Unger 1832.

Class: **Cystobasidiomycetes** R. Bauer, Begerow, J. P. Samp., M. Weiß & Oberw., *Mycol. Progr.* **5**: 46 (2006).

Equivalent to the *Erythrobasidium–Naohidea–Sakaguchia* clade (Swann *et al.* 2001) and *Cystobasidiaceae* lineage (Weiß *et al.* 2004). Genera of *Cystobasidiomycetes* that are not placed in any order include *Sakaguchia* Y. Yamada, K. Maeda & Mikata 1994, and *Cyrenella* Goch. 1981 (Aime *et al.* 2007; Bauer *et al.* 2006).

Order: **Cystobasidiales** R. Bauer, Begerow, J. P. Samp., M. Weiß & Oberw., *Mycol. Progr.* 5: 46 (2006).

Exemplar genera: Cystobasidium (Lagerh.) Neuhoff 1924, *Occultifur* Oberw. 1990, *Rhodotorula* F.C. Harrison 1927 *pro parte*.

Order: **Erythrobasidiales** R. Bauer, Begerow, J. P. Samp., M. Weiß & Oberw., *Mycol. Progr.* 5: 46 (2006).

Exemplar genera: Erythrobasidium Hamam. 1988, Sugiyama & Komag. 1988, *Rhodotorula* F.C. Harrison 1927 *pro parte*, *Sporobolomyces* Kluyver & C.B. Niel 1924 *pro parte*, *Bannoa* Hamam. 2002.

Order: **Naohideales** R. Bauer, Begerow, J. P. Samp., M. Weiß & Oberw., *Mycol. Progr.* 5: 46 (2006).

Exemplar genus: Naohidea Oberw. 1990.

Class: **Agaricostilbomycetes** R. Bauer, Begerow, J. P. Samp., M. Weiß & Oberw., *Mycol. Progr.* 5: 45 (2006).

Equivalent to *Agaricostilbomycetidae* (Swann *et al.* 2001; Weiß *et al.* 2004).

Order: **Agaricostilbales** Oberw. & R. Bauer, *Sydowia* 41: 240 (1989).

Exemplar genera: Agaricostilbum J.E. Wright 1970 (emend. Wright, Bandoni & Oberw. 1981), *Chionosphaera* D.E. Cox 1976, *Kondoa* Y. Yamada, Nakagawa & I Banno 1989 (emend. Fonseca, Sampaio, Inácio & Fell 2000).

Order: **Spiculogloales** R. Bauer, Begerow, J. P. Samp., M. Weiß & Oberw., *Mycol. Progr.* 5: 45 (2006).

Equivalent to *Mycogloea* group (Weiß *et al.* 2004).

Exemplar genera: Mycogloea L. S. Olive 1950, *Spiculogloea* P. Roberts 1996, *Sporobolomyces* Kluyver & C. B. Niel 1924 *pro parte*.

Class: **Microbotryomycetes** R. Bauer, Begerow, J. P. Samp., M. Weiß & Oberw., *Mycol. Progr.* 5: 47 (2006).

Equivalent to *Microbotryomycetidae* (Swann *et al.* 2001; Weiß *et al.* 2004). The backbone of the *Microbotryomycetes* remains poorly resolved, and several genera of *Microbotryomycetes* are not placed in any order, including *Colacogloea* Oberw. & R. Bauer 1991, *Atractocolax* R. Kirschner, R. Bauer & Oberw. 1999, *Krieglsteinera* Pouzar 1987, *Camptobasidium* Marvanová & Suberkr. 1990, *Kriegeria* Bres. 1891 and certain species of the polyphyletic genera *Sporobolomyces* Kluyver & C. B. Niel 1924 *pro parte*, *Rhodotorula* F. C. Harrison 1927 *pro parte*, and *Leucosporidium* Fell, Statzell, I.L. Hunter & Phaff 1970, and others (Aime *et al.* 2007; Bauer *et al.* 2006; Sampaio *et al.* 2004; Weiß *et al.* 2004).

Order: **Heterogastridiales** Oberw. & R. Bauer, *Mycologia* 82: 57 (1990).

Exemplar genus: Heterogastridium Oberw. & Bauer 1990.

Bauer *et al.* (2006) placed *Colacogloea*, *Atractocolax* and *Krieglsteinera* in the *Heterogastridiales*. However, analyses of Bauer *et al.* (2006) and Aime *et al.* (2007) suggest that *Heterogastridium* and *Colacogloea* do not form a clade, while *Atractocolax* and *Krieglsteinera* have yet to be sampled in molecular phylogenetic studies.

Order: **Microbotryales** R. Bauer & Oberw., in Bauer *et al.*, *Can. J. Bot.* 75: 1309 (1997).

Exemplar genera: Microbotryum Lév. 1847, *Ustilentyloma* Savile 1964.

Order: **Leucosporidiales** J. P. Samp., M. Weiß & R. Bauer, in Sampaio *et al.*, *Mycol. Progr.* **2**: 61 (2003).

Exemplar genera: Leucosporidiella J. P. Samp. 2003, *Leucosporidium* Fell, Statzell, I. L. Hunter & Phaff 1970, *Mastigobasidium* Golubev 1999.

Order: **Sporidiobolales** J. P. Samp., M. Weiß & R. Bauer, in Sampaio *et al.*, *Mycol. Progr.* **2**: 66 (2003).

Exemplar genera: Sporidiobolus Nyland 1949, *Sporobolomyces* Kluyver & C. B. Niel 1924, *Rhodosporidium* I. Banno 1967, *Rhodotorula* F. C. Harrison 1927 *pro parte*.

Class: **Atractiellomycetes** R. Bauer, Begerow, J. P. Samp., M. Weiß & Oberw., *Mycol. Progr.* **5**: 45 (2006).

Order: **Atractiellales** Oberw. & Bandoni, *Can. J. Bot.* **60**: 1740 (1982).

Emend. Oberw. & Bauer, *Sydowia* **41**: 239 (1989).

Exemplar genera: Atractiella Sacc. 1886, *Saccoblastia* A. Møller 1895, *Helicogloea* Pat. 1892, *Phleogena* Link 1833.

Class: **Classiculomycetes** R. Bauer, Begerow, J. P. Samp., M. Weiß & Oberw., *Mycol. Progr.* **5**: 46 (2006).

Order: **Classiculales** R. Bauer, Begerow, Oberw. & Marvanová, *Mycologia* **95**: 763 (2003).

Exemplar genera: Classicula R. Bauer, Begerow, Oberw. & Marvanová 2003,
Jaculispora H. J. Huds. & Ingold 1960.

Class: **Mixiomycetes** R. Bauer, Begerow, J. P. Samp., M. Weiß & Oberw., *Mycol. Progr.* 5: 47 (2006).

Order: **Mixiales** R. Bauer, Begerow, J. P. Samp., M. Weiß & Oberw., *Mycol. Progr.* 5: 47 (2006).

Exemplar genus: Mixia C. L. Kramer 1958.

Class: **Cryptomycocolacomycetes** R. Bauer, Begerow, J. P. Samp., M. Weiß & Oberw., *Mycol. Progr.* 5: 46 (2006).

Order: **Cryptomycocolacales** Oberw. & R. Bauer, *Mycologia* 82: 672 (1990).

Exemplar genera: Cryptomycocolax Oberw. & Bauer 1990, *Colacosiphon* R. Kirschner, R. Bauer & Oberw. 2001.

Subphylum: **Ustilaginomycotina** R. Bauer, Begerow, J. P. Samp., M. Weiß & Oberw., *Mycol. Progr.* 5: 45 (2006).

Equivalent to *Ustilaginomycetes* (Bauer *et al.* 1997, . 2001; Swann & Taylor 1995). The classification of *Ustilaginomycotina* employed here largely parallels that of Begerow *et al.* (2007), with the primary differences being that here the *Entorrhizomycetes* are classified as *incertae sedis* among *Basidiomycota* (rather than being a class within *Ustilaginomycotina*).

Class: **Ustilaginomycetes** R. Bauer, Oberw. & Vánky, *Can. J. Bot.* **75**: 1311 (1997).

Emend. Begerow, Stoll & Bauer, *Mycologia* **98**: XXX (2007) ["2006"].

Equivalent to *Ustilaginomycetidae* Jülich as emmended by Bauer & Oberwinkler (Bauer *et al.* 1997, 2001; Weiß *et al.* 2004).

Order: **Urocystales** R. Bauer & Oberw., *in* Bauer *et al.*, *Can. J. Bot.* **75**: 1311 (1997).

Exemplar genera: *Urocystis* Rabenh. ex Fuckel 1870, *Ustacystis* Zundel 1945, *Doassansiopsis* (Setch.) Dietel 1897.

Melanotaenium de Bary 1874 has also been placed in this order (Bauer *et al.* 2001; Weiß *et al.* 2004), but analyses of Begerow *et al.* (2007) and Matheny *et al.* (2007b) have supported its transfer to *Ustilaginales*.

Order: **Ustilaginales** G. Winter, *Rabenh. Krypt.-Fl.* 2nd ed. **1**(1.1): 73 (1880), as "*Ustilagineae*".

Emend. Bauer & Oberwinkler, *in* Bauer *et al.*, *Can. J. Bot.* **75**: 1311 (1997).

Exemplar genera: *Ustilago* (Pers.) Roussel 1806, *Cintractia* Cornu 1883.

Thecaphora Fingerh. 1836 has also been placed in this order (Bauer *et al.* 2001), but analyses of Begerow *et al.* (2007) and Matheny *et al.* (2007b) have suggested that it is not nested in *Ustilaginales*. *Thecaphora* may be the sister group of *Urocystales* (Matheny *et al.* 2007b).

Class: **Exobasidiomycetes** Begerow, Stoll & Bauer, *Mycologia* **98**: XXX (2007) [“2006”].

Equivalent to *Exobasidiomycetidae* Jülich 1981 *emend.* Bauer & Oberwinkler, except for exclusion of *Malasseziales* (Bauer *et al.* 1997, 2001; Weiß *et al.* 2004). Monophyly of the *Exobasidiomycetidae*, as delimited here, is supported with high Bayesian posterior probability in analyses of *rpb1*, *rpb2*, and *tef1*, and nuclear *lsu*, *ssu*, and 5.8S ribosomal genes (Matheny *et al.* 2007b), but it is weakly supported in analyses using *atp6*, β -tubulin, and *nuc-lsu* ribosomal RNA genes (Begerow *et al.* 2007). See comments regarding *Malasseziales*.

Order: **Doassansiales** R. Bauer & Oberw., in Bauer *et al.*, *Can. J. Bot.* **75**: 1312 (1997).

Exemplar genera: *Doassansia* Cornu 1883, *Rhamphospora* D.D. Cunn. 1888, *Nannfeldtiomyces* Vánky 1981.

Order: **Entylomatales** R. Bauer & Oberw., in Bauer *et al.*, *Can. J. Bot.* **75**: 1311 (1997).

Exemplar genera: *Entyloma* de Bary 1874, *Tilletiopsis* Derx 1948.

Begerow *et al.* (2007) erected the monotypic order *Ceraceosorales* Begerow, Stoll & R. Bauer for *Ceraceosorus bombacis* (B. K. Bakshi) B. K. Bakshi 1976, which was weakly supported as the sister group of *Tilletiopsis albescens* Gokhale 1972. The *Ceraceosorus-T. albescens* clade was placed as the sister group of *Entylomatales*, again with weak support. *Ceraceosorales* is not included in the present classification, pending more robust resolution of the relationships among *Ceraceosorus*, *Tilletiopsis*, and *Entyloma*.

Order: **Exobasidiales** Henn., in Engler & Prantl (eds), *Nat. Pflanzenfam.* **1**(1**): 103 (1897), as "*Exobasidiineae*".

Emend. Bauer, Oberwinkler & Vánky, *Can. J. Bot.* **75**: 1312 (1997).

Exemplar genera: *Exobasidium* Woronin 1867, *Climoconidium* Pat. 1898, *Dicellomyces* L. S. Olive 1945.

Order: **Georgefischeriales** R. Bauer, Begerow & Oberw., *in* Bauer *et al.*, *Can. J. Bot.* **75**: 1311 (1997).

Exemplar genera: *Georgefischeria* Thirum. & Naras. *emend.* Gandhe 1980, *Phragmotaenium* R. Bauer, Begerow, A. Nagler & Oberw. 2001, *Tilletiaria* Bandoni & B. N. Johri 1972, *Tilleteopsis* Derx 1948 *pro parte*.

Order: **Microstromatales** R. Bauer & Oberw., *in* Bauer *et al.*, *Can. J. Bot.* **75**: 1311 (1997).

Exemplar genera: *Microstroma* Niessl 1861, *Sympodiomyopsis* Sugiy., Tokuoka & Komag. 1991, *Volvocisporium* Begerow, R. Bauer & Oberw. 2001.

Order: **Tilletiales** Kreisel ex R. Bauer & Oberw., *in* Bauer *et al.*, *Can. J. Bot.* **75**: 1311 (1997).

Exemplar genera: *Tilletia* Tul. & C. Tul. 1847, *Conidiosporomyces* Vánky 1992, *Erratomyces* M. Piepenbr. & R. Bauer 1997.

Ustilaginomycotina *incertae sedis* (not placed in any class):

Order: **Malasseziales** R. T. Moore, *Bot. Mar.* **23**: 371 (1980).

Emend. Begerow, Bauer & Boekhout, *Mycol. Res.* **104**: 59 (2000).

Exemplar genus: *Malassezia* Baill. 1889.

Analyses of the protein-coding genes *rpb1*, *rpb2*, and *tef1*, alone or in combination with nuclear *lsu*, *ssu*, and 5.8S ribosomal genes, suggest that *Malasseziales* are included in the *Ustilaginomycetes*, but analyses of nuclear ribosomal genes alone or in combination with *atp6* and β -tubulin suggest that *Malasseziales* is in the *Exobasidiomycetes* (Bauer *et al.* 2001; Begerow *et al.* 2007; Matheny *et al.* 2007b; Weiß *et al.* 2004).

Subphylum: **Agaricomycotina** R. Bauer, Begerow, J. P. Samp., M. Weiß & Oberw.,
Mycol. Progr. 5: 45 (2006).

