CLADISTIC ANALYSIS OF DASYUROMORPHIAN (MARSUPIALIA) PHYLOGENY USING CRANIAL AND DENTAL CHARACTERS

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Dasyuromorphian relationships were investigated using cladistic analysis for 24 species using 77 cranial and dental features. Among the 7 extinct taxa used were 6 recently described fossil species, each well represented by cranial and dental material (3 ameridelphians, 1 peramelemorphian, and 2 dasyuromorphians). Monophyly for the Dasyuromorphia and several clades widely recognized therein is supported, but in many instances, relationships among extant dasyurids departs greatly from general consensus. Where congruence with previous investigations is evident, few taxa are united by unique synapomorphies within Marsupialia. Many clades are united by combinations of locally derived features only. Bootstrap and Bremer support is weak for most clades. Thus, although sup- ported by cladistic analysis, the status of many synapomorphies identified in the course of this study are tentative. However, for some groups, notably Dasyuridae and a dasyurid clade inclusive of all modern subfamilies, the synapomorphic nature of some derived features appears to be robust, even where they also are present in some outgroup taxa. This argument applies to shared apomorphies of the basicranium in particular. No potential sister taxon to Dasyuromorphia is favored. The case for australidelphian and microbiotheriid affinity of some American and Antarctic fossil taxa was considered to be highly equivocal.

Key words: Ameridelphia, Australidelphia, Dasyuridae, Dasyuromorphia, *Badjcinus, Barinya, fossil*, Myrmecobiidae, phylogeny, Thylacinidae

A rapidly expanding body of both paleontological and molecular data has shed new light on Australian marsupicarnivore phylogeny (Cifelli and Muizon 1998; Kirsch et al. 1991, 1997; Krajewski et al. 1996, 1997a, 1997b; Marshall and Muizon 1995; Muirhead 1992, 1997; Muirhead and Wroe 1998; Muizon 1994,1998; Muizon et al. 1997; Springer et al. 1994, 1997; Wroe 1997b, 1998, 1999a, 1999b). Prior to description of the unallocated Eocene taxon *Djarthia murgonensis* (Godthelp et al. 1999), all Australian marsupicamivores had been placed in a single order, Dasyuromorphia. Following publication of molecular and morphological results in the early 1980s (Lowenstein et al. 1981; Szalay 1982), longrunning controversy over the possible borhyaenoid affinity of thylacinids has been laid to rest, with all subsequent authors treating Dasyuromorphia as monophyletic. This position has been reaffirmed by many subsequent investigators (Kra-

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jewski et al. 1997a; Muirhead and Wroe 1998; Thomas et al. 1989; Wroe 1996).

Widely accepted as the most generalized elements of the Australian marsupial radiation, dasyuromorphians, especially dasyurids, have figured prominently in discussions of Australian marsupial origins. Alternatively, Aplin and Archer (1987) and Wroe (1999a) expressed a contradictory view with respect to Dasyuridae. The position of D. murgonensis relative to dasyuromorphians is unclear. This new species. less derived for almost all dental features than any other Australian marsupial, may represent a basal element of Dasyuromorphia. However, given that D. murgonensis also is plesiomorphic within Marsupialia for most known features, it actually may be didelphimorphian (Godthelp et al. 1999). The relationship of dasyuromorphians to other marsupials also remains controversial. Possible sister taxa to Dasyuromorphia proposed in previous investigations include Didelphidae (Archer 1976b; Bensley 1903), Peramelemorphia (Kirsch 1977), Microbiotheriidae (Szalay 1994), and Microbiotheriidae plus Diprotodontia plus Notoryctidae (Kirsch et al. 1997). Marshall et al. (1990) and Woodburne and Case (1996) suggested a clade inclusive of all non-dasyuromorphian Australian marsupials as the sister clade to Dasyuromorphia. Those same authors proposed that the Paleocene Bolivian species Andinodelphys cochabambensis is the sister taxon to all Australian marsupials. The various interpretations of higher level phylogeny for these taxa have been reviewed in several recent papers (Aplin and Archer 1987; Kirsch et al. 1997; Luckett 1994; Marshall et al. 1990; Springer et al. 1997). Microbiotherians are central to the formulation of many hypotheses on Australian marsupial evolution and thus require special attention. In particular, efforts to link fossil microbiotherians and other American taxa with the Australian marsupial radiation demand more detailed consideration. We reviewed the literature bearing on the origins and phylogeny of Dasyuromorphia and performed a cladistic analysis using morphological features and new fossil material.

Possible fossil microbiotherians.-Microbiotherians from South America. Antarctica. and Australia have been considered the sister "on to dasyuromorphians (Szalay 1994), diprotodontians (Kirsch et al. 1997), the entire Australian marsupial radiation plus Andinodelphys cochabambensis (Marshall et al. 1990; Woodburne and Case 1996), and all other marsupials (Hershkovitz 1992, 1995, 1999). Of particular relevance here are proposals of monophyly for the ?microbiotheriid Mirandatherium. the Australian taxon Ankotarinja tirarensis (Marshall 1987), and A. cochabambensis and all Australian marsupials, a position argued by Marshall et al. (1990) and Woodburne and Case (1996) but rejected by Muizon (1992) and Muizon et al.. (1997). Other pertinent issues include the possible microbiotherian affinities of fossil material from the Eocene deposits of Murgon (Archer et al. 1993; Kirsch et al. 1997) and Seymour Island, Antarctica (Goin and Carlini 1995; Marenssi et al. 1994).

