DOI 10.1007/5584_2015_110

© Springer International Publishing Switzerland 2015

Published online: 19 March 2015

Pathophysiology of Clinical Symptoms in Acute Viral Respiratory Tract Infections

E. Kuchar, K. Miśkiewicz, Aneta Nitsch-Osuch, and L. Szenborn

Abstract

In this article we discuss the pathophysiology of common symptoms of acute viral respiratory infections (e.g., sneezing, nasal discharge, sore throat, cough, muscle pains, malaise, and mood changes). Since clinical symptoms are not sufficient to determine the etiology of viral respiratory tract infections, we believe that the host defense mechanisms are critical for the symptomatology. Consequently, this review of literature is focused on the pathophysiology of respiratory symptoms regardless of their etiology. We assume that despite a high prevalence of symptoms of respiratory infection, their pathogenesis is not widely known. A better understanding of the symptoms' pathogenesis could improve the quality of care for patients with respiratory tract infections.

Keywords

Common cold • Coronavirus • Flu • Influenza • Rhinovirus • RSV

E. Kuchar (⊠)

Department of Pediatric Infectious Diseases, Wroclaw Medical University, 2A Chalubinskiego Str., 50-368 Wroclaw, Poland

Department of Pediatrics and Observation Ward Medical University of Warsaw, 61 Żwirki i Wigury Str., 02-091 Wrocław, Poland

e-mail: ernest.kuchar@gmail.com

K. Miśkiewicz and L. Szenborn Department of Pediatric Infectious Diseases, Wrocław Medical University, 2A Chalubinskiego Str., 50-368

Wroclaw, Poland

A. Nitsch-Osuch Department of Family Medicine, Medical University of Warsaw, Warsaw, Poland

1 Introduction

1.1 Respiratory Tract Infections

The literature reports that each year up to 25 million people in the US visit their doctor because of respiratory tract infections. The upper respiratory tract infections, better known as common colds, are the most common clinical presentation of said infections (Gonzales et al. 2001). Viral lower respiratory tract infections, bronchitis, bronchiolitis, and pneumonia; e.g., with respiratory syncytial virus (RSV) or influenza are generally more common in infants, young children, and in patients with chronic medical conditions, whereas older children and healthy adults usually suffer from

milder upper respiratory tract infections (Eccles 2005). Common cold was the third most common diagnosis made during ambulatory care visits by patients of all ages in the US (Hsiao et al. 2010). It has been estimated that an average adult suffers from 2 to 4 colds per year and a schoolchild suffers from 6 to 10 colds per year (Spector 1995; Johnston and Holgate 1996; Winther et al. 1998; Eccles 2005). Therefore, respiratory tract infections represent a significant clinical and therapeutic problem, and an economic burden (Wat 2004). Upper respiratory tract infections are generally mild, selflimiting, and viral in their origin (Johnston and Holgate 1996; Snow et al. 2001; Turner 2011; Kennedy et al. 2012). In experimental studies, the infections have been defined as a short mild illness, with the early symptoms being headaches, chills, sneezing, and a sore throat, and delayed symptoms being nasal obstruction or discharge, cough, and malaise (Jackson et al. 1958). The duration of symptoms varies from 7 to 10 days, with a peak occurring on the 2-3rd day. However, some symptoms may be observed up to 3 weeks after the onset of the infection (Heikkinen and Järvinen 2003; Eccles 2005). Common colds are triggered by rhinovirus (RV) coronavirus, influenza and parainfluenza viruses, and RSV (Wat 2004; Eccles 2005; Kennedy et al. 2012). The diagnosis of upper respiratory tract infections is based on symptoms and, with the exception of antivirals which may be used in influenza, treatment remains symptomatic. Studies on different viruses responsible for the upper respiratory tract infections have shown that it is not possible to identify the virus based on the symptoms (Johnston and Holgate 1996; Eccles 2002). Not only is the clinical presentation of these infections similar regardless of their etiology, but the order in which the symptoms develop is also similar. If the etiology of infections cannot be determined on the basis of clinical symptoms, the host reaction must play a major role in their pathogenesis. The similarities in the clinical presentation of viral upper respiratory tract infections stems from the same immune response pattern to different etiologic agents (Eccles 2005). Furthermore, in acute respiratory tract infections a routine diagnosis to determine the etiology is not usually carried out in primary care settings.

Advances in molecular biology help explain the mechanisms that generate the symptoms of viral respiratory tract infections. Nevertheless, the practical use of the pathophysiology of common symptoms seems relatively poor compared with the amount of scientific knowledge available.

1.2 Symptomatology of Respiratory Tract Infections

Clinical manifestations of respiratory tract infections are familiar and well-known (Eccles 2005; Turner 2011). Although the symptomatology depends, to some extent, on the type and location (e.g., pharyngitis, rhinitis, sinusitis, and bronchitis), the etiology of respiratory infection (viral or bacterial), patient's age, general health, co-morbidities, immunity, and on whether the infection is primary or secondary, e.g., in RSV or influenza there is a great amount of variation and overlap in both etiology and symptoms of individual infections. Consequently, even defining the exact syndrome, like common cold or influenza-like morbidity, is difficult and problematic (Eccles 2005).

The most significant signs of viral respiratory tract infections include sneezing, rhinorrhea nose and nasal discharge), congestion, cough, tachypnea, and fever. Subjective symptoms include a sore throat, malaise, shivering (chills), shortness of breath, muscle aches and weakness, fatigue, and a loss of appetite and headaches (Snow et al. 2001; Wat 2004; Eccles 2005; Kennedy et al. 2012). Febrile seizures are a rare but important symptom in young children up to 6 years of age (Schuchmann et al. 2011). Symptoms of upper respiratory tract infections have been traditionally classified as early or late (Jackson et al. 1958; Eccles 2005). The early symptoms are those that develop quickly and resolve rapidly after 1–2 days, like headaches, sneezing, chills, sore throat, and malaise. In children a high fever may be observed, complicated by seizures in some cases (Monto et al. 2000; Schuchmann et al. 2011). Late symptoms include nasal discharge, nasal obstruction, and cough. The later symptoms develop over several days and are present one week after experimental infection (Jackson et al. 1958; Eccles 2005). The development of sneezing before coughing in patients with a common cold may be partly explained by the involvement of the upper airways first and the infection subsequent spread to the lower respiratory tract (Eccles 2005).

The aim of this review is to discuss the pathophysiology of symptoms of respiratory tract infections. We focused on the most significant symptoms of acute viral respiratory infections: sneezing, nasal discharge and obstruction, sore throat, coughing, muscle pains, malaise and mood changes, fever, and febrile seizures in children. We believe that despite a high prevalence of the symptoms, their pathogenesis is not widely known and a better understanding should have a beneficial effect on the therapeutic approach and should improve the quality of patient care.