Equivalent to *Hymenomycetes* (Swann & Taylor 1995) or *Basidiomycetes* (Kirk *et al.* 2001; Hibbett 2007).

Class: **Tremellomycetes** Hibbett, Matheny & Binder, **class. nov.**

Mycobank no.: MB 501295

Fungi dimorphici; basidiomata gelatinosa vel absentia; parenthesesmata sacculata vel raro absentia; basidia septata vel aseptata.

Typus: Tremella Pers. 1794.

Dimorphic fungi. Fruiting bodies gelatinous or absent, parenthesesmata sacculate or absent, basidia septate or non-septate. The least inclusive clade containing *Tremellales*, *Filobasidiales* and *Cystofilobasidiales*.

Equivalent to *Tremellomycetidae* sensu Swann & Taylor (1995) and Weiß *et al.* (2004). The name *Tremellomycetidae* was earlier published by Locquin (1984), but without a Latin diagnosis, and it is therefore invalid under the *Code*.

Order: **Cystofilobasidiales** Fell, Roeyjmans & Boekhout, *Int. J. Syst. Bacteriol.* **49**: 911 (1999).

Exemplar genera: Cystofilobasidium Oberw. & Bandoni 1983, *Mrakia* Y. Yamada & Komag. 1987, *Itersonilia* Derx 1948.

Order: **Filobasidiales** Jülich, *Bibliotheca Mycol.* **85**: 324 (1981).

Exemplar genera: Filobasidiella Kwon-Chung 1976, *Cryptococcus* Vuill. 1901 (*pro parte*).

Order: **Tremellales** Fr. *Syst. Mycol.* **1**: 2 (1821), as "*Tremellinae*".

As delimited here, the group includes *Trichosporonales* Boekhout & Fell 2001 (Fell *et al.* 2001) and *Christianseniales* F. Rath 1991 (Wells & Bandoni 2001). *Filobasidiales*, which Weiß *et al.* (2004) included in *Tremellales s. lat.*, has been resolved as the sister group of *Tremellales* (Fell *et al.* 2001; Matheny *et al.* 2007b; Swann & Taylor 1995).

Exemplar genera: Tremella Pers. 1794, *Trichosporon* Behrend 1890, *Christiansenia* Hauerslev 1969.

Class: **Dacrymycetes** Hibbett, Matheny, Binder & M. Weiß, **class. nov.**

Mycobank no.: MB 501296

Basidiomata gelatinosa; basidia bifurcata, raro unispora; parenthesomata imperforata.

Typus: Dacrymyces Nees 1816.

Fruiting bodies gelatinous, basidia furcate (rarely unisporous), parenthesomes imperforate.

Containing the single order *Dacrymycetales* (Wells & Bandoni 2001).

Order: **Dacrymycetales** Henn., in Engler & Prantl (eds), *Nat. Pflanzenfam.* **1**(1**): 96 (1898), as "*Dacryomycetinae*".

Exemplar genera: *Dacrymyces* Nees 1861, *Calocera* (Fr.) Fr. 1828, *Guepiniopsis* Pat. 1883.

Class: **Agaricomycetes** Matheny, Hibbett & Binder, **class. nov.**

MycoBank no.: MB 501297

Basidiomata hymenomycetoidea vel gasteroidea; basidia 2-8 sporas formantia; parenthesesmata perforata vel imperforata.

Typus: *Agaricus* L. 1753

Fruiting bodies hymenomycetous or gasteroid, basidia two- to eight-spored, parenthesesmata perforate or imperforate. The least-inclusive clade containing *Auriculariales*, *Sebacinales*, *Cantharellales*, *Phallomycetidae* and *Agaricomycetidae*.

This group is approximately equivalent to *Homobasidiomycetes sensu* Hibbett & Thorn (2001) plus *Auriculariales* and *Sebacinales*.

Subclass: **Agaricomycetidae** (Fr.) Parm., *Windahlia* **16**: 16 (1986).

The least-inclusive clade containing *Agaricales*, *Boletales* and *Atheliales*.

The delimitation of *Agaricomycetidae* adopted here differs from that of Parmasto (1986), who described *Agaricomycetidae* as a subclass of *Cantharellomycetes* Parm. 1986. For example, many of the resupinate forms in the *Agaricomycetidae* were placed by Parmasto in the *Corticiumycetes* Parm. 1986. The name *Agaricomycetidae* was also published by Locquin (1984), but without a Latin diagnosis and it is therefore invalid under the *Code*.

Order: **Agaricales** Underw., *Moulds, Mildews Mushrooms*: 97 (1899).

Equivalent to euagarics clade (Hibbett & Thorn, 2001).

Exemplar genera: *Agaricus* L. 1753, *Coprinus* Pers. 1797, *Pleurotus* (Fr.) P. Kumm. 1871.

Order: **Atheliales** Jülich, *Bibliotheca Mycol.* **85**: 343 (1981).

Equivalent to athelioid clade (Binder *et al.* 2005; Larsson *et al.* 2004).

Exemplar genera: *Athelia* Pers. 1822, *Piloderma* Jülich 1969, *Tylospora* Donk 1960.

Order: **Boletales** E.-J. Gilbert, *Livres Mycol.* **3**: 83 (1931).

Equivalent to bolete clade (Binder & Hibbett 2006; Hibbett & Thorn 2001).

Exemplar genera: *Boletus* Fr. 1821, *Scleroderma* Pers. 1801, *Coniophora* DC 1815, *Rhizopogon* Fr. & Nordholm 1817.

Subclass: **Phallomycetidae** K. Hosaka, Castellano & Spatafora, *Mycologia* **98**: XXX (2007) ["2006"].

Equivalent to *Phallales sensu* Kirk *et al.* (2001), and the gomphoid-phalloid clade (Hibbett & Thorn 2001; Hosaka *et al.* 2007).

Order: **Geastrales** K. Hosaka & Castellano, *Mycologia* **98**: XXX (2007) ["2006"].

Exemplar genera: *Geastrum* Pers. 1794, *Radiigera* Zeller 1944, *Sphaerobolus* Tode 1790.

Order: **Gomphales** Jülich, *Bibliotheca Mycol.* **85**: 348 (1981).

Exemplar genera: Gomphus (Fr.) Weinm. 1826, *Gautieria* Vittad. 1831, *Ramaria* Holmsk. 1790.

Order: **Hysterangiales** K. Hosaka & Castellano, *Mycologia* **98**: XXX (2007) [“2006”].

Exemplar genera: Hysterangium Vittad. 1831, *Phallogaster* Morgan 1893, *Gallacea* Lloyd 1905, *Austrogautieria* E. L. Stewart & Trappe 1985.

Order: **Phallales** E. Fisch., in Engler & Prantl (eds), *Nat. Pflanzenfam.* **1**(1**): 276 (1898).

Equivalent to *Phallomycetidae* Locq. (Locquin 1984), which was invalidly published, owing to the absence of a Latin diagnosis.

Exemplar genera: Phallus Junius ex L. 1753, *Clathrus* P. Micheli ex L. 1753, *Claustula* K. M. Curtis 1926.

Agaricomycetes *incertae sedis* (not placed in any subclass):

Order: **Auriculariales** J. Schröt., in Cohn (ed.), *Krypt.-Fl. Schlesien* **1**: 382 (1889).

Exemplar genera: Auricularia Bull. ex Juss. 1789, *Exidia* Fr. 1822, *Bourdotia* (Bres.) Trotter 1913.

Order: **Cantharellales** Gäum., *Vergl. Morph. Pilze*: 495 (1926).

Equivalent to the cantharelloid clade (Hibbett & Thorn 2001; Moncalvo *et al.* 2007). The *Cantharellales* as delimited here includes *Tulasnella*, which is distinguished by unusual basidia with inflated sterigmata, and has been classified in a separate order, *Tulasnellales* Rea 1922 (e.g. Weiß *et al.* 2004). Extreme evolutionary rate heterogeneity in the nuclear ribosomal RNA genes of *Tulasnella*, *Cantharellus* and *Craterellus* is a source of error in

phylogenetics of *Cantharellales*. Analyses of Matheny *et al.* (2006b) suggest that *Tulasnella* is nested within the *Cantharellales*, but it could also be the sister group to *Cantharellales s.str.* (Moncalvo *et al.* 2007). If so, then it may be appropriate to segregate *Tulasnellales* from *Cantharellales s.str.*

Exemplar genera: Cantharellus Fr. 1821, *Botryobasidium* Donk 1931, *Craterellus* Pers. 1825, *Tulasnella* J. Schröt. 1888.

Order: **Corticiales** K. H. Larss., **ord. nov.**

MycoBank no.: MB 501299

Basidiomata resupinata, effuso-reflexa vel discoidea; hymenophora laevia; systema hypharum monomiticum; dendrohyphidia raro absentia; basidia saepe e probasidiis oriuntur. Cystidia presentia vel absentia. Sporae hyalinae, tenuitunicatae, albae vel aggregatae roseae.

Typus: Corticium Pers. 1794.

Basidiomycetes with effused or discoid (*Cytidia*) basidiomata, a smooth hymenophore, and a monomititic hyphal system with clamped, rarely simple-septate, hyphae.

Dendrohyphidia common. Species with or without cystidia. A probasidial resting stage is present in many species. Spores smooth, in masses white to pink. Saprotrophic, parasitic, or lichenicolous.

Equivalent to *Vuilleminiales* Boidin, Mugnier & Canales 1998 and the corticioid clade (Binder *et al.* 2005; Larsson *et al.* 2004). Boidin *et al.* (1998) explicitly included *Corticium* in their new order, as a member of the family *Vuilleminiaceae* Maire 1902. Jülich (1981) also placed *Corticium* in *Vuilleminiaceae* but referred them to *Aleurodiscales* Jülich 1981. *Corticium* is the type of *Corticaceae* Herter 1910, a family name conserved

against *Vuilleminiaceae*. The introduction of *Corticiales* as a new name for this order is, therefore, the preferred option.

Exemplar genera: Corticium Pers. 1794, *Vuilleminia* Maire 1902, *Punctularia* Pat. 1895.

Order: Gloeophyllales Thorn, **ord. nov.**

MycoBank no.: MB 501300

Basidiomata annua vel perennia, resupinata, effuso-reflexa, dimidiata vel pileata; hymenophora laevia, merulioidea, odontioidea vel poroidea. Systema hypharum monomiticum, dimiticum vel trimiticum. Hyphae generativae fibulatae vel efibulatae. Leptocystidia ex trama in hymenium projecta, hyalina vel brunnea, tenuitunicata vel crassitunicata. Basidiosporae laeves, hyalinae, tenuitunicatae, ellipsoideae vel cylindricae vel allantoideae, inamyloideae. Lignum decompositum brunneum vel album.

Typus: Gloeophyllum P. Karst. 1882.

Fruiting bodies perennial or annual and long-lived, with hymenium maturing and thickening over time. Stature resupinate, effused-reflexed or dimidiate, with smooth, wrinkled, dentate, lamellate or regularly poroid hymenophore, or pileate-stipitate with lamellae. (Aborted, coralloid or flabelliform fruiting bodies may be formed under conditions of darkness or high CO₂ concentration). Leptocystidia or hyphoid hairs originating in the context and extending into or protruding from the hymenial layer (or lamellar margin in *Neolentinus*) are common; these often with thick brown walls and brownish incrustation. Context brown (but pallid in *Neolentinus*) and generally darkening in KOH (the brownish incrustation in *Boreostereum* turning green in KOH). Monomitic (if so, with sclerified generative hyphae), dimitic, or trimitic; generative hyphae with or without clamp connections. Basidiospores hyaline, ellipsoid to

cylindrical or subballantoid, with thin, smooth walls, and neither amyloid, dextrinoid nor cyanophilous. Where this is known, basidiospores are binucleate and sexuality is heterothallic and bipolar (but tetrapolar in *V. berkeleyi*).

Causing brown rots (*Gloeophyllum*, *Neolentinus*, *Veluticeps*) or stringy white rot (*Boreostereum*, *Donkioporia*) of wood of gymnosperms, monocots and dicots. Occurrence on "wood in service" (e.g. railway ties, paving blocks, wooden chests) seems to be common (in *Donkioporia*, *Gloeophyllum*, *Heliocybe* and *Neolentinus*); often on charred wood (*Boreostereum* and *Veluticeps*).

Equivalent to *Gloeophyllum* clade (Binder *et al.* 2005).

Exemplar genera: *Gloeophyllum* P. Karst. 1882, *Neolentinus* Redhead & Ginns 1985, *Veluticeps* (Cooke) Pat. 1894.

Order: **Hymenochaetales** Oberw., in Frey *et al.* (eds), *Beitr. Biol. niederen Pflanz.*: 89 (1977).

Equivalent to the hymenochaetoid clade (Hibbett & Thorn 2001; Larsson *et al.* 2007).

Exemplar genera: *Hymenochaete* Lév. 1846, *Phellinus* Quél. 1886, *Trichaptum* Murrill 1904.

Order: **Polyporales** Gäum., *Vergl. Morph. Pilze*: 503 (1926).

Equivalent to polyporoid clade (Hibbett and Thorn 2001).

Exemplar genera: *Polyporus* Fr. 1815, *Fomitopsis* P. Karst. 1881, *Phanerochaete* P. Karst. 1889.

Order: **Russulales** Kreisel ex P.M. Kirk, P.F. Cannon & J.C. David, in Kirk *et al.*, *Ainsworth & Bisby's Dict. Fungi* (9th edn): xi (2001).

Equivalent to the russuloid clade (Hibbett & Thorn 2001; Larsson & Larsson 2003; Miller *et al.* 2007).

Exemplar genera: Russula Pers. 1796, *Aleurodiscus* Rabenh. ex J. Schröt. 1888, *Bondarzewia* Singer 1940, *Hericium* Pers. 1794, *Peniophora* Cooke 1879, *Stereum* Pers. 1794.

Order: **Sebacinales** M. Weiß, Selosse, Rexer, A. Urb. & Oberw., *Mycol. Res.* **108**: 1007 (2004).

Exemplar genera: Sebacina Tul. 1871, *Tremellodendron* G.F. Atk. 1902, *Piriformospora* Sav. Verma, Aj. Varma, Rexer, G. Kost & P. Franken 1998.