Inclusion of Mirandatherium within Microbiotheria was formally recognized by Marshal (1987:148), who stated that this taxon is "clearly a member of the subfamily Microbiotheriinae." Eomicrobiotherium, Pachybiotherium, and a pediomyid clade, which included Monodelphopsis, also were treated as microbiotheriids. Marshall et al. (1990) likewise treated pediomyids and Pachybiotherium as microbiotherians but reinterpreted Mirandatherium as а didelphine after reconsidering the state of 1 character, centrocrista straight or linear (Godthelp et al. 1999). In their cladogram, Marshall et al. (1990:438) portrayed A. cochabambensis as monophyletic with the marsupial clade. Australian Monodelphopsis, a pediomyine under Marshall's (1987) classification, was rediagnosed as a microbiotheriid by Marshall et al. (1990). Previously, pediomyids had been placed as the sister taxon to Borhyaenoidea (Marshall

1977). Phylogenies presented by Marshall (1987) and Marshall et al. (1990) contained many clades that appeared to be based, at least in part, on presence of shared plesiomorphies or overall similarity (Godthelp et al. 1999; Muizon 1992). For example, presence of a linear centrocrista was used as part of the basis for treating Ankotarinja tir- arensis as a possible sister taxon to Mir- and atherium by Marshall (1987), but the same author treated that feature as a mar- supial plesiomorphy. Moreover, in our view, the centrocrista of A. tirarensis is V- shaped. We consider other proposed syna- pomorphies to be highly equivocal. Marshall et al. (1990:442) listed 4 possible shared derived features to support their con- cept of Microbiotheria. None of these fea- tures are exclusive to Microbiotheria (sensu Marshall et al. 1990). Some characters are extremely labile and have appeared inde- pendently in several marsupial clades (e.g., reduction of the stylar shelf; stylar cusp B reduced, with D the largest cusp). Of the 13 proposed anatomical synapomorphies uni- fying Australidelphia (sensu Marshall et al. 1990), -2 are marsupial plesiomorphies (i.e., entoconid and hypoconid large, subequal in size; molars semibrachyodont). Others traits are locally derived dental features, often found in many other marsupicarni- vore clades (e.g., subequal stylar cusps B and D; talonid wider than trigonid on M2- 3; cristid obliqua meets rear of trigonid la- bial to protocristid notch; entoconid notch weak; upper incisors spatulate shaped).

Godthelp et al. (1999) discussed problems with interpretation of dental data used to support monophyly for the putative australidelphian *Andinodelphys cochabambensis* and Australian marsupials, as suggested by Marshall et al. (1990) and Woodburne and Case (1996), and found those arguments unconvincing, a view shared by Kirsch et al. (1997). Most features used in support of this special relationship are highly mutable among marsupials, some at the intraspecific and even individual level. For example, presence of twinned cusps in the

C position, present in then-known specimens of A. cochabambensis and undescribed peramelid material and treated by Marshall et al. (1990) as a possible synapomorphy uniting A. cochabambensis and Australian marsupials, was considered a poor basis for the postulation of special relationship (cL Muizon 1992). This skepticism has been vindicated with the discovery of additional specimens of A. cochabambensis wherein twinned cusps in the C position are variably present and sometimes present on 1 side of a specimen but not the other (Muizon et al. 1997). Furthermore, from new well-preserved cranial material, it is clear that A. cochabambensis is plesiomiorphic within Marsupialia for almost all cranial features. The only derived cranial feature identified by Muizon et al. (1997) was considered supportive of a special relationship with 2 other Tiupampan ameridelphians, Pucadelphys and Inavulestes ferox.

Because Microbiotheriidae (sensu Marshall 1987; Marshall et al. 1990) is not well defined on the basis of dental data. referral of new fossil taxa to this clade should be treated with circumspection. The suggestion that undescribed Pachybiotherium-like material from Murgon constitutes evidence for existence of microbiotheriids in Australia (Archer et al. 1993) requires further substantiation. This view is strengthened when it is considered that Microbiotheriidae was defined almost exclusively on the basis of cranial features by Marshall et al. (1990), wherein only 1 dental feature was noted, but Pachybiotherium, treated as a microbiotheriid by the same authors, is known only from dental material. Similar problems arise with the classification of other possible fossil microbiotherians. Woodbume and Case (1996:133) regarded Khasia from the Paleocene Tiupampa Local Fauna of Bolivia as "completely specialized toward microbiotheres." Proposed synapomorphies supporting the position were "reduction of stylar shelf and cusps, stylar cusp A reduced versus stylar cusps B, C, and D, the

paraconid reduced and positioned anteriorly, the oblique orientation of the protocristid, and the talonid wider than the trigonid." Of those characters, it is unclear as to how 1, i.e., oblique orientation of the protocristid, can be interpreted as suggesting affinity with microbiotheres. This feature is certainly derived within Marsupialia but is not common to Dromiciops, Microbiotherium, or pediomyids. All of the remaining features are shared with 1_demonstrably nonmicrobiotherian clade, with some features, such as reduction of the stylar shelf and stylar cusp A relative to other stylar cusps, independently derived in many didelphimorphian and dasyuromorphian taxa.

The microbiotherian status of some Early Tertiary Antarctic material also is tenuous. Marenssi et al. (1994) informally and Goin and Carlini (1995) formally referred a partial edentulate dentary from Seymour Island to Microbiotheriidae. That specimen (MLP 88-I-1-1) originally was referred to Didelphimorphia by S. E Vizcaino et al. (in litt.). Goin and Carlini (1995:205) cited 2 synapomorphies of the dentary, originally presented by Marshall (1982), in support of a microbiotheriid affinity for MLP 88-I-1-1, i.e., "large mental foramen below P, and depth of ramus relatively constant from back to front." A well-developed mental foramen below p2 is present in at least some peramelemorphians (e.g., Echymipera kalubu, Perameles nasuta), dasyuromorphians (e.g., Neophascogale lorentzii, Dasyurus maculatus, Thylacinus cynocephalus), borhyaenoids (e.g., Mayulestes ferox), didelphoids (e.g., Pucadelphys andinus), and peradectids (e.g., Alphadon eatoni). On the basis of distribution, the phylogenetic value of this feature within Marsupialia is questionable. Given its presence in peradectids, basal representatives of Didelphoidea and Borhyaenoidea, and the undoubted microbiotheriid Dromiciops gliroides, it seems likely that this feature 'is plesiomorphic within Marsupialia. The degree of confidence that can be attributed to the 2nd feature is difficult to assess without examining

the specimen. Marshall et al. (1990:438) used neither of these characters to define Australidelphia, Microbiotheria, or Microbiotheriidae in their cladogram.