2 Methods of Literature Selection

We searched the literature to identify relevant Medline, Scopus, Web of Science, Cochrane databases and respiratory-specific journals such as Respiratory Physiology & Neurobiology, American Journal of Respiratory and Critical Care Medicine, Thorax, Chest, Journal of Allergy, Asthma and Clinical Immunology, European Respiratory Journal, American Journal of Respiratory Cell and Molecular Biology, American Journal of Physiology – Lung Cellular and Molecular Physiology, BMC Pulmonary Medicine, Respiratory Research, Current Opinion in Pulmonary Medicine, Expert Review of Respiratory Medicine, Respiratory Research, Seminars in Respiratory, Critical Care Medicine, and Lancet Infectious Diseases were searched with the following key words: (Pathophys* or Pathogen*) and (Resp* symptom or Resp* infection or Common cold) from January 1990 through May 2014. If possible the results of the search were sorted according to 'relevance'. Due to a large number of articles, exceeding 20,000 (e.g., PubMed), only the first 250 were analyzed. References from relevant articles were also identified. Studies were included if they met the following two criteria: published in English and containing valid, consistent, and relevant data. The first two authors of the present review independently assessed all titles and abstracts to determine whether the inclusion criteria were satisfied. If either of the assessors considered the abstract potentially suitable, full-text articles were retrieved and then assessed by both assessors for their suitability for inclusion. Eventually, 129 publications were selected and studied by each of the authors before the manuscript was prepared.

3 Results

3.1 Significant Etiologic Viral Agents of Upper Respiratory Tract Infections

3.1.1 Rhinoviruses

Rhinoviruses are the most common etiologic agents of common cold. They are responsible for one-third to half of all upper respiratory tract infections reported in adults annually (Proud et al. 1990; Hendley 1998; Heikkinen and Järvinen 2003; Ruuskanen et al. 2013). Improved knowledge about the structure and function of rhinoviruses has been acquired in recent years using virus culture techniques and new molecular genetics methods available. Currently, more than 100 serotypes have been identified with HRV-A and HRV-B being the most important of them all (Heymann et al. 2005; Peltola et al. 2008; Bochkov et al. 2011; Kennedy et al. 2012). It has been demonstrated that the pathomechanism of symptoms in rhinoviral respiratory tract infections does not result from the cell damage, unlike influenza virus or RSV action (Winther et al. 1990). Rather, rhinovirus disrupts the tight junctions of the epithelial barrier, which facilitates the translocation of pathogens and stimulates the host's innate and adaptive immune responses (Rezaee et al. 2011; Kennedy et al. 2012). Over 90 % of rhinovirus serotypes enter the nasal epithelial cells through the host receptor with the intercellular adhesion molecule-1 (ICAM-1) being a glycoprotein immunoglobulin expressed on non-ciliated epithelial cells and on the basal surface of the

ciliated epithelium of nasopharyngeal mucosa, while only around ten rhinovirus serotypes use a minor group of receptors, low-density lipoprotein (LDL) (Bardin et al. 1994; Winther et al. 1997; Whiteman et al. 2003; Bella and Rossmann 2000; Vlasak et al. 2005; Kennedy et al. 2012). Newly discovered and sequenced human rhinovirus-C (HRV-C) is somehow unique as the virus isolates do not grow in typically used cell cultures, e.g., embryonic lung fibroblasts. In vitro growth of HRV-C was successfully performed using the sinus mucosal tissue as a substrate (Bochkov et al. 2011; Kennedy et al. 2012). Furthermore, studies on the structure and function of HRV-C have shown that the virus enters the cells using neither the ICAM-1 nor LDL receptor and the pathomechanism of the HRV-C infection remains unclear (Arden and Mackay 2010; Simmonds et al. 2010; Bochkov and Gern 2012; Lee et al. 2012; Ruuskanen et al. 2013).

In otherwise healthy individuals, rhinovirus infections are generally limited to the upper respiratory tract with rhinorrhea and nasal obstruction being the most prominent symptoms (Kennedy et al. 2012). Entry of the rhinovirus triggers the release of interleukin-8 (IL-8) a major cytokine in the recruitment of polymorphonuclear cells (Hendley 1998). IL-8, which is produced locally, increases the production of nasal secretions and causes an influx of neutrophils (Douglass et al. 1994). Oxidative stress caused by viral infection is probably responsible for the cellular mechanisms that lead to the production and release of IL-8 (Zhu et al. 1997). Studies of volunteers infected with a rhinovirus show a local infection in a small proportion (1–2 %) of nasopharyngeal epithelial cells (Turner et al. 1982; Bardin et al. 1994; Arruda et al. 1995). The higher the nasopharyngeal viral load the more severe is the disease (Esposito et al. 2014). Vasoactive kinin peptides are other important mediators produced on-site in nasal mucosa and submucosa in rhinovirus-infected humans. Kinins released in the nose following plasma exudation increase the symptoms of rhinoviral infection and cause an increase in vascular permeability, vasodilatation, and glandular secretion. Bradykinin applied to the nasal mucosa causes symptoms that mimic the common cold, including rhinitis, nasal obstruction, and sore throat (Proud et al. 1988, 1990). Symptom scores correlate with an increase in the kinin concentration. While nasal secretion in adults with symptomatic experimentally-induced rhinovirus infection contain significantly higher concentrations of bradykinin and lysyl-bradykinin, asymptomatic infections do not result in increased kinin concentrations (Naclerio et al. 1988). Interestingly, the presence of rhinovirus has been detected by RT-PCR in 20 % of asymptomatic people who had an infected family member (Jartti et al. 2008).

3.1.2 Coronaviruses

The coronavirus (CoV) is the second etiologic agent of upper respiratory tract infections (Eccles 2005; Mesel-Lemoine et al. 2012). The most common human-infecting coronaviruses include 229E, OC43, SARS-CoV, and the recently discovered NL63 and HKU1 (Arden et al. 2005; Esper et al. 2005; Vabret et al. 2005; Pyrc et al. 2007). The virus is transmitted by aerosol inhalation and reinfections often occur due to a short-lived immunity (Callow et al. 1990; Wat 2004). As a result, coronavirus accounts for 15-30 % of upper respiratory tract infections in humans. These infections are limited predominantly to the upper respiratory tract and rarely spread to lower airways and lungs (Mesel-Lemoine et al. 2012). The coronavirus infection can occasionally involve other organs (Arbour al. 2000: Collins 2002: et Desforges et al. 2006). It has been reported that the two new members of the coronavirus family, NL63 and HKU1, especially the NL63, could also trigger severe lower respiratory tract infections and abdominal disorders such as pain and diarrhea (Arden et al. 2005; Esper et al. 2005; Vabret et al. 2005; Pyrc et al. 2007). Epidemics caused by CoV-NL63 and CoV-HKU1 have been observed every 2–3 years (Kahn 2006; Jartti et al. 2012). Studies have confirmed that the infection with CoV-NL63 is associated with croup and acute respiratory disease mostly in children, the elderly, and patients with chronic diseases, with some fatal cases being reported (van der Hoek et al. 2005; Han et al. 2007; Wu et al. 2008; Oosterhof et al. 2010; Sung et al. 2010; Milewska et al. 2013). The growth of coronaviruses using standard tissue cultures is poor and advanced molecular methods are needed to isolate the virus, so that most infections may remain undiagnosed (Walsh et al. 2013). Human aminopeptidase N (hAPN), a zinc-binding protein with endopeptidase activity, is used by CoV-229E for entry into the epithelial cells, whereas CoV-OC43 hemagglutinin-esterase (HE) and spike (S) – its own surface glycoproteins - to enter the cell (Tyrrell et al. 1993; Künkel and Herrler 1996; Wat 2004). Recent studies have shown the ability of CoV-229E to destroy the dendritic cells, which are essential components of the respiratory tract's immune system, which may explain multiple reinfections with the same type of CoV (Mesel-Lemoine et al. 2012). Although the pathogenesis of infection caused by the two main groups of CoV - 229E and OC43 - is different, the clinical symptomatology is similar (Tyrrell et al. 1993; Wat 2004).