Order: **Thelephorales** Corner ex Oberw., *Sydowia* **78**: 361 (1976).

Equivalent to the thelephoroid clade (Hibbett & Thorn 2001).

Exemplar genera: Thelephora Ehrh. ex Willd. 1787, *Bankera* Coker & Beers ex Pouzar 1955, *Polyozellus* Murrill 1910.

Order: **Trechisporales** K.H. Larss., **ord. nov.**

Mycobank no.: MB 501301

Basidiomata resupinata, stipitata vel clavarioidea. Hymenophora laevia, grandinioidea, hydnoidea vel poroidea. Systema hypharum monomiticum vel dimiticum. Hyphae fibulatae, septa hypharum interdum inflata (ampullata). Cystidia praesentia vel absentia. Basidia 4-6 sterigmata formantia. Sporae laeves vel ornatae. Species lignicolae vel terricolae.

Typus: Trechispora P. Karst. 1890.

Basidiomycetes with effused, stipitate or clavarioid basidiomata. Hymenophore smooth, grandinoid, hydroid or poroid. Hyphal system monomitic, hyphae clamped,

subicular hyphae with or without ampullate septa. Cystidia present in some species, mostly lacking. Basidia with four to six sterigmata. Spores smooth or ornamented. On wood or soil.

Equivalent to *Hydnodontales* Jülich 1981 and trechisporoid clade (Binder *et al.* 2005; Larsson *et al.* 2004). *Hydnodon* Banker 1913 was recently placed in synonymy under *Trechispora* (Ryvarden 2002) and this synonymy is supported by molecular data (K.H. Larsson, unpubl.). The introduction of a new name for the group, a name that connects to the clade name already established and that is based on the most species-rich genus is, therefore, justified.

Exemplar genera: Trechispora P. Karst. 1890, *Sistotremastrum* J. Erikss. 1958, *Porpomyces* Jülich 1982.

Basidiomycota *incertae sedis* (not placed in any subphylum):

Class: **Wallemiomycetes** Zalar, de Hoog & Schroers, *Antonie van Leeuwenhoek* **87**: 322 (2005).

Analyses of *rpb1*, *rpb2*, *tef1*, and *nuc-lsu*, *nuc-ssu*, and 5.8S ribosomal RNA genes suggest that the *Wallemiomycetes* is the sister group of the rest of the *Basidiomycota* (possibly along with *Entorrhizomycetes*, see below), but subsets of this dataset produce alternative placements (Matheny *et al.* 2007b; Zalar *et al.* 2005).

Order: **Wallemiales** Zalar, de Hoog & Schroers, *Antonie van Leeuwenhoek* **87**: 322 (2005).

Exemplar genus: Wallemia Johan-Olsen 1887.

Class: **Entorrhizomycetes** Begerow, Stoll & R. Bauer, *Mycologia* **98**: XXX (2007) ["2006"].

Equivalent to *Entorrhizomycetidae* R. Bauer & Oberw. (Bauer *et al.* 1997). So far, only ribosomal RNA genes have been sequenced in *Entorrhizomycetes*. Analyses with broad sampling across all groups of *Basidiomycota* and including *Ascomycota* and *Glomeromycota* as outgroups suggest that *Entorrhizomycetes* is not nested within any subphylum, and may be the sister group of the rest of the *Basidiomycota* (Matheny *et al.* 2007a; also see Begerow *et al.* 1997).

Order: **Entorrhizales** R. Bauer & Oberw., in Bauer *et al.*, *Can. J. Bot.* **75**: 1311 (1997).

Exemplar genus: Entorrhiza C. A. Weber 1884.

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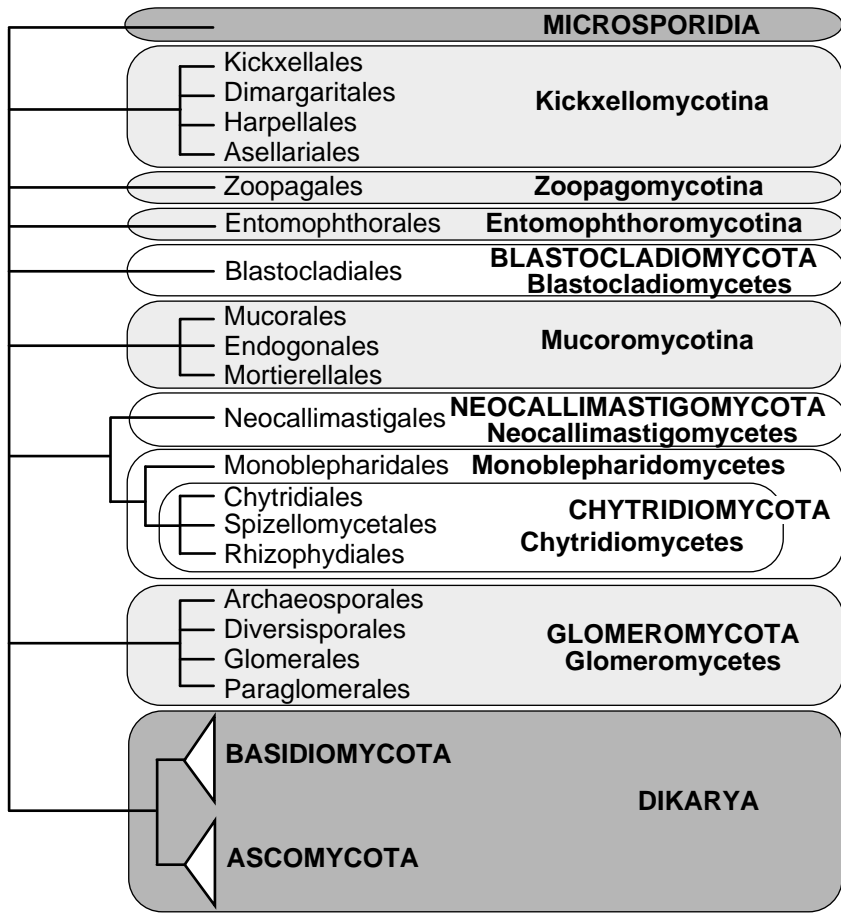
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Fig 1– Phylogeny and classification of *Fungi*. Basal *Fungi* and *Dikarya*. Branch lengths are not proportional to genetic distances. See Table 1 for support values for clades.

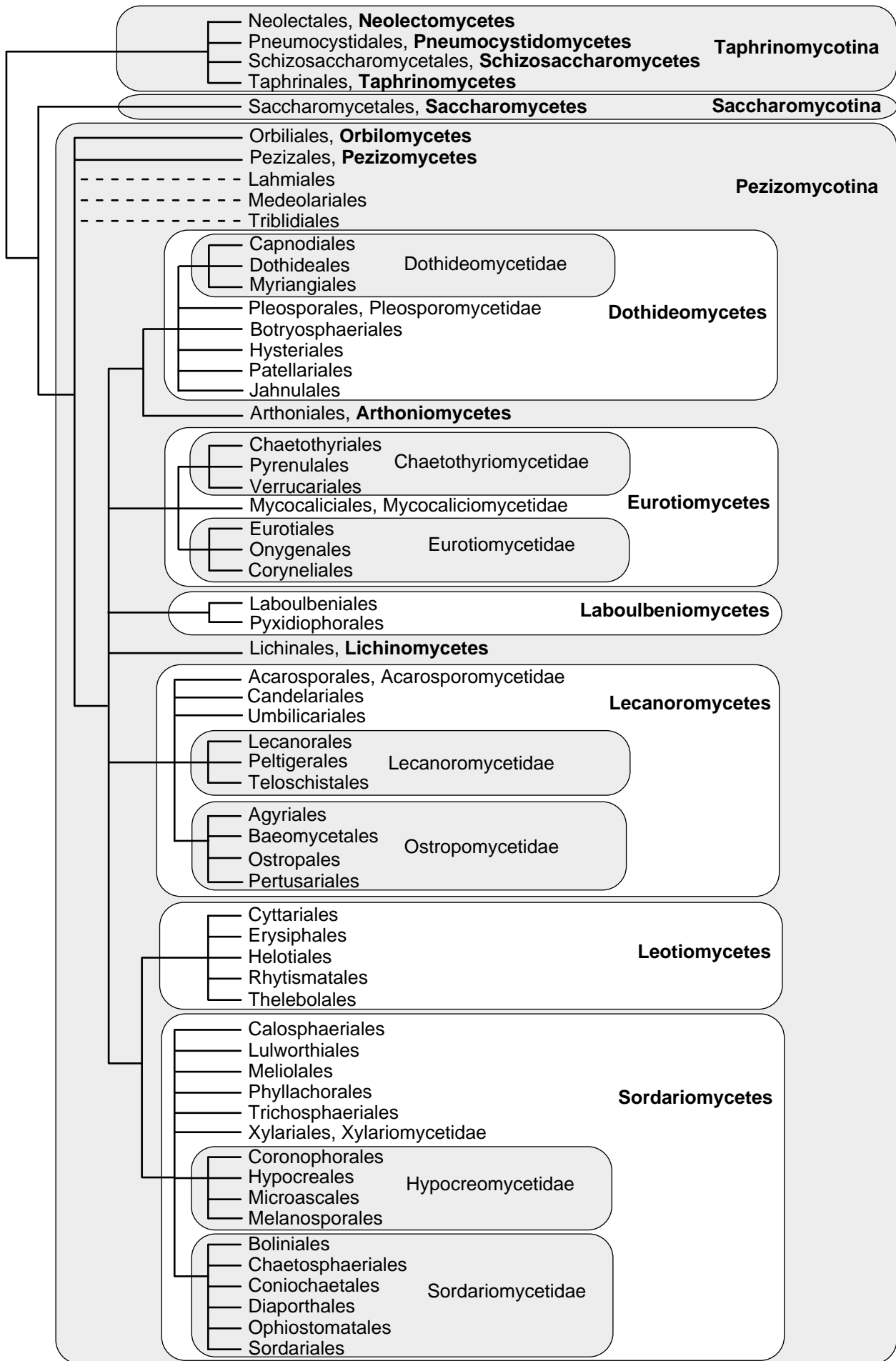
Fig 2 – Phylogeny and classification of *Fungi*. *Ascomycota*. See Table 2 for support values for clades. Dashed lines indicate taxa that are of uncertain placement.

Fig 3 – Phylogeny and classification of *Fungi*. *Basidiomycota*. See Table 3 for support values for clades. Dashed lines indicate taxa that are of uncertain placement.



Traditional Zygomycota

Traditional Chytridiomycota



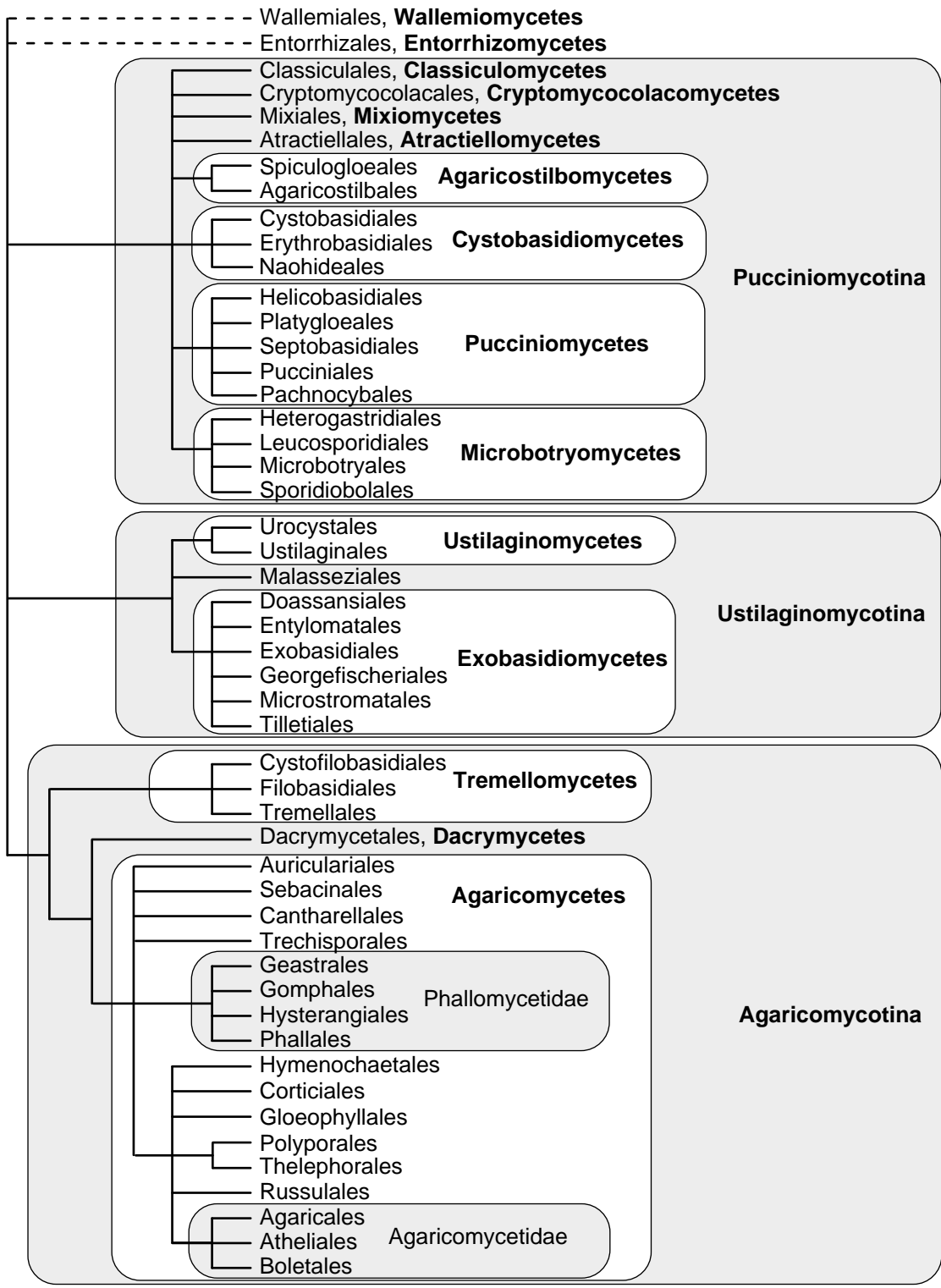


Table 1. Support for major groups of Fungi in selected phylogenetic studies: Basal Fungi and Dikarya. **Rank:** K = kingdom, SK = subkingdom, P = phylum, SP = subphylum, C = class, SC = subclass, O = order, G = genus. Taxa with only one subsidiary taxon included (i.e., redundant taxa) are listed on a single line, with rank abbreviations divided by a slash (e.g., the class Agaricostilbomycetes, which contains a single order, Agaricostilbales, is indicated as C/O). **Data:** lsu, ssu, and 5.8S refer to nuclear rRNA genes, whereas mt-lsu and mt-ssu refer to mitochondrial rRNA genes, other genes follow standard abbreviations. Some datasets contain missing sequences. **OTUs:** indicates the number of OTUs in the specified clade, not the total number of OTUs in the dataset. **Support:** bs = bootstrap %, jk = jackknife %, MP = maximum parsimony (equally weighted), WP = weighted parsimony, NJ = neighbor joining, ML = maximum likelihood, RML = RaxML, PML = PhyML, ME = minimum evolution, Bpp = Bayesian posterior probability, NA = not applicable because the group is monotypic, or only a single species was sampled in the reference study.