Although the possibility that some or all of the fossil ?microbiotherian taxa considered above may in fact be microbiotherian can and should not be dismissed, their position remains contentious. Use of these taxa in the construction of phylogenetic and biogeographic scenarios should be qualified.

Previous morphology-based cladistic analyses.-Apart from tins study, only 3 investigations have addressed higher level phylogeny of Australian marsupicarnivores using parsimony-based analysis of anatomical data: Kirsch and Archer (1982), Springer et al. (1997), and Rougier et al. (1998). Kirsch and Archer (1982) found that because many anomalous phylogenies were produced, particularly at higher levels, the value of parsimony was called into question. Their study is of particular interest with regard to our own because it represents the only previous anatomy-based investigation that used species as the operational taxonomic unit. Kirsch and Archer (1982) clearly demonstrated that varying the number of taxa included could affect outcomes and inclusion of fossil taxa could significantly impact results. For example, they found that Thylacinus cynocephalus formed a monophyletic clade with the South American taxon Borhyaena tuberata when no other borhyaenids were included but that support for thylacinid-borhyaenid monophyly disappeared when additional borhyaenids were added. Additionally, they noted that no clades were founded on uniquely derived features; only unique combinations of features and sequences of change served to distinguish taxa in their analyses. Springer et al. (1997) presented both morphology- and molecule-based analyses. Their anatomy-based investigation included diprotodontian, marsupicarnivore, and peramelemorphian taxa, and in further contrast to the work of Kirsch and Archer (1982), they used the family rather than the species as the operational taxonomic unit. Their analysis produced a single with most-parsimonious tree dasyuromorphians plus notoryctids shown as the sister taxon to a clade inclusive of all living marsupials, other а result incongruous with all previous findings. In common with some recent molecular **Dromiciops** was studies. gliroides monophyletic with diprotodontians (Springer et al. 1997). No interordinal relationships were supported by bootstrap values -50%. Most recently, Rougier et al. (1998) presented a cladistic analysis including a wide range of nontherian, eutherian, and metatherian taxa, but only 3 of the 7 orders examined were extant. The results supported monophyly of D. gliroides and Dasyuridae. Sister taxa to this clade in decreasing order of relatedness were Didelphidae, Andinodelphys, Pucadelphys, Mayulestes, Borhyaenidae, and a monophyletic clade inclusive of 9 North American and Asian taxa. Elucidation of dasyuromorphian phylogeny was clearly not a prime objective of that analysis (neither thylacinids nor Myrmecobius were included), and relationships between dasyurids and non-Australian taxa were not discussed (Rougier et al. 1998). Consequently, we have restricted our comments on those findings. in determining their value However. affinities of Australian marsupicarnivores is limited because only dasyurids were included. As argued by Wroe (1999a, 1999b, in press) and supported by results of the present study, dasyurids, especially the modem subfamilies, constitute a specialized marsupial clade and may have been derived quite recently. Thus, they are unlikely to represent approximations of the "ancestral" Australian marsupial. There are a number of potential synapomorphies, especially in the basicranium, uniting dasyurids and D. gliroides. However, to acknowledge these features as synapomorphies uniting these 2 taxa demands that a suite of basicranial plesiomorphies present in thylacinids and myrmecobiids or numerous dental plesiomorphies in *D. gliroides* be consigned to the status of apomorphic reversals to a plesiomorphic condition.

Several possible explanations, including

inappropriate methods of analysis and exclusion of appropriate range of specimens, may account for failure of previous analyses to resolve dasyuromorphian phylogeny. Moreover, the characters themselves may be relatively uninformative, even when analyzed correctly within a cladistic framework. As shown by Kirsch and Archer (1982), inclusion of fossil taxa can significantly affect the outcome of parsimonybased analyses. Aided by access to highly significant new fossil material from Australian and South American deposits, we attempted to give added insight into the question of the evolution of Australian marsupicarnivores.

MATERIALS AND METHODS

Included taxa and scoring of characters .-Pucadelphys andinus, the least derived of South American marsupials known from near-complete cranial and dental data (Marshall and Muizon 1995), was treated as the outgroup. Thus, the remaining 6 South American taxa and the peramelemorphians were treated as ingroup taxa to test previous hypotheses of phylogeny. In all, representatives of the following possible sister taxa to Dasyuromorphia were included: Pucadelphydae, Didelphidae, Mayulestidae, Microbiotheriidae, and Peramelemorphia. Notoryctidae, another potential sister taxon, was not included because extreme specialization of cranial and dental features in this zalambdodont taxon produced a character profile consisting of >70% features that were either unknown or autapomorphic. Novacek et al. (1988) recommended that 30% of the features be scored for robust elucidation of relationships. However. new material representing a possible structurally intermediate notoryctemorphian, recently from Miocene recovered deposits of Riversleigh (M. Gott and M. Archer, in litt.), may be of value in future analyses. Diprotodontians were not included for 2 reasons. This taxon was not suggested as a potential sister to Dasyuromorphia, except as part of a monophyletic clade with Dromiciops gliroides (Kirsch et al. 1997), already included in the analysis. Also, the majority of characters, if scored for diprotodontians in the present study, would have constituted autapomorphies. A more detailed character analysis incorporating

extant and new nearly complete fossil diprotodontian taxa is in progress.