3.1.3 Respiratory Syncytial Virus

Respiratory syncytial virus (RSV) is responsible for many flu-like illnesses (Zambon et al. 2001). RSV replication starts in the nasopharynx and then the virus infects the bronchiolar epithelium presumably by cell-to-cell spread or aspiration of secretions. The virus spares the basal cells and subsequently extends the alveolar to pneumocytes. The pathologic findings of RSV are characterized by necrosis of epithelial cells, infiltration with T cells and monocytes around arterioles and with neutrophils between the vascular structures and small airways (Johnson et al. 2007). This leads to airway obstruction, air trapping, increased airway resistance, and is associated with neutrophilia bronchoalveolar lavage (Everard et al. 1994). RSV has never been isolated from blood (Peebles and Graham 2005). The immune response to RSV, especially cytokines and chemokines, seems to be responsible for the symptoms and severity of bronchiolitis (Garofalo et al. 2001; Legg et al. 2003). The cytokines IL-8, IL-6, TNF-alpha, and IL-1 beta were detected in respiratory secretions from infected children and high IL-6 concentrations are associated with severe manifestations of the disease (Matsuda et al. 1995; Noah et al. 1995; Smyth et al. 1997). Respiratory secretions from infected children contain chemokines expressed and secreted by T-cells (chemokine ligand 3 - CCL3, i.e. macrophage inflammatory protein-1 - MIP-1 alpha; chemokine ligand 2 - CCL2, i.e. monocyte chemoattractant protein-1 _ MCP-1; chemokine 11 - CCL11 - eotaxin, and chemokine ligand 5 - CCL5, i.e. RANTES; regulated on activation). MIP-1 alpha, and to a lesser extent other beta-chemokines, primarily secreted by activated immune cells, are associated with severe manifestations of the disease (Noah et al. 1995; Welliver et al. 2002; Garofalo et al. 2005). Experimental infection of explanted polarized respiratory epithelium in tissue culture generates IL-8 and CCL5 (Mellow et al. 2004). Nonetheless, it is unknown whether the cytokines and chemokines are the cause of disease or are by-products of enhanced inflammatory responses (Barr and Graham 2014).

3.1.4 Influenza Viruses

Whether respiratory tract infections caused by influenza viruses present as common colds with typical symptoms or as severe lower respiratory tract diseases depends on the type of virus, pre-existing immunity, the patient's underlying disorders, and multiple other factors (Wat 2004). The phenomena such as antigenic shift or drift have led to the formation of more recent and increasingly virulent variations of the influenza virus and, consequently, to more serious clinical manifestations (Gething et al. 1980; Treanor 2004). As an example, pandemic influenza A (H1N1)pdm09 affected all age groups, but it was more prevalent in younger patients and children in whom there was the highest rate of hospitalization and pneumonia (Kuchar et al. 2013). It has previously been shown that coughing and fever are the best predictive factors of influenza infections having a positive predictive value (PPV) of 79 % (Monto et al. 2000). However, neither symptom is sufficiently predictive in children aged 1–4 (Ohmit and Monto 2006). Overall, influenza viruses are generally responsible for 5–15 % of acute upper respiratory tract infections in humans (Wat 2004). The influenza virus causes damage to the epithelial cells and its replication occurs in the airways with predilection to the lower respiratory tract (Hers and Mulder 1961; Hers 1966; Wat 2004). The two main glycoproteins of the influenza virus surface - hemagglutinin (HA) and neuraminidase (NA) play an essential role in the infection spread. HA targets cells for infection binding to the epithelial sialylated glycans (specific for upper or lower airways) (Shinya et al. 2006; de Wit et al. 2010; Fukuyama and Kawaoka 2011), while NA is responsible for effective viral replication (Pappas et al. 2008). Another viral protein, nonstructural protein 1 (NS1) is also important due to its counteracting IFN- α production in infected cells (Fukuyama and Kawaoka 2011). Viral replication is possible in host cells due to activation of nuclear factor kappa B (NF-κB) and the Raf/MEK/ERK cascade, and then proinflammatory cytokines are produced with interleukin 6 (IL-6) being the most important of them (Kaiser et al. 2001; Pinto et al. 2011; Wine and Alper 2012). IL-6, tumor necrosis factor- α (TNF- α), interferon- α (IFN- α), IL-8, and IL-1 β increase significantly in response to the viral invasion resulting in the development of fever, nasopharyngeal mucous production, and respiratory and systemic symptoms. Viral replication and the intensity of the main influenza symptoms are correlated with the level of cytokines, particularly with IL-6 and TNF- α (Hayden et al. 1998; Kaiser et al. 2001).

4 Pathophysiology of Common Respiratory Signs and Symptoms

Evidence presented in our review of the pathophysiology of signs and symptoms in the four most common viral upper respiratory tract infections suggest that the immune system's response to infection rather than the virusspecific damage to the respiratory tract is responsible for the symptomatology (Turner 1997; Hendley 1998; Eccles 2005). Studies on different viruses responsible for upper respiratory tract infections have shown that it is not possible to identify the virus based on the symptoms (Eccles 2005). The pathology of rhinovirus infections consists of the influx of polymorphonuclear leukocytes at the beginning of the infection (Winther et al. 1984). Macrophages play a key role in triggering an acute phase response with the production of cytokines (Beutler 2003), while release the proinflammatory cytokines and other mediators cause upper respiratory tract infection symptoms (Eccles 2000a, b). Cytokines are responsible for the systemic symptoms (e.g., fever) and bradykinin plays a major role in local symptoms of respiratory tract infections (e.g., sore throat and nasal congestion) (Proud et al. 1988; Shibayama et al. 1996; Conti et al. 2004).

4.1 Sore Throat

A sore throat, irritation, and pain in the pharynx usually appear at the beginning of a respiratory tract infection. A sore throat is most likely caused by the action of prostaglandins and bradykinin on sensory nerve endings in the upper respiratory tract. Intranasal administration of bradykinin causes symptoms of rhinitis and a sore throat, so it is likely to be responsible for these symptoms (Rees and Eccles 1994; Proud 1998). The sensation of pain is mediated by the cranial nerves supplying the nasopharynx. Similar symptoms have been observed in bacterial upper respiratory tract infections, pharyngitis, and tonsillitis (Georgitis 1993).

4.2 Nasal Congestion

Nasal congestion is a subsequent symptom of respiratory infection that develops over the first week of symptoms (Tyrrell et al. 1993). The mechanism of nasal congestion relies on the dilation of the venous sinuses in the nasal epithelium in response to the vasodilator mediators such as

bradykinin (Proud et al. 1990; Widdicombe 1997). Symptom scores increase as kinin concentrations rise (Proud et al. 1990). Dilatation of the sinuses in the narrow nasal valve region causes obstruction of the nasal airway (Eccles 2000b). The so-called nasal cycle (alternating congestion and decongestion of the nasal passages controlled by the sympathetic vasoconstrictor nerves) is accentuated and the asymmetry of nasal airflow is more pronounced during respiratory infection (Eccles et al. 1996).