Rank	Taxon	Reference	Data	OTUs	Support
K	FUNGI	Keeling (2003)	α -tub, β -tub	38	MLbs = 98 NJbs = 94
P	CHYTRIDIOMYCOTA	Baldauf <i>et al.</i> (2000)	<i>act</i> , α -tub, β -tub, <i>tef1</i>	12	MLbs = 85 MPbs = 95
		James <i>et al.</i> (2007)	lsu, ssu, 5.8S	84	Bpp \geq 0.95
		Seif <i>et al.</i> (2005)	mt-genome	5	Bpp = 1.00 MLbs = 100
C	Chytridiomycetes	James <i>et al.</i> (2006)	lsu, ssu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i>	8	Bpp \geq 0.95 MLbs \geq 70
		James <i>et al.</i> (2007)	lsu, ssu, 5.8S	75	Bpp \geq 0.95 MLbs \geq 70
		Keeling (2003)	α -tub, β -tub	5	MLbs = 90 NJbs = 95
O	Chytridiales	James <i>et al.</i> (unpublished)	lsu, ssu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i> , <i>Atp6</i>	9	MLbs = 98
O	Rhizophydiales	James <i>et al.</i> (2006)	lsu, ssu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i>	2	Bpp \geq 0.95 MLbs \geq 70
		Letcher <i>et al.</i> (2006)	lsu, 5.8S	96	MPbs=100 Bpp = 1.00
O	Spizellomycesales	James <i>et al.</i> (2007)	lsu, ssu, 5.8S	9	MPbs = 100
C/O	Monoblepharidomycetes, Monoblepharidales	James <i>et al.</i> (2007)	lsu, ssu, 5.8S	9	Bpp \geq 0.95 MLbs \geq 70
		Bullerwell <i>et al.</i> (2003)	<i>cox 1,2,3; cob, atp6,9; nad 1,2,3,4, 4L,6</i>	4	MPbs \geq 70 MLbs = 100

P/C/O	NEOCALLIMASTIGOMYCOTA, <i>Neocallimastigomycetes</i> , <i>Neocallimastigales</i>	James <i>et al.</i> (2007)	Isu, ssu, 5.8S	6	Bpp \geq 0.95 MLbs \geq 70 MPbs \geq 70 Bpp \geq 0.95
P/C/O	BLASTOCLADIOMYCOTA, <i>Blastocladiomycetes</i> , <i>Blastocladiiales</i>	James <i>et al.</i> (2007)	Isu, ssu, 5.8S	10	
P	MICROSPORIDIA	Liu <i>et al.</i> (2006)	<i>rpb1</i> , <i>rpb2</i>	3	Bpp = 1.00 MPbs = 100
P/C	GLOMEROMYCOTA, <i>Glomeromycetes</i>	James <i>et al.</i> (2006)	Isu, ssu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tefl</i>	5	Bpp \geq 0.95 MLbs \geq 70 MLbs = 100 NJbs = 97
O	<i>Archaeosporales</i>	Schüßler <i>et al.</i> (2001)	ssu	72	NJbs \geq 90
O	<i>Diversisporales</i>	Schüßler <i>et al.</i> (2001)	ssu	5	NJbs \geq 95
O	<i>Glomerates</i>	Schüßler <i>et al.</i> (2001)	ssu	32	NJbs \geq 95
O	<i>Paraglomerates</i>	Schüßler <i>et al.</i> (2001)	ssu	32	NJbs \geq 95
SP	<i>Subphyla incertae sedis (not placed in any phylum)</i> <i>Mucoromycotina</i>	Schüßler <i>et al.</i> (2001)	ssu	3	NJbs \geq 95
O	<i>Mucorales</i>	James <i>et al.</i> (2006)	Isu, ssu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tefl</i> <i>rpb1</i>	11	Bpp = 1.00
O		Tanabe <i>et al.</i> (2004)	<i>rpb1</i>	4	NJbs = 82
		James <i>et al.</i> (2006)	Isu, ssu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tefl</i> <i>rpb1</i>	3	Bpp \geq 0.95 MLbs \geq 70
		Tanabe <i>et al.</i> (2004)	<i>rpb1</i>	3	NJbs = 100
		Keeling (2003)	α - <i>tub</i> , β - <i>tub</i>	4	MLbs = 96 NJbs = 98
O	<i>Endogonales</i>	White <i>et al.</i> (2007)	Isu, ssu, 5.8S	28	Bpp = 1.00 MPbs \geq 70
O	<i>Mortierellales</i>	White <i>et al.</i> (2007)	Isu, ssu, 5.8S	2	Bpp = 1.00 MPbs \geq 70
SP/O	<i>Entomophthoromycotina</i> , <i>Entomophthorales</i>	White <i>et al.</i> (2007)	Isu, ssu, 5.8S	6	Bpp = 1.00 MPbs \geq 70
SP/O	<i>Zoopagomycotina</i> , <i>Zoopagales</i>	James <i>et al.</i> (2006)	Isu, ssu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tefl</i>	2	Bpp \geq 0.95 MLbs \geq 70
		James <i>et al.</i> (2006)	Isu, ssu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tefl</i> <i>rpb1</i>	2	Bpp \geq 0.95 MLbs \geq 70
		Tanabe <i>et al.</i> (2004)	<i>rpb1</i>	3	MLbs \geq 70 NJbs = 86

SP	<i>Kickxellomycotina</i>	Tanabe <i>et al.</i> (2004)	<i>rpb1</i>	6	N bs = 84
O	<i>Kickxellales</i>	O'Donnell <i>et al.</i> (1998)	<i>ssu</i>	7	MPbs = 100
O	<i>Dimargaritales</i>	Tanabe <i>et al.</i> (2000)	<i>ssu</i>	3	N bs = 100
O	<i>Harpellales</i>	Tanabe <i>et al.</i> (2004)	<i>rpb1</i>	3	N bs = 98
O	<i>Asellariales</i>	O'Donnell <i>et al.</i> (1998)	<i>ssu</i>	4	MPbs = 100
SK	<i>DIKARYA</i>	---	---	---	---
		James <i>et al.</i> (2006)	<i>lsu, ssu, 5.8S, rpb1, rpb2, tef1</i>	161	Bpp = 1.00 MLbs = 71
		Steenkamp <i>et al.</i> (2006)	<i>act, α-tub, β-tub, tef1</i>	10	Bpp = 1.00 MLbs = 84 MPbs = 82
		Seif <i>et al.</i> (2005)	mt-genome	10	N bs = 96 Bpp = 1.00 MLbs = 100
		Liu <i>et al.</i> (2006)	<i>rpb1, rpb2</i>	27	Bpp = 1.00 MPbs = 100

Table 2. Support for major groups of Fungi in selected phylogenetic studies: Ascomycota. See Table 1 for explanation.

Rank	Taxon	Reference	Data	OTUs	Support
P	ASCOMYCOTA	James <i>et al.</i> (2006a, fig. 1) Spatafora <i>et al.</i> (2007, fig. 2)	ssu, lsu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i> lsu, ssu, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i>	111 177	MLbs = 94 Bpp = 1.00 WPbs = <50 MLbs = 100 Bpp = 1.00 NJbs = 67 Bpp = 1.00 MLbs = 98 Bpp = 1.00 WPbs = <50 MLbs = 98 Bpp = 1.00 Bpp = 1.00 Bpp = 1.00
SP	<i>Taphrinomycotina</i>	Lutzoni <i>et al.</i> (2004, fig. 2) James <i>et al.</i> (2006a, fig. 2) Spatafora <i>et al.</i> (2007, fig. 2)	lsu, ssu ssu, lsu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i> lsu, ssu, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i>	276 4 8	Bpp = 1.00 NJbs = 67 Bpp = 1.00 MLbs = 98 Bpp = 1.00 WPbs = <50 MLbs = 98
C/O	<i>Taphrinomycetes</i> , <i>Taphrinales</i>	Liu <i>et al.</i> (2006, fig. 3) Sugiyama <i>et al.</i> (2007, fig. 2) Kurtzman & Sugiyama (2001, fig. 7) Sugiyama <i>et al.</i> (2007, fig. 2)	<i>rpb1</i> , <i>rpb2</i> lsu, ssu <i>rpb2</i> , β - <i>tub</i> ssu	3 11 8	Bpp = 1.00 Bpp = 1.00 Bpp = 1.00 NJbs = 54
C/O	<i>Nelectomycetes</i> , <i>Nelectales</i>	Kurtzman & Sugiyama (2001, fig. 7) Nishida & Sugiyama (1994, fig. 1) Lutzoni <i>et al.</i> (2004, fig. 2) Sugiyama <i>et al.</i> (2007, fig. 2)	lsu, ssu <i>rpb2</i> , β - <i>tub</i> ssu ssu	6 4 5	Bpp = 1.00 NJbs = 100 NJbs = 100
C/O	<i>Pneumocystidomycetes</i> , <i>Pneumocystidales</i>	Landvik <i>et al.</i> (2001, fig. 1) Sugiyama <i>et al.</i> (2007, fig. 2)	lsu, ssu lsu, ssu, β - <i>tub</i> , <i>rpb2</i> β - <i>tub</i>	1 2 2	NA Bpp = 1.00 MPbs = 100
C/O	<i>Schizosaccharomycetes</i> , <i>Schizosaccharomycetales</i>	Lutzoni <i>et al.</i> (2004, fig. 2) Sugiyama <i>et al.</i> (2007, fig. 2)	lsu, ssu ssu, lsu, <i>rpb2</i> , β -	1 1	NA NA

	<p><i>Taphrinomycotina incertae sedis (not placed in any subphylum)</i> <i>Saitoella</i></p>	<p>Lutzoni <i>et al.</i> (2004, fig. 2)</p>	<p><i>tub</i> lsu, ssu</p>	<p>2</p>	<p>Bpp = 1.0 NJbs = 100</p>
G		<p>Sugiyama <i>et al.</i> (2007, fig. 2)</p>	<p>ssu, lsu, <i>rpb2</i>, β-<i>tub</i> ssu</p>	<p>1</p>	<p>NA</p>
SP/C/O	<p><i>Saccharomycotina</i>, <i>Saccharomycetes</i>, <i>Saccharomycetales</i></p>	<p>Nishida & Sugiyama (1994, fig. 1) Spatafora <i>et al.</i> (2007, fig. 2)</p>	<p>lsu, ssu, <i>rpb1</i>, <i>rpb2</i>, <i>tefl</i></p>	<p>12</p>	<p>WPbs = 55 MLbs = 100 Bpp = 1.00 MPbs = 99</p>
SP	<p><i>Pezizomycotina</i></p>	<p>Suh <i>et al.</i> (2007, fig. 2)</p>	<p>lsu, ssu</p>	<p>87</p>	<p>Bpp = 1.00 Bpp = 1.00 MLbs = 94 MPbs = 94-100</p>
SP		<p>James <i>et al.</i> (2006a, fig. 1)</p>	<p>ssu, lsu, 5.8S, <i>rpb1</i>, <i>rpb2</i>, <i>tefl</i> Genomes</p>	<p>46</p>	<p>Bpp = 1.00 Bpp = 1.00 MLbs = 94 MPbs = 94-100</p>
SP		<p>Robbertse <i>et al.</i> (2006, figs. 4,5,6)</p>		<p>11</p>	
C/O	<p><i>Arthoniomycetes</i>, <i>Arthoniales</i></p>	<p>Spatafora <i>et al.</i> (2007, fig. 2)</p>	<p>lsu, ssu, <i>rpb1</i>, <i>rpb2</i>, <i>tefl</i></p>	<p>157</p>	<p>NJbs = 100 MLbs = 100 WPbs = 100 MLbs = 97</p>
C/O		<p>Spatafora <i>et al.</i> (2007, fig. 2)</p>	<p>lsu, ssu, <i>rpb1</i>, <i>rpb2</i>, <i>tefl</i></p>	<p>4</p>	<p>Bpp = 1.00 WPbs = 100 MLbs = 100</p>
C	<p><i>Dothideomycetes</i></p>	<p>Lumbsch <i>et al.</i> (2005, fig. 1)</p>	<p>lsu, ssu, mt-ssu, mt-lsu</p>	<p>6</p>	<p>Bpp = 1.00 MPbs = 100</p>
C		<p>Schoch <i>et al.</i> (2007, fig.1)</p>	<p>lsu, ssu, <i>rpb2</i>, <i>tefl</i></p>	<p>96</p>	<p>Bpp = 1.00 Bpp = 1.00 MPbs < 50 MLbs = 70</p>
C		<p>Spatafora <i>et al.</i> (2007, fig. 2)</p>	<p>lsu, ssu, <i>rpb1</i>, <i>rpb2</i>, <i>tefl</i></p>	<p>17</p>	<p>WPbs < 50 MLbs = 84</p>
C		<p>Kruys <i>et al.</i> (2006, fig.1)</p>	<p>lsu, ssu, mt-ssu</p>	<p>51</p>	<p>Bpp = 1.00 Bpp > 0.95</p>