Polarity decisions regarding the majority of characters used m the present study based work. were previous on detailed treatment Consequently, was unnecessary except where there has not been general consensus. The character analysis is given in Appendix 1. Species were used as the operational taxonomic unit in this analysis for several reasons. First, the investigator introduces assumptions of phylogeny by using higher taxonomic units. Because the phylogenetic position of many marsupicaniivores is extremely contentious (for some species, this uncertainty extends to the ordinal level), such assumptions may confound the generation of accurate results. Second, because elucidation of relationships among new fossil species was a primary goal, they were treated at the specific level. Third, as observed by Simmons (1993), the enduring legacy of parsimony-based phylogenetic analyses is most likely to be the data. Because higher taxonomic units are certain to be redefined, with taxa added or removed in light of new evidence, data based at this level are sure to become redundant. Moreover, particularly where diverse and speciose taxa represent the subject material few investigators can claim to have studied all species referred to higher clades. These difficulties do not arise where data are tabulated at the specific level. An obvious drawback to the methodology applied here is that all available species could not be included. However, this limitation was overridden by the aforementioned advantages.

Seven South American species (3 fossil and 4 extant) and 17 Australian species (2 fossil, 1 recently extinct, and 14 extant) were included. The American taxa were andinus Pucadelphys (Pucadelphydae), Andinodelphys cochabambensis Didelphis (Pucadelphydae), marsupialis (Didelphidae). Metachirus naudicaudatus (Didelphidae), Lestodelphys halli (Didelphidae), Mayulestes ferox (Mayulestidae), and Dromiciops gliroides (Microbiotheriidae). Australian taxa were 3 peramelemorphians (Yarala burchfieldi, *Echymipera kalubu, and Perameles nasuta*) dasyuromorphians, and 14 i.e., Myrmecobius fasciatus (Myrmecobiidae), 2 thylacinids (Thylacinus cynocephalus and *Badjcinus turnbulli),* and 11 dasyurids. Dasyurid taxa comprised representatives from each of the 4 dasyurid subfamilies recognized in the present study: Barinya wangala (Barinyainae), Sminthopsis crassicaudata, S. murina, and Planigale maculata (Sminthopsinae), Murexia longicaudata, Phascogale tapoatafa, and Antechinus flavipes (Phascogalinae), and 4 dasyurines (Neophascogale lorentzii, Dasyurus hallucatus, Dasyurus maculata, and Sarcophilus harrisii). Only fossil taxa known on the basis of complete or nearly complete cranial and dental material were used. Outgroup taxa were the 3 exceptionally well-preserved Paleocene taxa from the Tiupampa Local Fauna of Bolivia (P. andinus, A. cochabambensis, and M. ferox). Australian fossil taxa included were the Miocene bandicoot taxon Y. burchfieldi and the only 2 pre-Pleistocene Australian dasyuromorphians represented bv significant cranial material (B. turnbulli and B. wangala).

Phylogenetic analysis. The data matrix included 24 taxa (6 fossil, 1 recently extinct, and 17 extant) and 77 characters (44 dental and 33 cranial). Analyses were conducted with PAUP 3.1.1 (Swofford 1993) using 50 replicates, ACCTRAN, and TBR branch swapping (random input order) and with characters unordered. Relative stability of clades was assessed using bootstrap (Felsenstein 1985) and Bremer support 1988. 1994) (Bremer analyses. Bootstrapping was based on 1,000 replicates, and pseudoreplicate bias was reduced by using multiple tree-space searches, with 3 trees sampled per iteration (Colgan et al. 1998; Reid 1996). Bremer support indices were determined bv studying suboptimal trees. Because different investigators have offered conflicting opinions regarding character states of some important features, only species that were examined directly by us were included in this study. Dental nomenclature follows Flower (1867) and Luckett (1993) regarding the molar-premolar boundary, such that the adult (unreduced) postcanine cheektooth formula of marsupials is P1-3 and M1-4. Systematic terminology was as used by Wroe (1996, 1997b), adapted from that of Marshall et al. (1990) and Krajewski et al. (1994). Institutional abbreviations are QM (Queensland Museum Fossil). AMNH (American Museum of Natural History), Paleontologie, NMHN (Laboratoire de Museum National d'Histoire Naturelle), and (Paleontology YPBF Pal collection of Yacimientos Petroliferos Fiscales de Bolivia in the Centro de Temologia Petrolera). Material examined included that used by Wroe (1997b) and holotype material of D. murgonensis (QMF31458), B. wangala (QMF31408), P. andinus (YPFB Pal 6105), A. cochabambensis (NMHN 8264), and M. ferox (NMHN 1249).

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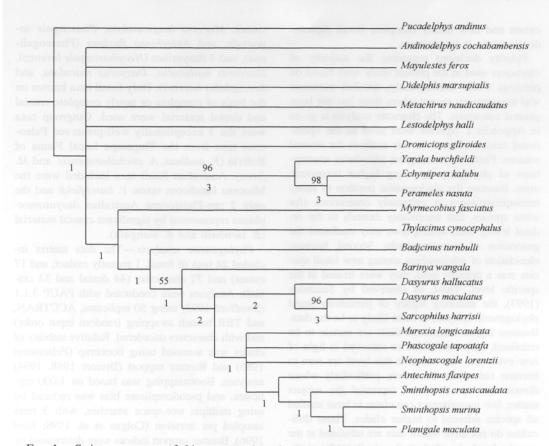


FIG. 1.—Strict consensus of 64 most-parsimonious trees for 24 taxa and 77 dental and cranial characters (length 245, consistency index = 0.4160, homoplasy index = 0.5840, and retention index = 0.6106). Bootstrap values >50% are shown above branches. Bremer support is given below branches.