4.3 Rhinorrhea

A watery nasal secretion often accompanied by sneezing is an early symptom of a respiratory tract infection. Nasal discharge in respiratory infections is a complex mixture of plasma and glandular exudates with cellular elements (e.g., goblet cells, plasma cells, and neutrophils) of variable composition that changes over the course of the infection and severity of the inflammatory response (Eccles 1983). The first phase of nasal discharge consists of a glandular secretion reflex caused by stimulation of the upper airway's trigeminal nerves. Studies demonstrated that intranasal administration of ipratropium inhibits nasal secretions in the first 4 days of a common cold (e.g., when it is caused by coronavirus) (Akerlund et al. 1993; Hayden et al. 1996). The color of the nasal discharge may change from watery clear to yellow and green during the course of the respiratory tract infection and this reflects the severity of the inflammatory response rather than the etiology of the infection (Stockley et al. 2001). The green or yellow color of nasal discharge is often regarded as a clinical marker of bacterial superinfection and clinical indication to antibiotic treatment, but there is no evidence that supports this concept (Murray et al. 2000). The color change is related to the recruitment of leukocytes into the airway lumen (Stockley et al. 2001). Neutrophils and activated monocytes contain chromatic, green (azurophil granules) containing myeloperoxidase with heme pigment. The more leukocytes present in nasal discharge the more

colorful the nasal discharge appears (Stockley et al. 2001). Although the literature is related to sputum color changes, the same mechanisms apply to nasal discharge.

4.4 Sneezing

A watery nasal secretion in the infection's early stage is often accompanied by sneezing. Sneezing together with a sore throat are the early symptoms of respiratory tract infections. Sneezing is a reflex mediated by the trigeminal nerves which supply the nasal epithelium (Leung and Robson 1994; Eccles 2005). The sneeze center in the brainstem coordinates the sneeze reflex. Sneezing is related to inflammatory responses in the nose and nasopharynx that stimulate the trigeminal nerves (Eccles 2005). As intranasal administration of histamine causes sneezing, sneezing is probably mediated by histamine receptors on the trigeminal nerves (Mygind et al. 1983; Eccles 2005).

4.5 Cough

Coughing is the most common clinical symptom and the most frequent reason for visits to see a doctor (McGarvey and Morice 2006). Coughing is a protective reflex that prevents the aspiration of food and fluids into the airway and cleans the respiratory tract of mucus and other foreign bodies. The reflex is mediated exclusively by the vagus nerve (Eccles 2005). Coughing is initiated in the airway through stimulation of the sensory nerves in the larynx or below (Widdicombe 1995). The airway inflammation associated with rhinitis must reach the larynx to cause coughing. The external ear and esophagus are also supplied by the vagus nerve and coughing can also be triggered by gastroesophageal reflux (Morice 2002). Respiratory tract infections are often accompanied by redundant, dry, and unproductive coughing in the first days. The unproductive coughing may be caused by the inflammatory process spreading to the larynx since nasal inflammation causes sneezing rather

coughing. Coughing in respiratory tract infections is believed to be mediated by hyperreactivity of the cough reflex due to the effects of inflammatory mediators on the airway's sensory nerve endings (Lee et al. 2002; Eccles and Lee 2004). When the larynx is inflamed and hyperreactive, coughing may occur spontaneously or in response to stimuli that would not normally cause coughing, e.g., cold air. It may persist for three weeks or longer. Some coughs may be voluntary and related to airway irritation (Lee et al. 2002). Productive coughing usually occurs later in the course of respiratory tract infection and is related to the mucus production associated with inflammation of the lower airways (Eccles 2005). Rhinovirus and coronavirus do not usually cause significant damage to the airway cells and the common cold is typically associated with little, if any, coughing while the influenza virus may cause substantial cellular damage to the respiratory epithelium and an influenza infection is usually associated with coughing (Monto et al. 2000).

4.6 Malaise and Mood Changes

Respiratory tract infections are associated with psychomotor impaired function et al. 1998). Mood changes and malaise may be explained by the unpleasant objective symptoms of respiratory tract infections such as nasal congestion, rhinorrhea, and coughing (Eccles 2005). These symptoms may cause discomfort and lower the patient's quality of life. However, there is increasing evidence that mood changes may also be caused by the effects of cytokines on the central nervous system (Mahoney and Ball 2002). Interferon alpha treatment for chronic hepatitis B and C is associated with flu-like adverse effects similar to those observed in respiratory tract infections: malaise, fever, myalgia, and mood changes (Schaefer et al. 2002). Psychiatric adverse effects such as depression, irritability, impaired concentration, and psychoses have been reported with interferon alpha therapy. It has been reported that cytokines, e.g., tumor necrosis factor- α (TNF- α) and interleukins 1, 2, and 6 cause mood changes with anhedonia, cognitive dysfunction, anxiety, irritability, psychomotor slowing, anorexia, sleep alterations, and a lower pain threshold (Capuron and Miller 2004). The production of these cytokines is also associated with respiratory tract infections which may mediate mood changes associated with these infections. The exact mechanisms of cytokine action in the brain are poorly understood, but there is a growing body of evidence suggesting that anorexia associated with respiratory infections is mediated by cytokines that act directly on the feeding center in the hypothalamus (Langhans 2000).

4.7 Headache

Headaches are a common early symptom of respiratory tract infections. The majority (60 %) of patients with respiratory tract infections with a sore throat reported headaches in a clinical trial (Eccles et al. 2003). The mechanism of a headache associated with a respiratory tract infection is unknown but headaches may be triggered by cytokines released in response to a viral infection (Smith 1992). It has been shown that the administration of some cytokines such as tumor necrosis factor and interferons cause headaches (Smith 1992; Gold et al. 2005; van Zonneveld et al. 2005).

4.8 Muscle Pain

Myalgia is a common symptom of respiratory tract infections. Around half of patients with a common cold complain about muscle pain (Eccles et al. 2003; Eccles 2005). Myalgia occurs in the acute immune response to infection phase and is related to the effects of cytokines on skeletal muscles (Baracos et al. 1983). Proinflammatory cytokines including TNF-α have been implicated in the breakdown of muscle proteins (Kotler 2000). Fever and myalgia associated with respiratory tract infection may be caused by the production of prostaglandin E2 response to cytokines (Baracos et al. 1983). The cytokine-related synthesis of prostaglandin E2 and the breakdown of skeletal muscle has been inhibited *in vitro* by non-steroidal anti-inflammatory agents and similarly fever and myalgia accompanying acute respiratory infection are relieved with acetylsalicylic acid, a classic anti-inflammatory agent (Eccles et al. 2003). Since prostaglandin E2 is a pain mediator, its increased synthesis may explain the myalgia associated with acute respiratory tract infections.

4.9 Fever and Chills

Fever is a classic response to infection. It is a manifestation of cytokine release in response to a variety of stimuli. It is believed to be beneficial for the host's response to infection (Cabanac 1990; Eccles 2005) and is usually associated with novel or severe viral infections such as influenza (Monto et al. 2000). Consequently, fever is a common symptom in infants, probably because viruses responsible for acute respiratory tract infections are new to the infant and induce a strong immune response. However, in adults who had been exposed to numerous common cold viruses in the past, subsequent infections do not elicit a strong cytokine response and fever is a rare symptom of a common cold in adults (Eccles 2005). On the contrary, some patients experience a transient fall in body temperature during the early stages of acute benign respiratory tract infection and about 1/3 of all patients experience chills (Eccles et al. 2003). Chills associated with a fall in skin temperature related to vasoconstriction of the skin's blood vessels may be explained as an initial stage of fever, but chills may also be unrelated to changes in skin temperature. Chills have developed after administration of exogenous pyrogens, even if the subjects maintained a neutral skin temperature (34.5 °C in water) in experimental human studies (Guieu and Hellon 1980). Chills occur together with shivering and the latter symptom is probably related to the cerebral cortex influence over the shivering control. Both chills and shivering may be caused by cytokines acting on the temperature control center of the hypothalamus. Many cytokines act as endogenous pyrogens and are released from leukocytes in response to infection (Conti et al. 2004). The proinflammatory cytokines (IL-1 and IL-6) are regarded as the most important cytokines for causing fever (Netea et al. 2000; Leon 2002). They are believed to cross the blood-brain barrier and increase the thermal set point in the temperature control center. The hypothalamus then induces shivering, constriction of the skin's blood vessels, and chills (Eccles 2005).