SC	<i>Dothideomycetidae</i>	Schoch <i>et al.</i> (2007, fig.1)	lsu, ssu, <i>rpb2</i> , <i>tefl</i>	26	MPbs < 50 Bpp = 1.00 MPbs > 50 MLbs > 0.7 Bpp > 0.95 MPbs < 50 Bpp = 1.00 MPbs > 70 MLbs > 70 Bpp = 1.00 MPbs > 70 MLbs > 70 Bpp > 0.95 MPbs = 100 MLbs = 91 NJbs = 100 Bpp = 1.00 MPbs > 70 MLbs > 70 Bpp = 1.00 MPbs > 70 MLbs > 70 Bpp = 1.00 MPbs = 100
O	<i>Capnodiales</i>	Kruys <i>et al.</i> (2006, fig.1)	lsu, ssu, mt-ssu	11	
O	<i>Dothideales</i>	Schoch <i>et al.</i> (2007, fig.1)	lsu, ssu, <i>rpb2</i> , <i>tefl</i>	11	
O		Schoch <i>et al.</i> (2007, fig.1)	lsu, ssu, <i>rpb2</i> , <i>tefl</i>	9	
O		Kruys <i>et al.</i> (2006, fig.1)	lsu, ssu, mt-ssu	4	
O	<i>Myriangiales</i>	Lindemuth <i>et al.</i> (2001)	lsu, ssu	6	
O		Schoch <i>et al.</i> (2007, fig.1)	lsu, ssu, <i>rpb2</i> , <i>tefl</i>	5	
SC/O	<i>Pleosporomycetidae, Pleosporales</i>	Schoch <i>et al.</i> (2007, fig.1)	lsu, ssu, <i>rpb2</i> , <i>tefl</i>	48	
O	<i>Dothideomycetes incertae sedis (not placed in any subclass)</i> <i>Botryosphaeriales</i>	Kruys <i>et al.</i> (2006, fig.1)	lsu, ssu, mt-ssu	35	
O	<i>Hysteriales</i>	Schoch <i>et al.</i> (2007, fig.1)	lsu, ssu, <i>rpb2</i> , <i>tefl</i>	8	Bpp = 1.00 MPbs > 70 MLbs > 70
O	<i>Patellariales</i>	Schoch <i>et al.</i> (2007, fig.1)	lsu, ssu, <i>rpb2</i> , <i>tefl</i>	3	Bpp = 1.00 MPbs > 70 MLbs > 70
O	<i>Jahnulales</i>	Pang <i>et al.</i> (2002, fig. 26) Inderbitzin <i>et al.</i> (2001, fig. 18) Pang <i>et al.</i> (2002, fig. 26)	ssu ssu ssu	1 1 6	NA NA MPbs = 100

C	Eurotiomycetes	<p>Spatafora <i>et al.</i> (2007, fig. 2)</p> <p>Geiser <i>et al.</i> (2007, fig. 1)</p> <p>Ekman & Tønsberg (2002, fig. 1)</p> <p>Del Prado <i>et al.</i> (2005, fig. 1)</p> <p>Lumbsch <i>et al.</i> (2005, fig. 1)</p> <p>Lutzoni <i>et al.</i> (2004, fig. 5)</p> <p>Reeb <i>et al.</i> (2004, fig.1)</p> <p>Reeb <i>et al.</i> (2004, fig.1)</p> <p>Lutzoni <i>et al.</i> (2004, fig. 5)</p> <p>Del Prado <i>et al.</i> (2005, fig.1)</p> <p>Spatafora <i>et al.</i>(2006, fig.1)</p> <p>Geiser <i>et al.</i> (2007, fig.1)</p> <p>Lutzoni <i>et al.</i> (2004, fig. 2)</p> <p>Liu and Hall (2004, fig. 3)</p>	<p>lsu, ssu, <i>rpb1</i>, <i>rpb2</i>, <i>tef1</i></p> <p>ssu, <i>lsu</i>, <i>rpb1</i>, <i>rpb2</i>, <i>tef</i></p> <p>ssu</p> <p><i>lsu</i>, mt-ssu</p> <p><i>lsu</i>, ssu, mt-ssu, mt-<i>lsu</i></p> <p><i>lsu</i>, ssu, mt-SSU, <i>rpb2</i></p> <p>ssu, <i>lsu</i>, <i>rpb2</i></p> <p>ssu, <i>lsu</i>, <i>rpb2</i></p> <p><i>lsu</i>, ssu, mt-ssu, <i>rpb2</i></p> <p><i>lsu</i>, mit ssu</p> <p>ssu, <i>lsu</i>, <i>rpb1</i>, <i>rpb2</i>, <i>tef</i></p> <p>ssu, <i>lsu</i>, <i>rpb1</i>, <i>rpb2</i>, <i>tef</i></p> <p><i>lsu</i>, ssu</p> <p><i>rpb2</i></p>	<p>11</p> <p>49</p> <p>13</p> <p>15</p> <p>11</p> <p>8</p> <p>7</p> <p>5</p> <p>5</p> <p>11</p> <p>6</p> <p>21</p> <p>5</p> <p>5</p>	<p>WPbs = 89 MLbs = 84 Bpp = 1.00 Bpp = 1.00 MPbs = 100 WPbs = 100 MLbs = 100 Bpp = 0.99</p> <p>Bpp = 1.00 Bpp > 0.95 MPbs > 70 Bpp = 1.00 Bbs = 61 Bpp = 1.00 Bbs = 89</p> <p>Bpp = 1.00 Bbs = 100 MLbs = 100 Bpp = 1.00 Bbs = 100 NJbs = 99 MPbs = 98 Bpp = 1.00 Bpp = 1.00 MLbs = 100 WPbs > 70 Bpp = 1.00 MPbs = 100 WPBbs = 100 MLbs = 100 Bpp = 1.00 NJbs= 94 Bpp = 1.00 MPbs = 96</p>
SC	<i>Chaetothyriomycetidae</i>				
O	<i>Chaetothyriales</i>				

O	<i>Pyrenulales</i>	Spatafora <i>et al.</i> (2007, fig. 1)	ssu, lsu, <i>rpb1</i> , <i>rpb2</i> , <i>tef</i>	4	Bpp = 1.00 MLbs = 100 WPbs > 70
		Geiser <i>et al.</i> (2007, fig. 1)	ssu, lsu, <i>rpb1</i> , <i>rpb2</i> , <i>tef</i>	9	Bpp = 1.00 MPbs = 100 WPBbs = 100 MLbs = 100
		Lutzoni <i>et al.</i> (2004, fig. 8)	lsu, ssu, mt-ssu, <i>rpb2</i>	2	Bpp = 1.00 NJbs = 100
		Reeb <i>et al.</i> (2004, fig. 1)	lsu, ssu, <i>rpb2</i>	2	WPbs = 100 Bpp = 1.00 Bbs = 100 MLbs = 100
		Schmitt <i>et al.</i> (2004, fig. 1)	lsu, mt-ssu	2	Bpp = 1.00
		Geiser <i>et al.</i> (2007, fig. 1)	ssu, lsu, <i>rpb1</i> , <i>rpb2</i> , <i>tef</i>	5	Bpp = 1.00 MPbs = 100 WPBbs = 100 MLbs = 100
		Wedin <i>et al.</i> (2006, fig. 1)	lsu, mt-ssu	3	Bpp = 1.00 MPjk = 100
		Geiser <i>et al.</i> (2007, fig. 1)	ssu, lsu, <i>rpb1</i> , <i>rpb2</i> , <i>tef</i>	7	Bpp = 1.00 MPbs = 100 WPBbs = 100 MLbs = 100
		Lutzoni <i>et al.</i> (2004, fig. 2)	lsu, ssu	3	Bpp = 1.00 NJbs = 98
		Gueidan <i>et al.</i> (2007, fig. 2)	lsu, ssu, <i>rpb1</i>	83	Bpp = 1.00 MLbs = 100 MPbs = 100
		Geiser <i>et al.</i> (2007, fig. 1)	ssu, lsu, <i>rpb1</i> , <i>rpb2</i> , <i>tef</i>	24	Bpp = 1.00 MPbs = 100 WPBbs = 98 MLbs = 100
SC	<i>Eurotiomycetidae</i>	Lutzoni <i>et al.</i> (2004, fig. 2)	lsu, ssu	11	NJbs = 96 Bpp = 1.00

O	<i>Coryneliales</i>	Winka (2000, fig. 1)	ssu	2	MPbs = 100 NJbs = 100 NA
		Inderbitzin et al (2004, fig. 14)	ssu	1	
		Geiser <i>et al.</i> (2007, fig.1)	ssu, lsu, <i>rpb1</i> , <i>rpb2</i> , <i>tef</i>	3	Bpp = 1.00 MPbs = 100 WPBbs = 100 MLbs = 100
O	<i>Eurotiales</i>	Geiser <i>et al.</i> (2007, fig. 1)	ssu, lsu, <i>rpb1</i> , <i>rpb2</i> , <i>tef</i>	9	Bpp = 1.00 MPbs = 100 WPBbs = 100 MLbs = 100
O	<i>Omygenales</i>	Geiser <i>et al.</i> (2007, fig. 1)	ssu, lsu, <i>rpb1</i> , <i>rpb2</i> , <i>tef</i>	12	Bpp = 1.00 MPbs = 65 WPBbs = 68 MLbs = 88
SC/O	<i>Mycocaliciomycetidae, Mycocaliciales</i>	Tibell & Vinuesa (2005, fig. 1) Geiser <i>et al.</i> (2007, fig. 1)	lsu ssu, lsu, <i>rpb1</i> , <i>rpb2</i> , <i>tef</i>	20 4	Bpp = 1.00 Bpp = 1.00 MPbs = 100 WPBbs = 100 MLbs = 100 Bpp = 1.00
C	<i>Laboulbeniomycetes</i>	Ekman & Tønsberg (2002, fig. 1)	ssu	4	MPbs = 100 MPbs = 100
O	<i>Laboulbeniales</i>	Weir & Blackwell (2001, fig. 2) Henk, Weir, & Blackwell (2003, fig. 1) Weir & Blackwell (2001, fig. 1) Henk, Weir, & Blackwell (2003, fig. 2)	ssu ssu	4 6	MPbs = 100 MPbs = 100
O	<i>Pyxidiales</i>	Weir & Blackwell (2001, fig. 2) Henk, Weir, & Blackwell (2003, fig. 2)	ssu ssu	3 3	MPbs = 100 MPbs = 57
C	<i>Lecanoromycetes</i>	Lutzoni <i>et al.</i> (2004, fig. 5)	ssu ssu	1 2	NA MPbs = 99
		Spatafora <i>et al.</i> (2007, fig. 2)	lsu, ssu, <i>rpb2</i> , mt-ssu lsu, ssu, <i>rpb1</i> ,	34 38	Bpp = 1.00 Bbs = 56 WPbs <50

SC/O	<i>Acarosporomycetidae, Acarosporales</i>	<p>Miadlikowska <i>et al.</i> (2007, fig. 1)</p> <p>Hofstetter <i>et al.</i> (2006, fig. 1)</p> <p>Miadlikowska <i>et al.</i> (2007, fig. 1)</p> <p>Reeb <i>et al.</i> (2004, fig. 1)</p> <p>Lutzoni <i>et al.</i> (2004, fig. 4)</p>	<p><i>rpb2, tef1</i></p> <p>lsu, ssu, <i>rpb1</i>, <i>rpb2</i>, mt-ssu</p> <p>lsu, ssu, <i>rpb1</i>, <i>rpb2</i>, mt-ssu</p> <p>lsu, ssu, <i>rpb1</i>, <i>rpb2</i>, mt-ssu</p>	<p>264</p> <p>82</p> <p>15</p> <p>14</p> <p>14</p> <p>71</p> <p>54</p> <p>14</p> <p>86</p> <p>30</p> <p>14</p> <p>8</p> <p>46</p>	<p>MLbs = 93 Bpp = 1.00 RMLbs > 70</p> <p>Bpp > 0.95 RMLbs > 70</p> <p>Bpp > 0.95 RMLbs > 70%</p> <p>PMLbs > 70%</p> <p>Bpp > 0.95 MLbs = 100</p> <p>Bpp = 100</p> <p>Bpp = 1.00</p> <p>Nlbs = 100</p> <p>MPbs = 100</p> <p>RMLbs > 70%</p> <p>PMLbs > 70%</p> <p>Bpp > 0.95 RMLbs > 70</p> <p>Bpp > 0.95 MLbs = 73</p> <p>Bpp = 100 RMLbs > 70</p> <p>Bpp > 0.95 RMLbs > 70</p> <p>Bpp = 1.00 Bpp = 1.00 RMLbs > 70</p>
SC	<i>Lecanoromycetidae</i>	<p>Miadlikowska <i>et al.</i> (2007, fig. 1)</p>	<p>lsu, ssu, <i>rpb1</i>, <i>rpb2</i>, mt-ssu</p>	<p>71</p>	<p>RMLbs > 70%</p>
O	<i>Lecanorales</i>	<p>Hofstetter <i>et al.</i> (2006, fig. 1)</p> <p>Reeb <i>et al.</i> (2004, fig. 1)</p> <p>Miadlikowska <i>et al.</i> (2007, fig. 1)</p> <p>Hofstetter <i>et al.</i> (2006, fig. 1)</p>	<p>lsu, ssu, <i>rpb1</i>, <i>rpb2</i>, mt-ssu</p> <p>lsu, ssu, <i>rpb1</i>, <i>rpb2</i>, mt-ssu</p>	<p>54</p> <p>14</p> <p>86</p>	<p>Bpp > 0.95 RMLbs > 70</p> <p>Bpp > 0.95 MLbs = 73</p> <p>Bpp = 100 RMLbs > 70</p>
O	<i>Peltigerales</i>	<p>Lumbsch <i>et al.</i> (2004, fig. 1)</p> <p>Lucking <i>et al.</i> (2004, fig. 3)</p> <p>Miadlikowska <i>et al.</i> (2007, fig. 1)</p>	<p>lsu, ssu, <i>rpb1</i>, <i>rpb2</i>, mt-ssu</p> <p>lsu, mt-ssu</p> <p>lsu, mt-ssu</p> <p>lsu, ssu, <i>rpb1</i>, <i>rpb2</i>, mt-ssu</p>	<p>14</p> <p>8</p> <p>46</p>	<p>Bpp > 0.95 RMLbs > 70</p> <p>Bpp > 0.95 Bpp = 1.00</p> <p>Bpp = 1.00 RMLbs > 70</p> <p>Bpp > 0.95</p>