RESULTS

Sixty-four most-parsimonious trees of were produced steps 245(excluding characters. uninformative consistency index = 0.4160. homoplasv index = 0.5840. retention index = 0.6106). The strict consensus tree and the bootstrap values and Bremer support indices for resolved nodes are given in Fig. 1. Character state reconstructions for the list of the 64 mostparsimonious trees are given in Appendix II for the nodes shown in Fig. 2. At the ordinal level, only Dasyuromorphia and Peramelemorphia formed natural groups (Fig. 1). No single outgroup taxon emerged as a clear sister to Dasyuromorphia. Within Dasyuromorphia, the positions of **Myrmecobius**

fasciatus and Thylacinus cynocephalus were unresolved with respect to a clade comprising the thylacinid Badjcinus turnbulli and dasyurids. Barinya wangala was the sister taxon to a monophyletic clade inclusive of all extant dasyurids. Among extant subfamilies of dasyurids, only Sminthopsinae was monophyletic. The taxon--character matrix is given in Table 1.

DISCUSSION

Our cladistic treatment of morphological features provided resolution for dasyuromorphian phylogeny that, in at least some important respects, agrees with many previously published phylogenies. However,

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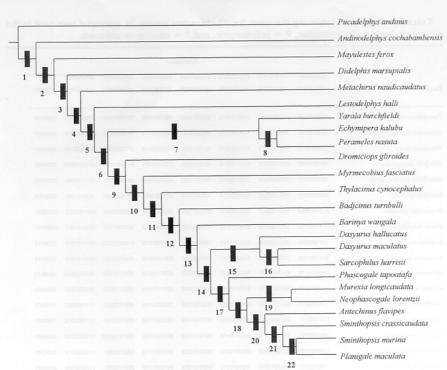


FIG. 2.--One of the 64 most-parsimonious trees of dasyuromorphians (see Appendix II for character state changes).

despite this correspondence, synapomorphies that actually support these clades often do not concur with those determined by subjective means used in most earlier anatomy-based works. Examples include cases where supporting synapomorphies, not previously suggested, appear in addition to those proposed in earlier works, e.g., only 4 of the 7 synapomorphies defining Dasyuridae (node 13, Fig. 2), changes in characters 61, 65, 66, and 70, were considered by Wroe (1999a). Examples also exist where previously proposed synapomorphies are not supported. These include development of a laterally extensive tubular ectotympanic (character 58), present in Thylacinus cynocephalus and Badjcinus turnbul*li*, as a synapomorphy for Thylacinidae

(Muirhead and Wroe 1998). However, the ectotympanic of an undescribed thylacinid from the Miocene of Riversleigh material (QMF36357) shows the same morphology as in T. cynocephalus and B. turnbulli, increasing the likelihood that this feature represents an actual thylacinid synapomorphy. Similar to Kirsch and Archer (1982), few groups are defined by unique synapomorand many characters phies, have undergone multiple reversals. Synapomorphies uniting clades some large numbers reversed include of characters. Of the 9 putative synapomorphies defining the B. turnbulli-dasyurid clade, 8 are reversals to a plesiomorphic state in dental features (see Appendix II). Overall, reversals to plesiomorphic states are more common in dental than cra-

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TABLE 1.—Data matrix showing the states for 77 characters among 24 marsupial taxa used in the present study: 0 = plesiomorphic, P = polymorphic, and ? = missing or unknown.

| | Characters | | | | | | | |
|------------------------------|------------|------------|------------|------------|------------|-------------|------------|---------|
| | 000000001 | 1111111112 | 2222222223 | 3333333334 | 444444445 | 5555555556 | 6666666667 | 7777777 |
| Taxon | 1234567890 | 1234567890 | 1234567890 | 1234567890 | 1234567890 | 1234567890 | 1234567890 | 1234567 |
| Pucadelphys andinus | 0000000101 | 0010000000 | 1000100010 | 0000011002 | 0007000010 | ???0000000 | 0000000110 | 0000000 |
| Andinodelphys cochabambensis | 0000000101 | 0010000000 | 1070100010 | 0000010002 | 0000101010 | 7770000710 | 0010000710 | 0000000 |
| Mayulestes ferox | 0000000112 | 0010000100 | 1000100010 | 0010011000 | 1110100010 | 7770030710 | 0000100710 | 0001001 |
| Didelphis marsupialis | 0010000000 | 0010000000 | 0000100010 | 1000021002 | 0100101001 | P110011000 | 00101100P3 | 0000000 |
| Metachirus naudicaudatus | 0010000111 | 0010000000 | 0000100010 | 1000011002 | 0000001010 | 0210021000 | 0011100000 | 0000000 |
| Lestodelphys halli | 0000000111 | 0011100010 | 0000100010 | 1000021002 | 2101001010 | 0110022000 | 1010121003 | 0000000 |
| Dromiciops gliroides | 0110000020 | 01011022u0 | 0000100000 | 0000021002 | 0001031010 | 0002022000 | 0011302012 | 0000001 |
| 'arala burchfieldi | 7770000111 | 0211100010 | 0000111111 | 1000021002 | 0002077010 | 0270001707 | 0010101017 | 0000111 |
| Echymipera kalubu | 1111000101 | 1211000000 | 0000011111 | 1000021001 | 0001001010 | 0100001000 | 0011101010 | 1110111 |
| Perameles nasuta | 0111001101 | 1211000010 | 0000011111 | 1000021001 | 0001031010 | 0200122000 | 0011121010 | 1110111 |
| lyrmecobius fasciatus | 1010100??? | 0777777777 | 7001110001 | 0010721007 | 0000110010 | 0000122000 | 0010213010 | 0000011 |
| arinya wangala | 1100010111 | 0010010010 | 1000110010 | 0000101002 | 0011171110 | 1000112000 | 1010321012 | 0000001 |
| minthopsis murina | 1100000111 | 0011011110 | 0000110710 | 0000001002 | 2002031110 | 0001122001 | 1111332012 | 1100001 |
| minthopsis crassicaudata | 1101000111 | 0010010010 | 0000111010 | 0000001000 | 0012031110 | 0001122001 | 1111332012 | 1100001 |
| lanigale maculata | 1000000121 | 0111111110 | 0000110n10 | 0011001002 | 2012031110 | 00012122001 | 1110332012 | 0000001 |
| furexia longicaudata | 1100000111 | 0010010010 | 0000010010 | 0000101002 | 001P011110 | 1002122001 | 1110332012 | 0000001 |
| hascogale tapoatafa | 1000000111 | 0010011010 | 1000010010 | 0000101002 | 0000001110 | 0001122001 | 1110332012 | 0000001 |
| ntechinus flavipes | 1000000111 | 0010010010 | 1000010010 | 0000101002 | 0012021110 | 0001122001 | 1110332012 | 1100001 |
| leophascogale lorentzii | 1110100101 | 0210000010 | 0000011010 | 0000101002 | 0010011P10 | 0002122001 | 1117332012 | 1000001 |
| asyurus hallucatus | 1000200111 | 0310011010 | 0000010010 | 0101111002 | 0021101110 | 00P1122001 | 1110332012 | 0000001 |
| asyurus maculatus | 1010200122 | 0310010011 | 0000110010 | 0111121102 | 0022111110 | P102122101 | 1110332012 | 0000001 |
| arcophilus harrisii | 1010210123 | 0311010011 | 0000110011 | 0112121102 | 1022121110 | 1101122201 | 1110332012 | 0000001 |
| adjeinus turnbulli | 1770777112 | 0210011110 | 0007010010 | 0000000000 | 0011117770 | 0100012100 | 0011100000 | 0000001 |
| hylacinus cynocephalus | 1010000113 | 0111112200 | 0001110010 | 0020111127 | 1102121111 | 2100112100 | 0011100000 | 0000001 |