4.10 Febrile Seizures

Febrile seizures are a rare but significant symptom of acute viral respiratory infections in children. They occur in 2-5 % of all children and the majority of febrile seizures are triggered by respiratory tract infection (Schuchmann et al. 2011). Febrile seizures are defined as occurring in children aged 6-60 months with a temperature ≥38.0 °C with no central nervous system infection, metabolic disturbance, or history of afebrile seizure. They are the most common type of seizure in children under 60 months (American Academy of Pediatrics 2011; Graves et al. 2012). Cytokines seem to play a crucial role in febrile seizures, however there is a lot of confusion about the relationship proinflammatory and anti-inflammatory cytokines and the febrile seizure risk. Is it generally accepted that the genotype IL- 1α -889 1/1 and IL-1 β -511 T/T homozygote as well as the serum concentration of IL-6 are associated with an increased risk of febrile seizures (Saghazadeh et al. 2014).

5 Therapeutic Points

The treatment of acute viral respiratory tract infections remains primarily supportive. There is evidence that medications like acetaminophen (paracetamol) and non-steroidal anti-inflammatory agents such as ibuprofen or aspirin relieve some symptoms of acute respiratory tract infections (fever, sore throat, pain, and malaise),

but it is debatable whether symptomatic treatment could speed up recovery (Eccles 2005). Many other common advices like drinking plenty of fluids or steam inhalation have not been scientifically proven. Some new agents seem to be promising. For example in a study by Asada et al. (2012) L-carbocysteine reduced the baseline and RS virus infection-induced secretion of pro-inflammatory cytokines, including IL-1β, IL-6, and IL-8 as well as virus titers in the supernatant of human tracheal epithelial cells culture. Although a virus-orientated approach and the development of anti-viral agents should be more beneficial, the diverse etiology makes the development of universal antivirals highly unlikely (Passioti et al. 2014).

6 Conclusions

Since clinical symptoms are not sufficient to determine the etiology of acute viral respiratory tract infections, we believe that the host defense mechanisms are critical to the symptomatology. Immune response seems to be fundamental for understanding the pathomechanisms of these infections. Inflammatory mediators such as prostaglandins and bradykinin are responsible for the local symptoms of nasal congestion and rhinorrhea, while cytokines are responsible for systemic symptoms. A better understanding of the immune response including cytokines interactions will eventually allow for better treatments to be developed and should improve the quality of care for patients with acute respiratory tract infections.

Conflicts of Interest The authors have no funding or conflicts of interest to disclose.

References

- Akerlund A, Greiff L, Andersson M, Bende M, Alkner U, Persson CG (1993) Mucosal exudation of fibrinogen in coronavirus-induced common colds. Acta Otolaryngol 113(5):642–648
- American Academy of Pediatrics (2011) Subcommittee on Febrile Seizures. Neurodiagnostic evaluation of the child with a simple febrile seizure. Pediatrics 127 (2):389–394

- Arbour N, Day R, Newcombe J, Talbot PJ (2000) Neuroinvasion by human respiratory coronaviruses. J Virol 74(19):8913–8921
- Arden KE, Mackay IM (2010) Newly identified human rhinoviruses: molecular methods heat up the cold viruses. Rev Med Virol 20:156–176
- Arden KE, Nissen MD, Sloots TP, Mackay IM (2005) New human coronavirus, HCoV-NL63, associated with severe lower respiratory tract disease in Australia. J Med Virol 75:455–462
- Arruda E, Boyle TR, Winther B, Pevear DC, Gwaltney JM Jr, Hayden FG (1995) Localization of human rhinovirus replication in the upper respiratory tract by in situ hybridization. J Infect Dis 17:1329–1333
- Asada M, Yoshida M, Hatachi Y, Sasaki T, Yasuda H, Deng X, Nishimura H, Kubo H, Nagatomi R, Yamaya M (2012) L-carbocisteine inhibits respiratory syncytial virus infection in human tracheal epithelial cells. Respir Physiol Neurobiol 180(1):112–118
- Baracos V, Rodemann HP, Dinarello CA, Goldberg AL (1983) Stimulation of muscle protein degradation and prostaglandin E2 release by leukocyticpyrogen (interleukin-1). A mechanism for the increased degradation of muscle proteins during fever. N Engl J Med 308(10):553–558
- Bardin PG, Johnston SL, Sanderson G, Robinson BS, Pickett MA, Fraenkel DJ, Holgate ST (1994) Detection of rhinovirus infection of the nasal mucosa by oligonucleotide in situ hybridization. Am J Respir Cell Mol Biol 10(2):207–213
- Barr FE, Graham BS (2014) Respiratory syncytial virus infection: clinical features and diagnosis. In: UpToDate [database online]. Wolters Kluwer Health. http://www.uptodate.com/contents/respiratory-syncy tial-virus-infection-clinical-features-and-diagnosis? source=search_result&search=rSV& selectedTitle=1~138. Updated September 02, 2014. Accessed 3 Sept 2014
- Bella J, Rossmann MG (2000) ICAM-1 receptors and cold viruses. Pharm Acta Helv 74(2–3):291–297
- Beutler B (2003) Science review: key inflammatory and stress pathways in critical illness the central role of the Toll-like receptors. Crit Care 7(1):39–46
- Bochkov YA, Gern JE (2012) Clinical and molecular features of human rhinovirus C. Microbes Infect 14 (6):485–494
- Bochkov YA, Palmenberg AC, Lee WM, Rathe JA, Amineva SP, Sun X, Pasic TR, Jarjour NN, Liggett SB, Gern JE (2011) Molecular modeling, organ culture and reverse genetics for a newly identified human rhinovirus C. Nat Med 17:627–632
- Cabanac M (1990) Phylogeny of fever. In: Bligh J, Voigt K (eds) Thermoreception and temperature regulation. Springer, Berlin, pp 284–296
- Callow KA, Parry HF, Sergeant M, Tyrrell DA (1990) The time course of the immune response to experimental coronavirus infection of man. Epidemiol Infect 105(2):435–446
- Capuron L, Miller AH (2004) Cytokines and psychopathology: lessons from interferon-alpha. Biol Psychiatry 56(11):819–824