		Miadlikowska & Lutzoni (2004, fig. 1)	lsu, ssu	59	MPbs < 70 Bpp = 0.92 Bjk = 99
O	<i>Teloschistales</i>	Wilklund & Wedin (2003, fig. 1)	lsu, ssu	31	
SC	<i>Ostropomycetidae</i>	Miadlikowska <i>et al.</i> (2007, fig. 1)	lsu, ssu, <i>rpb1</i> , <i>rpb2</i> , mt-ssu	13	RMLbs > 70
		Miadlikowska <i>et al.</i> (2007, fig. 1)	lsu, ssu, <i>rpb1</i> , <i>rpb2</i> , mt-ssu	58	Bpp > 0.95 RMLbs > 70
O	<i>Agyriales</i>	Grube <i>et al.</i> (2004, fig. 1)	mt-ssu	30	Bpp > 0.95
		Reeb <i>et al.</i> (2004, fig. 1)	lsu, ssu, <i>rpb2</i>	16	Bpp > 0.95 MLbs = 100
O		Miadlikowska <i>et al.</i> (2007, fig. 1)	lsu, ssu, <i>rpb1</i> , <i>rpb2</i> , mt-ssu	8	Bpp = 100 RMLbs > 70
O	<i>Baeomycetales</i>	Lucking <i>et al.</i> (2004, fig. 3)	lsu, mt-ssu	11	Bpp > 0.95
		Lutzoni <i>et al.</i> (2004, fig. 2)	lsu, ssu	4	Bpp = 1.00 NJbs = 100
O		Wedin <i>et al.</i> (2005, fig. 1)	lsu, mt-ssu	8	MPjk = 83 Bpp = 0.99
O		Miadlikowska <i>et al.</i> (2007, fig. 1)	lsu, ssu, <i>rpb1</i> , <i>rpb2</i> , mt-ssu	4	RMLbs > 70 PMLbs > 70
O	<i>Ostropales</i> s.l.	Wedin <i>et al.</i> (2005, fig. 1)	lsu, mt-ssu	3	Bpp > 0.95 MPjk = 99 Bpp = 1.0
O		Miadlikowska <i>et al.</i> (2007, fig. 1)	lsu, ssu, <i>rpb1</i> , <i>rpb2</i> , mt-ssu	21	RMLbs > 70
O		Schmitt <i>et al.</i> (2005, fig. 1)	lsu, mt-ssu	12	Bpp > 0.95
		Wedin <i>et al.</i> (2005, fig. 1)	lsu, mt-ssu	13	Bpp = 1.00 Bjk = 94
		Lutzoni <i>et al.</i> (2004, fig. 4)	lsu, ssu, <i>rpb2</i>	10	Bpp = 0.97 Bpp = 1.00 NJbs = 74 MPbs = 84

O	<i>Pertusariales</i>	Reeb <i>et al.</i> (2004, fig. 1)	lsu, ssu, <i>rpb2</i>	9	MLbs = 99 Bpp = 1.00 Bbs = 1.00 RMLbs > 70 Bpp > 0.95 Bpp = 1.00 Bpp = 1.00 Bpp = 1.00
O	<i>Lecanoromycetes incertae sedis</i> (not placed in any subclass) <i>Candelariales</i>	Miadlikowska <i>et al.</i> (2007, fig. 1) Lücking <i>et al.</i> (2004, fig. 3) Schmitt <i>et al.</i> (2005, fig. 1) Lutzoni <i>et al.</i> (2004, fig. 2) Wedin <i>et al.</i> (2005, fig. 1) Hofstetter <i>et al.</i> (2006, fig. 1) Miadlikowska <i>et al.</i> (2007, fig. 1) Miadlikowska <i>et al.</i> (2007, fig. 1) Miadlikowska <i>et al.</i> (2007, fig. 1) Hofstetter <i>et al.</i> (2006, fig. 1) Reeb <i>et al.</i> (2004, fig. 1) Spatafora <i>et al.</i> (2007, fig. 2) Wang <i>et al.</i> (2006, fig. 1) Wang <i>et al.</i> (2007, fig. 2) Wang <i>et al.</i> (2007, fig. 1)	lsu, ssu, <i>rpb1</i> , <i>rpb2</i> , mt-ssu lsu, mt-ssu lsu, mt-ssu lsu, ssu lsu, mt-ssu lsu, ssu, mt-ssu, <i>rpb1</i> , <i>rpb2</i> lsu, ssu, mt-ssu, <i>rpb1</i> , <i>rpb2</i> lsu, ssu, <i>rpb1</i> , <i>rpb2</i> , mt-ssu lsu, ssu, <i>rpb1</i> , <i>rpb2</i> , mt-ssu lsu, ssu, <i>rpb1</i> , <i>rpb2</i> , mt-ssu lsu, ssu, <i>rpb2</i> lsu, ssu, <i>rpb1</i> , <i>rpb2</i> , <i>tefl</i> lsu, ssu, 5.8S lsu, ssu, 5.8S	3 2 3 16 9 8 4 22 50 78 1	Jk = 100 Bpp = 0.96 RMLbs > 70 Bpp > 0.95 RMLbs > 70 PMLbs > 70 Bpp > 0.95 Bpp > 0.95 RMLbs > 70 PMLbs > 70 Bpp > 0.95 RMLbs > 70 MLbs = 70 Bpp = 1.00 Bbs = 88 WPbs = 100 MLbs = 100 Bpp = 1.00 Bpp = 1.00 MPbs = 61 Bpp = 1.00 NA
C	<i>Leotiomyces</i> (w/o <i>Geoglossaceae</i>)				
O	<i>Cyttariales</i>	Wang <i>et al.</i> (2007, fig. 1)	ssu, lsu, 5.8S	1	

O	<i>Erysiphales</i>	Rossmann <i>et al.</i> (2004, fig. 2) Wang <i>et al.</i> (2007, fig. 1)	lsu ssu, lsu, 5.8S	12 16	MPbs >55 MPbs = 63 Bpp = 0.97 NJbs = 99
O	<i>Helotiales (w/o Geoglossaceae)</i>	Takamatsu (2004, fig. 2)	ssu	10	Bpp < 0.90 MPbs >55
O	<i>Rhytismatales</i>	Wang <i>et al.</i> (2007, fig. 1) Rossmann <i>et al.</i> (2004, fig. 2) Wang <i>et al.</i> (2007, fig. 1)	ssu, lsu, 5.8S lsu ssu, lsu, 5.8S	40 4 5	MPbs = 100 Bpp = 1.00 MPbs = 56 NA
O	<i>Thelebolales</i>	de Hoog <i>et al.</i> (2005, fig. 3)	ssu	11	
C/O	<i>Lichinomycetes, Lichinales</i>	Spatafora <i>et al.</i> (2007, fig. 2)	lsu, ssu, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i>	1	
		Miadlikowska <i>et al.</i> (2007, fig. 1)	lsu, ssu, <i>rpb1</i> , <i>rpb2</i> , mt-ssu	2	RMLbs > 70 PMLbs > 70 Bpp > 0.95 MLbs = 100 Bbs = 100
		Reeb <i>et al.</i> (2004, fig. 1)	lsu, ssu, <i>rpb2</i>	3	
C/O	<i>Orbiliomycetes, Orbiliales</i>	Spatafora <i>et al.</i> (2007, fig. 2)	lsu, ssu, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i>	2	Bpp = 1.00 WPbs = 100 MLbs = 100
C/O	<i>Pezizomycetes, Pezizales</i>	Spatafora <i>et al.</i> (2007, fig. 2)	lsu, ssu, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i>	14	Bpp = 1.00 WPbs = 54 MLbs = 99
C	<i>Sordariomycetes</i>	Lutzoni <i>et al.</i> (2004, fig. 2) Spatafora <i>et al.</i> (2007, fig. 2)	lsu, ssu	21	Bpp = 1.00 Bpp = 0.96 NJbs = 70
		Zhang <i>et al.</i> (2007, fig. 2)	lsu, ssu, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i>	47	WPbs = 100 MLbs = 100 Bpp = 1.00 MPbs = 100 WPbs = 100 MLbs = 100
		Lutzoni <i>et al.</i> (2004, fig. 2)	lsu, ssu, <i>rpb2</i> , <i>tef1</i>	106	
		Zhang <i>et al.</i> (2007, fig. 2)	lsu, ssu	66	Bpp = 1.00 Bpp = 1.00 NJbs = 97
SC	<i>Hypocreomycetidae</i>	Zhang <i>et al.</i> (2007, fig. 2)	lsu, ssu, <i>rpb2</i> , <i>tef1</i>	42	MPbs = 92

O	Coronophorales	Lutzoni <i>et al.</i> (2004, fig. 2) Huhndorf <i>et al.</i> (2004b, figs. 38, 39) Zhang <i>et al.</i> (2007, fig. 2) Huhndorf <i>et al.</i> (2004b, figs. 38, 39) Miller & Huhndorf (2005, fig. 7) Zhang <i>et al.</i> (2007, fig. 2)	<i>tef1</i> lsu, ssu lsu lsu, ssu, <i>rpb2</i> , <i>tef1</i> lsu lsu, β - <i>tub</i> , <i>rpb2</i> lsu, ssu, <i>rpb2</i> , <i>tef1</i> lsu, ssu lsu, ssu, <i>rpb2</i> , <i>tef1</i> lsu, ssu, <i>rpb2</i> , <i>tef1</i>	26 21 2 16 2 21 31 2 15 10 40 16 54	WPbs = 96 MLbs = 90 Bpp = 1.00 NJbs < 50 Bpp = 1.00 MPbs = 67 Bpp >= 0.95 MPbs < 50 WPbs < 50 MLbs = 96 Bpp = 1.00 WPbs = 99 Bpp >= 95 WPbs = 100 Bpp >= 95 MPbs = 91 WPbs = 90 MLbs = 72 Bpp = 1.00 MPbs = 70 Bpp = 1.00 MPbs = 100 WPbs = 100 MLbs = 100 Bpp = 1.00 MPbs = 74 WPbs = 86 MLbs = 85 Bpp = 1.00 NJbs = 80 Bpp = 1.00 MPbs = 100 Bpp = 1.00 MPbs = 97 MPbs = 82
O	Hypocreales				
O	Melanosporales				
O	Microascales (incl. Halosphaeriales)				
SC	Sordariomycetidae	Kohlmeyer <i>et al.</i> (2000, fig. 1) Zhang <i>et al.</i> (2007, fig. 2)	lsu, ssu lsu, ssu, <i>rpb2</i>	16 54	Bpp = 1.00 MPbs = 97 MPbs = 82

O	Boliniales	Lutzoni <i>et al.</i> (2004, fig. 2) Zhang <i>et al.</i> (2007, fig. 2) Huhndorf <i>et al.</i> (2004a, fig. 1) Miller & Huhndorf (2005, fig. 7) Zhang <i>et al.</i> (2007, fig. 2) Miller & Huhndorf (2005, fig. 7) Shenoy <i>et al.</i> (2006, fig. 3) Zhang <i>et al.</i> (2007, fig. 2) Miller & Huhndorf (2005, fig. 7) Miller & Huhndorf (2004, fig. 10) Zhang <i>et al.</i> (2007, fig. 2) Castlebury <i>et al.</i> (2002, fig. 1) Lutzoni <i>et al.</i> (2004, fig. 2)	<i>tefl</i> lsu, ssu lsu, ssu, <i>rpb2</i> , <i>tefl</i> lsu lsu, β - <i>tub</i> , <i>rpb2</i> lsu, ssu, <i>rpb2</i> , <i>tefl</i> lsu, β - <i>tub</i> , <i>rpb2</i> lsu, <i>rpb2</i> lsu, ssu, <i>rpb2</i> , <i>tefl</i> lsu, β - <i>tub</i> , <i>rpb2</i> lsu lsu, ssu, <i>rpb2</i> , <i>tefl</i> lsu lsu, ssu	36 4 3 2 3 2 4 3 2 3 19 82 10	WPbs = 85 MLbs = 77 Bpp = 1.00 NJbs < 50 Bpp = 0.97 MPbs = 100 WPbs = 100 MLbs = 100 Bpp = 1.00 WPbs = 99 Bpp < 95 WPbs = 100 Bpp \geq 95 MPbs = 100 WPbs = 100 MLbs = 100 Bpp = 1.00 WPbs = 100 Bpp \geq 95 MPbs = 100 MPbs = 93 WPbs = 100 MLbs = 87 Bpp = 1.00 WPbs = 100 Bpp \geq 95 WPbs = 98 Bpp \geq 95 MPbs = 95 WPbs = 94 MLbs = 77 Bpp = 1.00 MPbs = 100 NJbs = 100 NJbs = 100
O	Chaetosphaeriales				
O	Coniochaetales				
O	Diaporthales				