nial features (19 versus 7). No reversals to plesiomorphic states occur in middle ear features at any branches in the strict consensus tree, a result consistent with the idea that complex adaptations of the middle ear are unlikely to he lost (MacPhee 1981).

Ambiguity of these results regarding the origin of Australian marsupicarnivores necessitates qualification in the erection of phylogenetic and biogeographic scenarios.

Andinodelphys, no special For relationship with any Australian taxon was supported. This finding is in keeping with the interpretation of Muizon et al. (1997), who on the basis of more complete material argued that this Tiupampan taxon was not australidelphian but didelphoid.

Our finding of monophyly for dasyuromorphians is consistent with results of all studies since those of Lowenstein et al. (1981) and Szalay (1982). However, both bootstrap values and Bremer support was low. Monophyly for the clade is based on acquisition of 9 synapomorphies (Appendix 11), but only 1 of these, reduction of upper incisor number, has been proposed previously (Aplin and Archer 1987; Marshall et al. 1990). None of these 9 synapomorphies represent uniquely derived features within Marsupialia. Thus, at present our ability to define Dasyuromorphia is limited to a handful of equivocal features. Moreover, perhaps the most important of these characters, incisor number, is unknown in significant but poorly represented fossil taxa not included in this analysis (e.g., *Djarthia murgonensis*).

Furthermore, our results lend no support to interpretations of dasyuromorphian relationships at the family level. In some previous anatomy-based studies (Aplin and Archer 1987; Archer 1984), but not in others (Wroe 1997b), Myrmecobius fasciatus has been suggested as the sister taxon to thylacinids and dasyurids. Among studies based on molecular evidence that have included *M. fasciatus*, Lowenstein et al. (1981) and Krajewski et al. (in press) supported the contention of plesiomorphic sister taxon status for Myrmecobiidae. Krajewski et al. (1997a) placed the numbat within Dasyuridae. However, this position was supported by very low bootstrap values. The finding that the 2 thylacinids included were not monophyletic is inconsistent with the results obtained by Muirhead and Wroe (1998). The position of Barinya wangala as a sister species to extant dasyurid subfamilies supports the conclusion of Wroe (1999a).

At lower taxonomic levels within Dasyuridae, relationships between some taxa were consistent with those of previous studies. Examples include monophyly for both *Dasyurus* and *Sarcophilus harrisii* (Archer 1982; Krajewski et al. 1994, 1997b) and Sminthopsinae (Archer 1982a; Krajewski et al. 1994), but other relationships were highly incongruent (e.g., paraphyly for Dasyurinae and Phascogalinae). Our results were perhaps not surprising given the long history of conflict over the intrafamilial relationships of dasyurids (see Krajewski et al. 1997b, in press; Wroe 1997b).

Monophyly was supported for a number of accepted natural groups that have previously proven resistant to resolution using cladistic analysis of anatomical data. These groups include Peramelemorphia, Dasyuromorphia, and Dasyuridae (including the fossil taxon *Barinya wangala*) and a clade inclusive of all extant dasyurids. In some instances, the actual status of many features identified as synapomorphies for these respective clades seems questionable, particularly regarding labile dental features. However, the standing of others, particularly basicranial apomorphies, is more concrete. Our results also support the argument of Wroe (1999a) that the common perception of extant dasyurids as Australian marsupial paradigms is no longer tenable.

Additional work remains to be done in the area of anatomy-based cladistic treatment of the origins of Dasyuromorphia, relationships among its constituent families, and intrafamilial affinities. Although monophyly for Dasyuromorphia and certain widely recognized clades was achieved in the present study, it is clear that most characters employed were inherently uninformative because there were very few nonhomoplastic synapomorphies at each node. Ongoing character analyses, inclusion of additional taxa, and new fossil discoveries will be of value in further analyses.