- Collins AR (2002) In vitro detection of apoptosis in monocytes/macrophages infected with human coronavirus. Clin Diagn Lab Immunol 9:1392–1395
- Conti B, Tabarean I, Andrei C, Bartfai T (2004) Cytokines and fever. Front Biosci 9:143–149
- de Wit E, Munster VJ, van Riel D, Beyer WE, Rimmelzwaan GF, Kuiken T, Osterhaus AD, Fouchier RA (2010) Molecular determinants of adaptation of highly pathogenic avian influenza H7N7 viruses to efficient replication in the human host. J Virol 84:1597–1606
- Desforges M, Miletti T, Gagnon M, Talbot PJ (2006) HCoV-229E infects and activates monocytes. Adv Exp Med Biol 581:511–514
- Douglass JA, Dhami D, Gurr CE, Bulpitt M, Shute JK, Howarth PH, Lindley IJ, Church MK, Holgate ST (1994) Influence of interleukin-8 challenge in the nasal mucosa in atopic and nonatopic subjects. Am J Respir Crit Care Med 150(4):1108–1113
- Eccles R (1983) Physiology of nasal secretion. Eur J Respir Dis 62:115–119
- Eccles R (2000a) Nasal airflow in health and disease. Acta Otolaryngol 120:580–595
- Eccles R (2000b) Pathophysiology of nasal symptoms. Am J Rhinol 14:335–338
- Eccles R (2002) Acute cooling of the body surface and the common cold. Rhinology 40(3):109–114
- Eccles R (2005) Understanding the symptoms of the common cold and influenza. Lancet Infect Dis 5 (11):718–725
- Eccles R, Lee PC (2004) Cough induced by airway vibration as a model of airway hyperreactivity in patients with acute upper respiratory tract infection. Pulm Pharmacol Ther 17:337–342
- Eccles R, Reilly M, Eccles KSJ (1996) Changes in the amplitude of the nasal cycle associated with symptoms of acute upper respiratory tract infection. Acta Otolaryngol 116:77–81
- Eccles R, Loose I, Jawad M, Nyman L (2003) Effects of acetylsalicylic acid on sore throat pain and other pain symptoms associated with acute upper respiratory tract infection. Pain Med 4:118–124
- Esper F, Weibel C, Ferguson D, Landry ML, Kahn JS (2005) Evidence of a novel human coronavirus that is associated with respiratory tract disease in infants and young children. J Infect Dis 191:492–498
- Esposito S, Daleno C, Scala A, Castellazzi L, Terranova L, Sferrazza Papa S, Longo MR, Pelucchi C, Principi N (2014) Impact of rhinovirus nasopharyngeal viral load and viremia on severity of respiratory infections in children. Eur J Clin Microbiol Infect Dis 33:41–48
- Everard ML, Swarbrick A, Wrightham M, McIntyre J, Dunkley C, James PD, Sewell HF, Milner AD (1994) Analysis of cells obtained by bronchial lavage of infants with respiratory syncytial virus infection. Arch Dis Child 71(5):428–432

- Fukuyama S, Kawaoka Y (2011) The pathogenesis of influenza virus infections: the contributions of virus and host factors. Curr Opin Immunol 23(4):481–486
- Garofalo RP, Patti J, Hintz KA, Hill V, Ogra PL, Welliver RC (2001) Macrophage inflammatory protein-1alpha (not T helper type 2 cytokines) is associated with severe forms of respiratory syncytial virus bronchiolitis. J Infect Dis 184(4):393–399
- Garofalo RP, Hintz KH, Hill V, Patti J, Ogra PL, Welliver RC Sr (2005) A comparison of epidemiologic and immunologic features of bronchiolitis caused by influenza virus and respiratory syncytial virus. J Med Virol 75(2):282–289
- Georgitis JW (1993) Nasopharyngitis, pharyngitis, and tonsillitis. Immunol Allergy Clin North Am 13:109–118
- Gething MJ, Bye J, Skehel J, Waterfield M (1980) Cloning and DNA sequence of double-stranded copies of haemagglutinin genes from H2 and H3 strains elucidates antigenic shift and drift in human influenza virus. Nature 287(5780):301–306
- Gold R, Rieckmann P, Chang P, Abdalla J (2005) The long-term safety and tolerability of high-dose interferon beta-1a in relapsing-remitting multiple sclerosis: 4-year data from the PRISMS study. Eur J Neurol 12:649–656
- Gonzales R, Malone DC, Maselli JH, Sande MA (2001) Excessive antibiotic use for acute respiratory infections in the United States. Clin Infect Dis 33:757–762
- Graves RC, Oehler K, Tingle LE (2012) Febrile seizures: risks, evaluation, and prognosis. Am Fam Physician 85(2):149–153
- Guieu JD, Hellon RF (1980) The chill sensation in fever. Pflugers Arch 384:103–104
- Han TH, Chung JY, Kim SW, Hwang ES (2007) Human coronavirus-NL63 infections in Korean children, 2004–2006. J Clin Virol 38:27–31
- Hayden FG, Diamond L, Wood PB, Korts DC, Wecker MT (1996) Effectiveness and safety of intranasal ipratropium bromide in common colds. A randomized, double-blind, placebo-controlled trial. Ann Intern Med 125:89–97
- Hayden FG, Fritz R, Lobo MC, Alvord W, Strober W, Straus SE (1998) Local and systemic cytokine responses during experimental human influenza A virus infection. Relation to symptom formation and host defense. J Clin Invest 101(3):643–649
- Heikkinen T, Järvinen A (2003) The common cold. Lancet 361:51–59
- Hendley JO (1998) The host response, not the virus, causes the symptoms of the common cold. Clin Infect Dis 26(4):847–848
- Hers JF (1966) Disturbances of the ciliated epithelium due to influenza virus. Am Rev Respir Dis 93(3), Suppl:162–177
- Hers JF, Mulder J (1961) Broad aspects of the pathology and pathogenesis of human influenza. Am Rev Respir Dis 83(2):84–97

E. Kuchar et al.

- Heymann PW, Platts-Mills TA, Johnston SL (2005) Role of viral infections, atopy and antiviral immunity in the etiology of wheezing exacerbations among children and young adults. Pediatr Infect Dis J 24(11)Suppl: S217–S222
- Hsiao CJ, Cherry DK, Beatty PC, Rechtsteiner EA (2010) National ambulatory medical care survey: 2007 summary. Natl Health Stat Rep 27:1–32
- Jackson GG, Dowling HF, Spiesman IG, Boand AV (1958) Transmission of the common cold to volunteers under controlled conditions. I. The common cold as a clinical entity. Arch Intern Med 101 (2):267–278
- Jartti T, Jartti L, Peltola V, Waris M, Ruuskanen O (2008) Identification of respiratory viruses in asymptomatic subjects: asymptomatic respiratory viral infections. Pediatr Infect Dis J 27(12):1103–1107
- Jartti T, Jartti L, Ruuskanen O, Söderlund-Venermo M (2012) New respiratory viral infections. Curr Opin Pulm Med 18(3):271–278
- Johnson JE, Gonzales RA, Olson SJ, Wright PF, Graham BS (2007) The histopathology of fatal untreated human respiratory syncytial virus infection. Mod Pathol 20(1):108–119
- Johnston S, Holgate S (1996) Epidemiology of viral respiratory infections. In: Myint S, Taylor Robinson D (eds) Viral and other infections of the human respiratory tract. Chapman & Hall, London, pp 1–38
- Kahn JS (2006) The widening scope of coronaviruses. Curr Opin Pediatr 18:42–47
- Kaiser L, Fritz RS, Straus SE, Gubareva L, Hayden FG (2001) Symptom pathogenesis during acute influenza: interleukin-6 and other cytokine responses. J Med Virol 64(3):262–268
- Kennedy LJ, Turner RB, Braciale T, Heymann PW, Borish L (2012) Pathogenesis of rhinovirus infection. Curr Opin Virol 2(3):287–293
- Kotler DP (2000) Cachexia. Ann Intern Med 133:622–634
- Kuchar E, Nitsch-Osuch A, Karpinska T, Kurpas D, Zycinska K, Wardyn K, Szenborn L (2013) Pandemic influenza in the 2009/2010 season in central Poland: the surveillance study of laboratory confirmed cases. Respir Physiol Neurobiol 187(1):94–98
- Künkel F, Herrler G (1996) Structural and functional analysis of the S proteins of two human coronavirus OC43 strains adapted to growth in different cells. Arch Virol 141(6):1123–1131
- Langhans W (2000) Anorexia of infection: current prospects. Nutrition 16:996–1005
- Lee P, Cotterill-Jones C, Eccles R (2002) Voluntary control of cough. Pulm Pharmacol Ther 15:317–320
- Lee WM, Lemanske RF Jr, Evans MD, Vang F, Pappas T, Gangnon R, Jackson DJ, Gern JE (2012) Human rhinovirus species and season of infection determine illness severity. Am J Respir Crit Care Med 186:886–891