O		Miller & Huhndorf (2005, fig. 7) Miller & Huhndorf (2004, fig. 10) Zhang <i>et al.</i> (2007, fig. 2)	lsu, b-tub, rpb2 lsu lsu, ssu, rpb2, tef1	2 3 3	Bpp = 1.00 WPbs = 100 Bpp ≥ 95 WPbs = 100 Bpp = >95 MPbs = 100 WPbs = 100 MLbs = 100 Bpp = 1.00 N bs = 99 MPbs = 99 MPbs = 80 WPbs = 77 MLbs = 84 Bpp = 1.00 WPbs = <50 Bpp < 95 WPbs = 65 Bpp ≥ 95 MPbs = 98 WPbs = 99 MLbs = 78 Bpp = 1.00 MPbs = 92
O	Ophiostomatales	Hausner & Reid (2004, fig. 1) Wingfield <i>et al.</i> (1999 fig. 3) Zhang <i>et al.</i> (2007, fig. 2)	ssu lsu lsu, ssu, rpb2, tef1	3 4 17	
O	Sordariales	Huhndorf <i>et al.</i> (2004a, fig. 1)	lsu	22	
SC/O	Xylariomycetidae, Xylariales	Miller & Huhndorf (2005, fig. 7) Zhang <i>et al.</i> (2007, fig. 2)	lsu, β-tub, rpb2 lsu, ssu, rpb2, tef1	41 8	
O	Sordariomycetes incertae sedis (not placed in any subclass) Calosphaeriales	Shenoy <i>et al.</i> (2006, fig. 1) Vijaykrishna <i>et al.</i> (2004, fig. 1) Réblová <i>et al.</i> (2004, fig. 1) Réblová (2006, fig. 1) Zhang <i>et al.</i> (2007, fig. 2)	lsu ssu lsu ssu lsu, ssu, rpb2, tef1	16 3 6 2 2	
O	Lulworthiales (incl. Spathulosporales)	Campbell <i>et al.</i> (2005, fig. 1)	lsu, ssu	56	MPbs = 100 MPbs = 53 MPbs = 68 MPbs = 100 WPbs = 100 MLbs = 100 Bpp = 1.00 Bpp = 1.00

O		Inderbitzin <i>et al.</i> (2004, fig. 15)	lsu	15	MPbs = 100 NJbs = 91 Bpp = 86 MPbs = 100 MPbs = 100 MPbs < 50 NA MPbs < 50
O	Meliolales	Kohlmeyer <i>et al.</i> (2000, fig. 1)	lsu, ssu	7	
O	Phyllachorales	Saenz & Taylor. (1998, fig. 1)	lsu	2	
O	Trichosphaeriales	Vijaykrishna <i>et al.</i> (2004, fig. 1)	ssu	2	
O	Pezizomycotina incertae sedis (not placed in any class)	Inderbitzin <i>et al.</i> (2004, fig. 14)	ssu	1	NA
O	Lahmiales	Réblová & Seifert (2004, fig. 1)	lsu	8	MPbs < 50
O	Medeolariales	Eriksson (1986)	---	---	---
O	Triblidiales	Inderbitzin <i>et al.</i> (2004, fig. 14) Eriksson (1992)	ssu ---	1 ---	NA ---

Table 3. Support for major groups of Fungi in selected phylogenetic studies: Basidiomycota. See Table 1 for explanation.

Ran k	Taxon	Reference	Data	OTUs	Support
SK	<i>DIKARYA</i>	James <i>et al.</i> (2006a)	lsu, ssu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i>	161	Bpp = 1.00 MLbs = 71
P	<i>BASIDIOMYCOTA</i>	James <i>et al.</i> (2006a)	lsu, ssu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i>	50	Bpp = 1.00 MLbs = 80
SP	<i>Pucciniomycotina</i>	Matheny <i>et al.</i> (2007a, fig. 4)	ssu, lsu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i>	17	Bpp = 1.00 MPbs = 100
		Aime <i>et al.</i> (2007, fig. 2)	lsu, ssu	109	Bpp = 1.00 MPbs = 100
C	<i>Pucciniomycetes</i>	Matheny <i>et al.</i> (2007a, fig. 4)	lsu, ssu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i>	7	Bpp > 0.95 MPbs > 70
		Matheny <i>et al.</i> (2007a, fig. 5)	lsu, ssu, 5.8S	24	Bpp = 0.97 MPbs ≥ 70
		Aime <i>et al.</i> (2007, fig. 2)	lsu, ssu	19	Bpp = 1.00 MPbs = 100
O	<i>Septobasidiales</i>	Aime <i>et al.</i> (2007, fig. 3)	lsu, ssu	41	MPbs = 86
O	<i>Pachnocybales</i>	Arun Kumar <i>et al.</i> (2007, fig. 7)	lsu, ssu	4	Bpp = 1.0 MPbs = 100
O	<i>Helicobasidiales</i>	Bauer <i>et al.</i> (2006, fig. 1)	lsu	1	NA
		Berres <i>et al.</i> (1995, fig. 4)	lsu	1	NA
		Aime <i>et al.</i> (2007, fig. 2)	lsu, ssu	2	Bpp = 1.00 MPbs = 96
O	<i>Platyglloeales</i>	Aime <i>et al.</i> (2007, fig. 3)	lsu, ssu	10	Nlbs = 98 MPbs = 87
		Aime <i>et al.</i> (2007, fig. 2)	lsu, ssu	4	Bpp = 1.00 MPbs = 100
O	<i>Pucciniales</i>	Aime <i>et al.</i> (2007, fig. 3)	lsu, ssu	8	Nlbs = 100 MPbs = 99
		Matheny <i>et al.</i> (2007a, fig. 4)	lsu, ssu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i>	2	Bpp > 0.95 MPbs > 70
O		Aime <i>et al.</i> (2007, fig. 2)	lsu, ssu	12	Bpp = 1.00

C	<i>Cystobasidiomycetes</i>	<p>Aime (2006) Wingfield <i>et al.</i> (2004) Matheny <i>et al.</i> (2007a, fig. 4) Aime <i>et al.</i> (2007, fig. 2)</p>	<p>Isu ssu Isu, ssu, 5.8S, <i>rpb1</i>, <i>rpb2</i>, <i>tef1</i> Isu, ssu</p>	<p>46 72 5 27</p>	<p>MPbs = 100 NJbs = 100 MPbs = 99 MPbs < 50 Bpp > 0.95 MPbs > 70 Bpp = 1.00 MPbs = 100 NJbs = 96 Bpp = 0.92 Bpp = 0.98 MLbs = 100 Bpp = 1.00 MPbs = 100 NJbs = 100 Bpp = 1.00</p>
O	<i>Cystobasidiales</i>	<p>Sampaio (2004, fig. 1) Sampaio (2004, fig. 2) Nagahama <i>et al.</i> (2006, fig. 2) Aime <i>et al.</i> (2007, fig. 2)</p>	<p>Isu Isu Isu, ssu, 5.8S, <i>tef1</i> Isu, ssu</p>	<p>11 26 9 12</p>	<p>NJbs = 96 Bpp = 0.92 Bpp = 0.98 MLbs = 100 Bpp = 1.00 MPbs = 100 NJbs = 100 Bpp = 1.00</p>
O	<i>Erythrobasidiales</i>	<p>Sampaio (2004, fig. 2) Nagahama <i>et al.</i> (2006, fig. 2) Aime <i>et al.</i> (2007, fig. 2)</p>	<p>Isu Isu, ssu, 5.8S, <i>tef1</i> Isu, ssu</p>	<p>8 21 14</p>	<p>Bpp = 1.00 MLbs = 72 Bpp = 1.00 MPbs = 83 NJbs = 91 Bpp = 1.00</p>
O	<i>Naohideales</i>	<p>Sampaio (2004, fig. 2) Aime <i>et al.</i> (2007, fig. 3) Weiß <i>et al.</i> (2004)</p>	<p>Isu Isu, ssu Isu</p>	<p>18 2 3</p>	<p>NJbs = 91 Bpp = 1.00 MPbs = 98 Bpp = 0.94 NJbs < 50 Bpp = 1.00 MPbs < 70 NJbs < 70 NJbs = 89 Bpp = 1.00 MPbs > 70 Bpp = 1.0 MPbs = 100 NJbs = 100 MPbs = 98</p>
C	<i>Agaricostilbomycetes</i>	<p>Aime <i>et al.</i> (2007, fig. 2)</p>	<p>Isu, ssu</p>	<p>25</p>	<p>NJbs < 50 Bpp = 1.00 MPbs < 70 NJbs < 70 NJbs = 89 Bpp = 1.00 MPbs > 70 Bpp = 1.0 MPbs = 100 NJbs = 100 MPbs = 98</p>
O	<i>Agaricostilbales</i>	<p>Bauer <i>et al.</i> (2006, fig. 2) Matheny <i>et al.</i> (2007a, fig. 5) Aime <i>et al.</i> (2007, fig. 2)</p>	<p>Isu, ssu Isu, ssu, 5.8S Isu, ssu</p>	<p>4 8 22</p>	<p>NJbs < 70 NJbs = 89 Bpp = 1.00 MPbs > 70 Bpp = 1.0 MPbs = 100 NJbs = 100 MPbs = 98</p>
		<p>Aime <i>et al.</i> (2007, fig. 2)</p>	<p>Isu, ssu</p>	<p>34</p>	<p>MPbs = 100 NJbs = 100 MPbs = 99 MPbs < 50 Bpp > 0.95 MPbs > 70 Bpp = 1.00 MPbs = 100 NJbs = 96 Bpp = 0.92 Bpp = 0.98 MLbs = 100 Bpp = 1.00 MPbs = 100 NJbs = 100 Bpp = 1.00</p>

O	<i>Spiculogloiales</i>	Sampaio (2004, fig. 1) Sampaio (2004, fig. 2) Fell <i>et al.</i> (2001) Aime <i>et al.</i> (2007, fig. 2)	lsu lsu lsu lsu, ssu	7 23 24 3	Bpp = 1.00 Bpp = 1.00 MPbs = 64 Bpp = 1.00 MPbs = 100 NJbs = 100 MPbs = 74 NJbs = 90 Bpp = 1.0 MPbs = 100 NJbs = 100 MPbs = 74 Bpp > 0.95 MPbs > 70 Bpp = 0.87 MPbs = 75
C	<i>Microbotryomycetes</i>	Aime <i>et al.</i> (2007, fig. 3) Bauer <i>et al.</i> (2006, fig. 2) Aime <i>et al.</i> (2007, fig. 2) Aime <i>et al.</i> (2007, fig. 3) Matheny <i>et al.</i> (2007a, fig. 4)	lsu, ssu lsu, ssu lsu, ssu lsu, ssu lsu, ssu lsu, ssu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tefl</i> lsu lsu	7 2 31 60 6 49* 78	MPbs = 100 NJbs = 74 Bpp = 0.98 MPbs = 85 NJbs = 100 MPbs = 67 Bpp = 1.00 MPbs = 74 NJbs = 68 MPbs = 69 Bpp = 0.98 Bpp = 1.00 MPbs = 80 NJbs = 96 MPbs = 68
O	<i>Heterogastridiales</i>	Sampaio (2004, fig. 2) Fell <i>et al.</i> (2001)	lsu, ssu lsu, ssu	1 4	NA Bpp = 1.0 MPbs = 99
O	<i>Microbotryales</i>	Bauer <i>et al.</i> (2006, fig. 2) Aime <i>et al.</i> (2007, fig. 2)	lsu, ssu lsu, ssu	12 3	NJbs = 94 MPbs = 82 Bpp = 0.98 MPbs = 85
O	<i>Leucosporidiales</i>	Aime <i>et al.</i> (2007, fig. 3) Aime <i>et al.</i> (2007, fig. 2)	lsu, ssu lsu, ssu	9 13	NJbs = 100 MPbs = 67 Bpp = 1.00 MPbs = 74
O	<i>Sporidiobolales</i>	Aime <i>et al.</i> (2007, fig. 3) Aime <i>et al.</i> (2007, fig. 2)	lsu, ssu lsu, ssu	17 20	NJbs = 68 MPbs = 69 Bpp = 0.98
C/O	<i>Atractiellomycetes</i> , <i>Atractiellales</i>	Sampaio (2004, fig. 2) Aime <i>et al.</i> (2007, fig. 2) Aime <i>et al.</i> (2007, fig. 3)	lsu, ssu lsu lsu, ssu lsu, ssu	4 8	Bpp = 1.00 MPbs = 80 NJbs = 96 MPbs = 68

C/O	Classiculomycetes, Classiculales	Bauer <i>et al.</i> (2006, fig. 2) Aime <i>et al.</i> (2007, fig. 2)	Isu, ssu Isu, ssu	7	NJbs = 68 Bpp = 1.00 MPbs = 100 NJbs = 100 Bpp = 1.00 NJbs = 99
C/O	Mixiomycetes, Mixiales	Weiß <i>et al.</i> (2004, figs. 1-2)	Isu	2	
C/O	Cryptomycolacomycetes, Cryptomycolacales	Aime <i>et al.</i> (2007, fig. 2) Bauer <i>et al.</i> (2006, fig. 2) Aime <i>et al.</i> (2007, fig. 3)	Isu, ssu Isu, ssu Isu, ssu	1 1 1	NA NA NA
SP	Ustilaginomycotina	Bauer <i>et al.</i> (2006, fig. 1) Matheny <i>et al.</i> (2007a, fig. 4)	Isu Isu, ssu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i>	2 24	NJbs = 100 Bpp = 1.00 MPbs = 100
C	Ustilaginomycetes	Matheny <i>et al.</i> (2007a, fig. 5) Bauer <i>et al.</i> (2006, fig. 2) Matheny <i>et al.</i> (2007a, fig. 4) Matheny <i>et al.</i> (2007a, fig. 5)	Isu, ssu Isu, ssu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i> Isu, ssu, 5.8S	59 21 12 25	Bpp = 1.00 MPbs > 70 NJbs = 100 Bpp > 0.95 MPbs > 70 Bpp = 1.00 MPbs > 70
		Begerow <i>et al.</i> (2007, fig. 1)	Isu, ITS, <i>atp6</i> , <i>βtub</i>	53	Bpp = 1.00 MPbs = 83 NJbs = 77
		Bauer <i>et al.</i> (2001, figs. 33-34)	Isu	36	MPbs = 79 NJbs = 93
O	Urocystales	Fell <i>et al.</i> (2001, fig. 24) Begerow <i>et al.</i> (2007, fig. 1)	Isu Isu, ITS, <i>atp6</i> , <i>βtub</i>	27 5	NJbs = 86 Bpp = 1.00 MPbs = 66 NJbs = 96
		Matheny <i>et al.</i> (2007a, fig. 4) Bauer <i>et al.</i> (2001, figs. 33-34)	Isu, ssu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i> Isu	1 9	NA MPbs = 95 ³ NJbs = 96 ³