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LITERATURE CITED

- APLIN, K., AND M. ARCHER. 1987. Recent advances in marsupial systematics, with a new, higher level classification of the Marsupialia. Pp. xv-lxxii in Possums and opossums: studies in evolution (M. Archer, ed.). Surrey Beatty and Sons, Sydney, New South Wales, Australia.
- ARCHER, M. 1976a. The basicranial region of marsupicarnivores (Marsupialia), interrelationships of carnivorous marsupials, and affinities of the insectivorous marsupial peramelids. Zoological Journal of the Linnean Society 59:217-322.
- ARCHER, M. 1976b. The dasyurid dentition and its relationship to that of didelphids, thylacinids, borhyaenids (Marsupicarnivora) and peramelids (Peramelina: Marsupialia). Australian Journal of Zoology, Supplementary Series 39:1-34.
- ARCHER, M. 1981. Results of the Archbold expeditions. 104. Systematic revision of the marsupial dasyurid genus *Sminthopsis* Thomas. Bulletin of the American Museum of Natural History 168:63-223.
- ARCHER, M. 1982a. Review of the dasyurid (Marsupialia) fossil record, integration of data bearing on phylogenetic interpretation and suprageneric classification. Pp. 397--443 in Carnivorous marsupials (M. Archer, ed.). Royal Zoological Society of New South Wales, Sydney, New South Wales, Australia.
- ARCHER, M. 1982b. A review of Miocene thylacinids (Thylacinidae, Marsupialia), the phylogenetic position of the Thylacinidae and the problem of a priorisms in character analysis. Pp. 445-476 in Carnivorous marsupials (M. Archer, ed.). Royal Zoological Society of New South Wales, Sydney, New South Wales, Australia.
- ARCHER, M. 1984. The Australian marsupial radiation. Pp. 633-808 in Vertebrate zoogeography and evolution in Australasia (M. Archer and G. Clayton,

eds.). Hesperian Press, Sydney, New South Wales, Australia.

- ARCHER, M., H. GODTHELP, AND S. HAND. 1993. Early Eocene marsupial from Australia. Kaupia 3:193-200.
- BENSLEY, B. A. 1903. On the evolution of the Australian Marsupialia with remarks on the relationships of marsupials in general. Transcripts of the Linnean Society of London (Zoology) 9:83-217.
- BREMER, K. 1988. The limits of amino acid sequence data in angiosperm phylogenetic reconstruction. Evolution 42:795-803.
- BREMER, K. 1994. Branch support and tree stability. Cladistics 10:295-304.
- CIFELLI, R. L. 1993. Theria of metatherian-eutherian grade and the origin of marsupials. Pp. 205-215 in Mammal phylogeny; Mesozoic differentiation, multituberculates, monotremes, early therians, and marsupials (E S. Szalay, M. J. Novacek, and M. C. McKenna, eds.). Springer-Verlag, New York.
- CIFELLI, R. L., AND C. DE MUIZON. 1998. Marsupial mammal from the Upper Cretaceous North Horn Formation, Central Utah. Journal of Paleontology 72:532-538.
- COLE. A. 1998. Histone H3 RNA DNA sequences and arthropod molecular evolution. Australian Journal of Zoology 46:419-437.
- CROCHET, J. Y. 1980. Les marsupiaux du Tertiare d'Europe. Editions Foundation Singer-Polinac, Paris, France.
- FELSENSTEIN, J. 1985. Confidence limits on phylogenies: an approach using the bootstrap. Evolution 39: 783-791.
- FLOWER, W. H. 1867. On the development and succession of teeth in the Marsupialia. Philosophical Transcripts of the Royal Society of London 157:631-641.
- GAUDIN, T. J., J. R. WIBLE, J. A. HOPSON, ~ W. D. TURNBULL. 1996. Reexamination of the morphological evidence for the cohort Epitheria (Mammalia, Eutheria). Journal of Mammalian Evolution 3:31-79.
- GODTHELP, H., S. WROE, AND M. ARCHER. 1999. A new marsupial from the Early Eocene Tingamarra Local Fauna of Murgon, Southeastern Queensland: the prototypical Australian marsupial? Journal of Mammalian Evolution 6:289-313.
- GOIN, E J., AND A. A. CARLINI. 1995. An Early Tertiary microbiotheriid marsupial from Antarctica. Journal of Vertebrate Paleontology 15:205-207.
- HFRSHKOVITZ, P. 1992. Ankle bones: the Chilean opossum *Dromiciops gliroides* Thomas, and marsupial phylogeny. Bonner Zoologische Beitrage 43:181-213.
- HERSHKOVITZ, P. 1995. The staggered marsupial third lower incisor: hallmark of cohort Didelphimorphia, and description of a new genus and species with staggered i3 from the Albanian (Lower Cretaceous) of Texas. Bonner Zoologische Beitrage 45:153-169.
- HERSHKOVITZ, P 1999. *Dromiciops gliroides* Thomas, 1894, last of the Microbiotheria (Marsupialia), with review of the family Microbiotheriidae. Fieldiana: Zoology, New Series 93:1-60.
- JOHANSON, Z. 1996. Revision of the late Cretaceous