- Legg JP, Hussain IR, Warner JA, Johnston SL, Warner JO (2003) Type 1 and type 2 cytokine imbalance in acute respiratory syncytial virus bronchiolitis. Am J Respir Crit Care Med 168(6):633–639
- Leon LR (2002) Invited review: cytokine regulation of fever: studies using gene knockout mice. J Appl Physiol 92:2648–2655
- Leung AK, Robson WL (1994) Sneezing. J Otolaryngol 23:125–129
- Mahoney T, Ball P (2002) Common respiratory tract infections as psychological entities: a review of the mood and performance effects of being ill. Aust Psychol 37:86–94
- Matsuda K, Tsutsumi H, Okamoto Y, Chiba C (1995) Development of interleukin 6 and tumor necrosis factor alpha activity in nasopharyngeal secretions of infants and children during infection with respiratory syncytial virus. Clin Diagn Lab Immunol 2(3):322–324
- McGarvey LPA, Morice AH (2006) Clinical cough and its mechanisms. Respir Physiol Neurobiol 152 (3):363–371
- Mellow TE, Murphy PC, Carson JL, Noah TL, Zhang L, Pickles RJ (2004) The effect of respiratory synctial virus on chemokine release by differentiated airway epithelium. Exp Lung Res 30(1):43–57
- Mesel-Lemoine M, Millet J, Vidalain PO, Law H, Vabret A, Lorin V, Escriou N, Albert ML, Nal B, Tangy F (2012) A human coronavirus responsible for the common cold massively kills dendritic cells but not monocytes. J Virol 86(14):7577–7587
- Milewska A, Ciejka J, Kaminski K, Karewicz A, Bielska D, Zeglen S, Karolak W, Nowakowska M, Potempa J, Bosch BJ, Pyrc K, Szczubialka K (2013) Novel polymeric inhibitors of HCoV-NL63. Antiviral Res 97(2):112–121
- Monto AS, Gravenstein S, Elliott M, Colopy M, Schweinle J (2000) Clinical signs and symptoms predicting influenza infection. Arch Intern Med 160:3243–3247
- Morice AH (2002) Epidemiology of cough. Pulm Pharmacol Ther 15:253–259
- Murray S, Del Mar C, O'Rourke P (2000) Predictors of an antibiotic prescription by GPs for respiratory tract infections: a pilot. Fam Pract 17:386–388
- Mygind N, Secher C, Kirkegaard J (1983) Role of histamine and antihistamines in the nose. Eur J Respir Dis Suppl 128:16–20
- Naclerio RM, Proud D, Lichtenstein LM, Kagey-Sobotka A, Hendley JO, Sorrentino J, Gwaltney JM (1988) Kinins are generated during experimental rhinovirus colds. J Infect Dis 157(1):133–142
- Netea MG, Kullberg BJ, Van der Meer JW (2000) Circulating cytokines as mediators of fever. Clin Infect Dis 31(Suppl 5):S178–S184
- Noah TL, Henderson FW, Wortman IA, Devlin RB, Handy J, Koren HS, Becker S (1995) Nasal cytokine

- production in viral acute upper respiratory infection of childhood. J Infect Dis 171(3):584–592
- Ohmit SE, Monto AS (2006) Symptomatic predictors of influenza virus positivity in children during the influenza season. Clin Infect Dis 43(5):564–568
- Oosterhof L, Christensen CB, Sengelov H (2010) Fatal lower respiratory tract disease with human corona virus NL63 in an adult haematopoietic cell transplant recipient. Bone Marrow Transplant 45:1115–1116
- Pappas C, Aguilar PV, Basler CF, Solorzano A, Zeng H, Perrone LA, Palese P, Garcia-Sastre A, Katz JM, Tumpey TM (2008) Single gene reassortants identify a critical role for PB1, HA, and NA in the high virulence of the 1918 pandemic influenza virus. Proc Natl Acad Sci U S A 105:3064–3069
- Passioti M, Maggina P, Megremis S, Papadopoulos NG (2014) The common cold: potential for future prevention or cure. Curr Allergy Asthma Rep 14(2):1–11
- Peebles RS Jr, Graham BS (2005) Pathogenesis of respiratory syncytial virus infection in the murine model. Proc Am Thorac Soc 2(2):110–115
- Peltola V, Waris M, Osterback R, Susi P, Hyypia T, Ruuskanen O (2008) Clinical effects of rhinovirus infections. J Clin Virol 43(4):411–414
- Pinto R, Herold S, Cakarova L, Hoegner K, Lohmeyer J, Planz O, Pleschka S (2011) Inhibition of influenza virusinduced NF-kappaB and raf/MEK/ERK activation can reduce both virus titers and cytokine expression simultaneously in vitro and in vivo. Antiviral Res 92(1):45–56
- Proud D (1998) The kinin system in rhinitis and asthma. Clin Rev Allergy Immunol 16(4):351–364
- Proud D, Reynolds CJ, Lacapra S, Kagey-Sobotka A, Lichtenstein LM, Naclerio RM (1988) Nasal provocation with bradykinin induces symptoms of rhinitis and a sore throat. Am Rev Respir Dis 137(3):613–616
- Proud D, Naclerio RM, Gwaltney JM, Hendley JO (1990) Kinins are generated in nasal secretions during natural rhinovirus colds. J Infect Dis 161(1):120–123
- Pyrc K, Berkhout B, van der Hoek L (2007) The novel human coronaviruses NL63 and HKU1. J Virol 81:3051–3057
- Rees GL, Eccles R (1994) Sore throat following nasal and oropharyngeal bradykinin challenge. Acta Otolaryngol 114:311–314
- Rezaee F, Meednu N, Emo JA, Saatian B, Chapman TJ, Naydenov NG, De Benedetto A, Beck LA, Ivanov AI, Georas SN (2011) Polyinosinic: polycytidylic acid induces protein kinase D dependent disassembly of apical junctions and barrier dysfunction in airway epithelial cells. J Allergy Clin Immunol 128(6):1216–1224.e11
- Ruuskanen O, Waris M, Ramilo O (2013) New aspects on human rhinovirus infections. Pediatr Infect Dis J 32 (5):553–555
- Saghazadeh A, Gharedaghi M, Meysamie A, Bauer S, Rezaei N (2014) Proinflammatory and anti-inflammatory cytokines in febrile seizures and epilepsy: systematic review and meta-analysis. Rev Neurosci 25(2):281–305