O	<i>Ustilaginales</i>	Matheny <i>et al.</i> (2007a, fig. 4)	Isu, ssu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i>	10	Bpp > 0.95 MPbs > 70
		Matheny <i>et al.</i> (2007a, fig. 5)	Isu, ssu, 5.8S	23	Bpp > 0.95 MPbs > 70
		Begerow <i>et al.</i> (2007, fig. 1)	Isu, ITS, atp6, β tub	46	Bpp = 1.00 MPbs < 60 NJbs < 60
C	<i>Exobasidiomycetes</i>	Matheny <i>et al.</i> (2007a, fig. 4)	Isu, ssu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i>	12	Bpp > 0.95 MPbs < 50
		Begerow <i>et al.</i> (2007, fig. 1)	Isu, ITS, atp6, β tub	35	Bpp < 0.60 MPbs < 60 NJbs < 60
		Bauer <i>et al.</i> (2001, figs. 33-34)	Isu	36	MPbs = 85 NJbs = 56
O	<i>Doassansiales</i>	Matheny <i>et al.</i> (2007a, fig. 4)	Isu, ssu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i>	1	NA
		Matheny <i>et al.</i> (2007a, fig. 5)	Isu, ssu, 5.8S	4	Bpp > 0.95 MPbs > 70
		Begerow <i>et al.</i> (2007, fig. 1)	Isu, ITS, atp6, β tub	4	Bpp = 1.00 MPbs = 84 NJbs = 77
		Bauer <i>et al.</i> (2001, figs. 33-34)	Isu	5	MPbs = 96 NJbs = 97
O	<i>Entylomatales</i>	Matheny <i>et al.</i> (2007a, fig. 4)	Isu, ssu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i>	4	Bpp > 0.95 MPbs > 70
		Begerow <i>et al.</i> (2007, fig. 1)	Isu, ITS, atp6, β tub	3	Bpp = 1.00 MPbs < 60 NJbs < 60
		Bauer <i>et al.</i> (2001, figs. 33-34)	Isu	9	MPbs = 72 NJbs = 91
O	<i>Exobasidiales</i>	Matheny <i>et al.</i> (2007a, fig. 4)	Isu, ssu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i>	2	Bpp > 0.95 MPbs > 70
		Matheny <i>et al.</i> (2007a, fig. 5)	Isu, ssu	6	Bpp > 0.95 MPbs > 70
		Begerow <i>et al.</i> (2007, fig. 1)	Isu, ITS, atp6, β tub	8	Bpp = 1.00

O	Geogfischeriales	<p>Matheny <i>et al.</i> (2007a, fig. 4)</p> <p>Begerow <i>et al.</i> (2007, fig. 1)</p> <p>Bauer <i>et al.</i> (2001, figs. 33-34)</p> <p>Matheny <i>et al.</i> (2007a, fig. 4)</p> <p>Matheny <i>et al.</i> (2007a, fig. 5)</p> <p>Begerow <i>et al.</i> (2007, fig. 1)</p>	<p>Isu, ssu, 5.8S, <i>rpb1</i>, <i>rpb2</i>, <i>tef1</i></p> <p>Isu, ITS, atp6, β<i>tub</i></p> <p>Isu</p> <p>Isu, ssu, 5.8S, <i>rpb1</i>, <i>rpb2</i>, <i>tef1</i></p> <p>Isu, ssu, 5.8S</p> <p>Isu, ITS, atp6, β<i>tub</i></p>	<p>2</p> <p>5</p> <p>9</p> <p>1</p> <p>3</p> <p>5</p> <p>2</p> <p>7</p> <p>5</p>	<p>MPbs < 60</p> <p>NJbs = 61</p> <p>Bpp > 0.95</p> <p>MPbs > 70</p> <p>Bpp < 0.60</p> <p>MPbs < 60</p> <p>NJbs < 60</p> <p>MPbs = 86</p> <p>NJbs = 65</p> <p>NA</p>
O	Microstromatales				
O	Tilletiales				
O	<i>Ustilaginomycotina incertae sedis</i> (not placed in any class)				
O	Malasseziales	<p>Matheny <i>et al.</i> (2007a, fig. 4)</p> <p>Begerow <i>et al.</i> (2007, fig. 1)</p>	<p>Isu, ssu, 5.8S, <i>rpb1</i>, <i>rpb2</i>, <i>tef1</i></p> <p>Isu, ITS, atp6, β<i>tub</i></p>	<p>1</p> <p>2</p>	<p>NA</p> <p>Bpp = 1.00</p> <p>MPbs = 100</p> <p>NJbs = 100</p> <p>MPbs = 100</p> <p>NJbs = 100</p>
SP	Agaricomycotina	<p>Bauer <i>et al.</i> (2001, figs. 33-34)</p> <p>Matheny <i>et al.</i> (2007b, fig. 6)</p>	<p>Isu</p> <p>Isu, ssu, 5.8S, <i>rpb2</i>, <i>tef1</i></p>	<p>4</p> <p>125</p>	<p>Bpp = 1.00</p> <p>MPbs = 100</p> <p>NJbs = 100</p> <p>MPbs = 100</p> <p>NJbs = 100</p> <p>Bpp = 1.00</p> <p>MPbs = 95</p>
C	Tremellomycetes	<p>Matheny <i>et al.</i> (2007a, fig. 4)</p>	<p>ssu, Isu, 5.8S, <i>rpb1</i>,</p>	<p>5</p>	<p>Bpp > 0.95</p>

O	Cystofilobasidiales	Fell <i>et al.</i> (2001, figs. 19, 22) Matheny <i>et al.</i> (2007a, fig. 5)	<i>rpb2, tef1</i> lsu lsu, ssu, 5.8S	139 5 16 34 5 89 4 9	MPbs = 50-69 MPbs = 100 Bpp = 1.00 MPbs ≥ 70 Bpp = 1.00 MPbs = 83 MPbs = 96 Bpp ≥ 0.95 MPbs ≥ 70 MPbs = 56 Bpp = 1.00 MPbs = 100 NJbs = 99
O	Filobasidiales	Fell & Scorzetti (2004, fig. 1)	lsu		
O	Tremellales	Fell <i>et al.</i> (2001, figs. 19, 22) Matheny <i>et al.</i> (2007a, fig. 5)	lsu lsu, ssu, 5.8S		
C/O	Dacrymycetes, Dacrymycetales	Fell <i>et al.</i> (2001, figs. 19, 22) Matheny <i>et al.</i> (2007b, fig. 6) Weiß & Oberwinkler (2001, fig. 6)	lsu lsu, ssu, 5.8S, <i>rpb2, tef1</i> lsu		
C	Agaricomycetes	Matheny <i>et al.</i> (2007b, fig. 6)	lsu, ssu, 5.8S, <i>rpb2, tef1</i>	119	Bpp = 1.00 MPbs = 95
SC	Agaricomycetidae	James <i>et al.</i> (2006a) Matheny <i>et al.</i> (2007b, fig. 6)	lsu, ssu, 5.8S, <i>rpb1, rpb2, tef1</i> lsu, ssu, 5.8S, <i>rpb2, tef1</i>	37 63	Bpp = 1.00 MLbs = 92 Bpp = 1.00 MPbs = 96
O	Agaricales	Binder & Hibbett (2007, fig. 2) Binder <i>et al.</i> (2005, fig. 1) Matheny <i>et al.</i> (2006, fig. 2) Matheny <i>et al.</i> (2006, fig. 3) Matheny <i>et al.</i> (2007b, fig. 6) Moncalvo <i>et al.</i> (2002, fig. 2) Larsson <i>et al.</i> (2004, fig. 1) Binder <i>et al.</i> (2005, fig. 4) Matheny <i>et al.</i> (2007b, fig. 6)	lsu, ssu, 5.8S, mt- lsu, atp6 lsu, ssu, mt-lsu, mt- ssu lsu, ssu, 5.8S lsu, ssu, 5.8S, <i>rpb1, rpb2</i> lsu, ssu, 5.8S, <i>rpb2, tef1</i> lsu lsu lsu, ssu, mt-lsu, mt- ssu	47 46 230 238 41 786 8 3 11	Bpp > 0.98 MLbs = 88 MPbs = 62 Bpp = 0.84 Bpp = 1.00 MPbs = 43 Bpp = 1.00 MPbs = 76 MPbs < 50 MPbs = 97 MPbs = 75 Bpp = 1.00 MPbs = 100
O	Atheliales				
O	Boletales				

SC	<i>Phallomycetidae</i>	Binder & Hibbett (2007, fig. 2)	lsu, ssu, 5.8S, mt-lsu, atp6 lsu lsu	42	Bpp > 0.98 MLbs = 99 Bpp > 0.98 Bpp = 1.00 MPbs = 98 Bpp = 1.00 MPbs = 100
O	<i>Geastrales</i>	Hosaka <i>et al.</i> (2007b, fig. 6)	lsu, mt-ssu, atp6, <i>rpb2</i> , <i>tef1</i> lsu, ssu, 5.8S, <i>rpb2</i> , <i>tef1</i>	3	Bpp = 1.00 MPbs = 100
O	<i>Gomphales</i>	Hosaka <i>et al.</i> (2007, fig. 2)	lsu, mt-ssu, atp6, <i>rpb2</i> , <i>tef1</i>	21	Bpp = 1.00 MPbs = 59
O	<i>Hysterangiales</i>	Hosaka <i>et al.</i> (2007, fig. 2)	lsu, mt-ssu, atp6, <i>rpb2</i> , <i>tef1</i>	61	Bpp = 1.00 MPbs = 63
O	<i>Phallales</i>	Hosaka <i>et al.</i> (2007, fig. 2)	lsu, mt-ssu, atp6, <i>rpb2</i> , <i>tef1</i>	99	Bpp = 1.00 MPbs = 98
O	<i>Agaricomycetes incertae sedis</i> (not placed in any subclass): <i>Auriculariales</i>	Hosaka <i>et al.</i> (2007, fig. 2)	lsu, mt-ssu, atp6, <i>rpb2</i> , <i>tef1</i>	41	Bpp = 1.00 MPbs = 84
O	<i>Cantharellales</i>	Matheny <i>et al.</i> (2007b, fig. 6)	lsu, ssu, 5.8S, <i>rpb2</i> , <i>tef1</i> lsu	3	Bpp = 1.00 MPbs = 100 NJbs < 60
O		Weiß & Oberwinkler (2001, fig. 6)		43	
O		Matheny <i>et al.</i> (2007b, fig. 6)	lsu, ssu, 5.8S, <i>rpb2</i> , <i>tef1</i>	11	Bpp = 1.00 MPbs = 69
O		Moncalvo <i>et al.</i> (2007, fig. 1)	lsu, ssu, mt-ssu, <i>rpb2</i>	29	Bpp < 0.50 MPbs < 50 MPbs < 50
O		Binder <i>et al.</i> (2005, fig. 4)	lsu, ssu, mt-lsu, mt-ssu	31	MPbs < 50
O	<i>Corticiales</i>	Larsson <i>et al.</i> (2004, fig. 1)	lsu	7	MPbs = 96
O		Binder <i>et al.</i> (2005, fig. 4)	lsu, ssu, mt-lsu, mt-ssu	8	MPbs = 81
O	<i>Gloeophyllales</i>	Thorn <i>et al.</i> (2000, fig. 5)	lsu	5	MPbs = 71
O		Binder <i>et al.</i> (2005, fig. 4)	lsu, ssu, mt-lsu, mt-ssu	6	MPbs = 54
O	<i>Hymenochaetales</i>	Matheny <i>et al.</i> (2007b, fig. 6)	lsu, ssu, 5.8S, <i>rpb2</i> , <i>tef1</i>	7	Bpp = 1.00 MPbs = 63

O	Polyporales	Larsson <i>et al.</i> (2007, fig. 3) Wagner & Fischer (2002, fig. 2) Matheny <i>et al.</i> (2007b, fig. 6)	lsu, 5.8S lsu lsu, ssu, 5.8S, <i>rpb2</i> , <i>tef1</i> lsu, ssu, mt-lsu, mt- ssu	174 104 16 122	Bpp = 1.00 NJbs = 100 Bpp = 1.00 MPbs = 85 MPbs < 50
O	Russulales	Matheny <i>et al.</i> (2007b, fig. 6)	lsu, ssu, 5.8S, <i>rpb2</i> , <i>tef1</i>	8	Bpp = 1.00 MPbs = 99
O	Sebacinales	Larsson & Larsson (2003, fig. 1) Miller <i>et al.</i> (2007, fig. 2) Matheny <i>et al.</i> (2007b, fig. 6)	lsu, 5.8S lsu, ITS lsu, ssu, 5.8S, <i>rpb2</i> , <i>tef1</i> lsu	127 143 2 9	MPbs = 96 MPbs = 100 Bpp = 1.00 MPbs = 100 NJbs = 99
O	Thelephorales	Weiß & Oberwinkler (2001, fig. 6) Matheny <i>et al.</i> (2007b, fig. 6)	lsu, ssu, 5.8S, <i>rpb2</i> , <i>tef1</i>	2	Bpp = 1.00 MPbs = 100
O	Trechisporales	Binder <i>et al.</i> (2005, fig. 4) Larsson <i>et al.</i> (2004, fig. 1) Matheny <i>et al.</i> (2007b, fig. 6)	lsu, ssu, mt-lsu, mt- ssu lsu, 5.8S lsu, ssu, 5.8S, <i>rpb2</i> , <i>tef1</i>	13 11 2	MPbs = 97 MPbs = 86 Bpp = 1.00 MPbs = 100
C/O	Basidiomycota incertae sedis (not placed in any subphylum): Wallemiomycetes, Wallemiales	Binder <i>et al.</i> (2005, fig. 4) Larsson <i>et al.</i> (2004, fig. 1)	lsu, ssu, mt-lsu, mt- ssu lsu, 5.8S	20 12	MPbs = 69 MPbs = 99
C/O	Entorrhizomycetes, Entorrhizales	Matheny <i>et al.</i> (2007a, fig. 4) Matheny <i>et al.</i> (2007a, fig. 5) Matheny <i>et al.</i> (2007a, fig. 5) Bauer <i>et al.</i> (2001, figs. 33-34)	lsu, ssu, 5.8S, <i>rpb1</i> , <i>rpb2</i> , <i>tef1</i> lsu, ssu, 5.8S lsu, ssu, 5.8S lsu	3 3 3 2	Bpp > 0.95 MPbs > 70 Bpp = 1.00 MPbs > 70 Bpp = 1.00 MPbs > 70 NJbs = 100