- Schaefer M, Schmidt F, Neumer R, Scholler G, Schwarz M (2002) Interferon-alpha, cytokines and possible implications for mood disorders. Bipolar Disord 4 (Suppl 1):111–113
- Schuchmann S, Hauck S, Henning S, Grüters-Kieslich A, Vanhatalo S, Schmitz D, Kaila K (2011) Respiratory alkalosis in children with febrile seizures. Epilepsia 52 (11):1949–1955
- Shibayama Y, Skoner D, Suehiro S, Konishi JE, Fireman P, Kaplan AP (1996) Bradykinin levels during experimental nasal infection with rhinovirus and attenuated influenza virus. Immunopharmacology 33:311–331
- Shinya K, Ebina M, Yamada S, Ono M, Kasai N, Kawaoka Y (2006) Avian flu: influenza virus receptors in the human airway. Nature 440:435–436
- Simmonds P, McIntyre C, Savolainen-Kopra C, Tapparel C, Mackay IM, Hovi T (2010) Proposals for the classification of human rhinovirus species C into genotypically assigned types. J Gen Virol 91:2409–2419
- Smith RS (1992) The cytokine theory of headache. Med Hypotheses 39:168–174
- Smith A, Thomas M, Kent J, Nicholson K (1998) Effects of the common cold on mood and performance. Psychoneuroendocrinology 23:733–739
- Smyth RL, Fletcher JN, Thomas HM, Hart CA (1997) Immunological responses to respiratory syncytial virus infection in infancy. Arch Dis Child 76 (3):210–214
- Snow V, Mottur-Pilson C, Gonzales R, American College of Physicians-American Society of Internal Medicine, American Academy of Family Physicians, Centers for Disease Control, Infectious Diseases Society of America (2001) Principles of appropriate antibiotic use for treatment of nonspecific upper respiratory tract infections in adults. Ann Intern Med 134(6):487–489
- Spector SL (1995) The common cold: current therapy and natural history. J Allergy Clin Immunol 95(5 Pt 2):1133–1138
- Stockley RA, Bayley D, Hill SL, Hill AT, Crooks S, Campbell EJ (2001) Assessment of airway neutrophils by sputum colour: correlation with airways inflammation. Thorax 56:366–372
- Sung JY, Lee HJ, Eun BW, Kim SH, Lee SY, Lee JY, Park KU, Choi EH (2010) Role of human coronavirus NL63 in hospitalized children with croup. Pediatr Infect Dis J 29:822–826
- Treanor J (2004) Influenza vaccine outmaneuvering antigenic shift and drift. N Engl J Med 350 (3):218–220
- Turner RB (1997) Epidemiology, pathogenesis, and treatment of the common cold. Ann Allergy Asthma Immunol 78:531–539
- Turner RB (2011) Chap. 369: The common cold. In: Goldman L, Schafer AI (eds) Goldman's Cecil medicine, 24th edn. Saunders Elsevier, Philadelphia

E. Kuchar et al.

- Turner RB, Hendley JO, Gwaltney JM Jr (1982) Shedding of infected ciliated epithelial cells in rhinovirus colds. J Infect Dis 145:849–853
- Tyrrell DAJ, Cohen S, Schlarb JE (1993) Signs and symptoms in common colds. Epidemiol Infect 111(1):143–156
- Vabret A, Mourez T, Dina J, van der Hoek L, Gouarin S, Petitjean J, Brouard J, Freymuth F (2005) Human coronavirus NL63, France. Emerg Infect Dis 11:1225–1229
- van der Hoek L, Sure K, Ihorst G, Stang A, Pyrc K, Jebbink MF, Petersen G, Forster J, Berkhout B, Uberla K (2005) Croup is associated with the novel coronavirus NL63. PLoS Med 2:e240
- van Zonneveld M, Flink HJ, Verhey E, Senturk H, Zeuzem S, Akarca US, Cakaloglu Y, Simon C, So TM, Gerken G, de Man RA, Hansen BE, Schalm SW, Janssen HL, HBV 99-01 Study Group (2005) The safety of pegylated interferon alpha-2b in the treatment of chronic hepatitis B: predictive factors for dose reduction and treatment discontinuation. Aliment Pharmacol Ther 21:1163–1171
- Vlasak M, Roivainen M, Reithmayer M, Goesler I, Laine P, Snyers L, Hovi T, Blaas D (2005) The minor receptor group of human rhinovirus (HRV) includes HRV23 and HRV25, but the presence of a lysine in the VP1 HI loop is not sufficient for receptor binding. J Virol 79(12):7389–7395
- Walsh EE, Shin JH, Falsey AR (2013) Clinical impact of human coronaviruses 229E and OC43 infection in diverse adult populations. J Infect Dis 208 (10):1634–1642
- Wat D (2004) The common cold: a review of the literature. Eur J Intern Med 15(2):79–88
- Welliver RC, Garofalo RP, Ogra PL (2002) Betachemokines, but neither T helper type 1 nor T helper type 2 cytokines, correlate with severity of illness during respiratory syncytial virus infection. Pediatr Infect Dis J 21:457–461
- Whiteman SC, Bianco A, Knight RA, Spiteri MA (2003) Human rhinovirus selectively modulates membranous

- and soluble forms of its intercellular adhesion molecule-1 (ICAM-1) receptor to promote epithelial cell infectivity. J Biol Chem 278(14):11954–11961
- Widdicombe JG (1995) Neurophysiology of the cough reflex. Eur Respir J 8:1103–1202
- Widdicombe JG (1997) Microvascular anatomy of the nose. Allergy 52:7–11
- Wine TM, Alper CM (2012) Cytokine responses in the common cold and otitis media. Curr Allergy Asthma Rep 12(6):574–581
- Winther B, Farr B, Turner RB, Hendley JO, Gwaltney JM, Mygind N (1984) Histopathologic examination and enumeration of polymorphonuclear leukocytes in the nasal mucosa during experimental rhinovirus colds. Acta Otolaryngol 413(Suppl):19–24
- Winther B, Gwaltney JM, Hendley JO (1990) Respiratory virus infection of monolayer cultures of human nasal epithelial cells. Am Rev Respir Dis 141 (4 Pt1):839–845
- Winther B, Greve JM, Gwaltney JM Jr, Innes DJ, Eastham JR, McClelland A, Hendley JO (1997) Surface expression of intercellular adhesion molecule 1 on epithelial cells in the human adenoid. J Infect Dis 176 (2):523–525
- Winther B, Gwaltney JM Jr, Mygind N, Hendley JO (1998) Viral-induced rhinitis. Am J Rhinol 12 (1):17–20
- Wu PS, Chang LY, Berkhout B, van der Hoek L, Lu CY, Kao CL, Lee PI, Shao PL, Lee CY, Huang FY, Huang LM (2008) Clinical manifestations of human coronavirus NL63 infection in children in Taiwan. Eur J Pediatr 167:75–80
- Zambon MC, Stockton JD, Clewley JP, Fleming DM (2001) Contribution of influenza and respiratory syncytial virus to community cases of influenza-like illness: an observational study. Lancet 358 (9291):1410–1416
- Zhu Z, Tang W, Gwaltney JM Jr, Wu Y, Elias JA (1997) Rhinovirus stimulation of interleukin-8 in vivo and in vitro: role of NF-kappaB. Am J Physiol 273(4 Pt 1):L814–L824