North American marsupial genus

- Alphadon. Palaeontographica 242:127-184. KIRSCH, J. A. W. 1977. The comparative serology of Marsupialia, and a classification of marsupials. Australian Journal of
- Zoology Supplementary Series 52: 1-152. KIRSCH, J. A. W., AND M. ARCHER. 1982. Polythetic cladistics, or, when parsimony's not enough: the relationships of carnivorous marsupials. Pp. 595-619 in Carnivorous marsupials (M. Archer, ed.). Royal Zoological Society of New South Wales, Sydney, New South Wales Australia.
- KIRSCH, J. A. W., A. W. DICKERMAN, A. R. OSVALDO, AND M. 5. SPRINGER. 1991. DNA hybridisation evidence for the Australasian affinity of the American marsupial Dromiciops gliroides. Proceedings of the National Academy of Sciences 88: 10465-10469.
- KIRSCH, J. A. W., E. LAPOINTE, AND M. S. SPRINGFR. 1997. DNA-hybridisation studies of marsupials and their implications for metatherian classification. Australian Journal of Zoology 45:211-280.
- KRAJEWSKI, C., L. BUCKLEY, AND M. WESTERMAN. 1997a. DNA phylogeny of the marsupial wolf resolved. Proceedings of the Royal Society of London, Series B 264:911-917.
- KRAJEWSKI, C., L. BUCKLEY, P. A. WOOLLEY, AND M. WESTFRMAN. 1996. Phylogenetic analysis of cytochrome b sequences in the dasyurid marsupial sub-family Phascogalinae: systematics and the evolution of reproductive strategies. Journal of Mammalian Evolution 3:81-91.
- KRAJEWSKI, C., L. BUCKLEY, AND M. WESTERMAN. 1994. Phylogenetic structure of the marsupial family Dasyuridae based on cytochrome b DNA sequences. Journal of Mammalian
- Evolution 2:25-35. KRAJEWSKI, C., S. WROE, AND M. WESTERMAN. In press. Molecular evidence for the phylogenetic relationships and the timing off cladogenesis in dasyurid marsupials. Zoological Journal of the
- Linnean Society. KRAJEWSKI, C., J. YOUNG, L. BUCKLEY, P. A. WOOLLEY, AND M. WESTERMAN. 1997b. Reconstructing the taxonomic radiation of dasyurine marsupials with cy-tochrome b, 12S RNA, and protamine PI gene trees. Journal of Mammalian Evolution 4:217-236.
- LOWENSTEIN, J. M., V. M. SARICH, AND B. J. ERICKSON. 198 1. Albumin systematics of the extinct mammoth and Tasmanian wolf. Nature 291:409-41 1.
- LUCKFTT, R W. 1993. An ontogenetic assessment of dental homologies in therian mammals. Pp. 182-284 in Mammal phylogeny; Mesozoic differentiation, multituberculates, monotremes, early therians, and marsupials (S. E Szalay, M. J. Novacek, and M. C. McKenna, eds.). Springer-Verlag, New York. LUCKETT, P. W. 1994. Suprafamilial
- relationships with- in Marsupialia: resolution and discordance from multidisciplinary data. Journal of Mammalian Evolution 2:255-283.
- MACPHEE, R. D. E. 198 1. Auditory regions of primates and eutherian insectivores: morphology, ontogeny and character analysis. Contributions to Primatology 18:1-282.
- MARENSSI, S. A., M. A. REGEURO, S. N. SANTILLANA, AND S. E VIZICNO. 1994. Eocene land mammals

from Seymour palaeobiogeographical Science 6:3-15.

Island, implications.

Antarctica: Antarctic

- MARSHALL, L. 0. 1977. Cladistic analysis of borhyaenoid, dasyuroid, didelphoid, and thylacinid (Marsupialia: Mammalia) affinity. Systematic Zoology 26: 410-425. MARSHALL, L. G. 1982. Systematics of the South
- Fieldiana family Microbiotheriidae. American
- (Geology) 10: 1-75. MARSHALL, L. G. 1987. Systematics of Itaboraian (middle Paleocene) age "opossum-like" marsupials from the limestone Quarry at 5Ao Jose de Itaborai, Brazil. Pp. 91-160 in Possums and opossums: studies in evolution. Vol. 1 (M. Archer, ed.). Surrey Beatty and Sons, Sydney, New South Wales, Australia.
- MARSHALL, L. G., J. A. CASF, AND M. O. WOOD]3URNE. 1990. Phylogenetic relationships of the families of marsupials. Pp. 433-505 in Current Mammalogy. Vol. 2 (H. H. Genoways, ed.). Plenum Publishing, New York.
- MARSHALL, L. G., AND C. DE MUIZON. 1995. *Pucadelphys andinus* (Marsupialia, Mammalia) from the early Paleocene of Bolivia. Part 11. The skull. Memoires du Museum National d'Histoire Naturelle 165:21-90.
- MUIRHEAD, J. 1992. A specialised thylacinid, *Thylacinus macknessi*, (Marsupialia: Thylacinidae) from Miocene deposits of Riversleigh, northwestern Queensland. Australian Mammalogy 15:67-76.
- MUIRHEAD, J. 1994. Systematics, evolution and palaeobiology of recent and fossil bandicoots Marsupialia: Peramelemorphia). Ph.D. dissertation, University of New South Wales, Sydney, New South Wales, Australia.
- MUIRHEAD, J. 1997. Two new thylacines (Marsupialia: Thylacinidae) from early Miocene sediments of Riversleigh, northwestern Queensland and a revision of the family Thylacinidae. Memoirs of the Queensland Museum 41:367-377.
- MUIRHEAD, J., AND S. FILAN. 1995. Yarala burchfieldi (Peramelemorphia) from Oligo-Miocene deposits of Riversleigh, northwestern Queensland. Journal of Paleontology 59:127-134.
 MUIRHEAD, J., AND S. WROE. 1998. A new genus and species, Badjcinus turnbulli (Thylacinidae:
- Marsupialia), from the late Oligocene of Riversleigh, northern Australia, and an investigation of thylacinid phylogeny. Journal of Vertebrate Paleontology 18: 612-626.
- MUIZON, C., DE. 1992. La fauna de mammiferos de Tiupampa (Paleoceno Inferior, Formacion Santa Lucia), Bolivia. Pp. 575-624 in Fossils y facies de Bolivia. Vol. 1. Vertebrados (R. Suarez-Soruco, ed.). Revista Teenica de Yacimientos Petroliferos Fiscales de Bolivia, Santa Cruz, Bolivia.
- MUIZON, C., DE. 1994. A new carnivorous marsupial from the Palaeocene of Bolivia and the problem of marsupial monophyly. Nature 370:208-21 1.
- MUIZON, C., DE. 1998. *Mayulestes ferox* a borhyaenoid (Metatheria, Mammalia) from the early Palaeocene of Bolivia: phylogenetic and paleobiologic implications. Geodiversitas 20:19-142.
- MUIŻON, C., DE, R. L., AND F. R. CESPEDA.

1997. The origin of the dog-like borhyaenoid marsupials of South America. Nature 389:486-489.

- NOVACEK, M. J., A. R. WYSS, AND M. C. MCKFNNA. 1988. The major groups of eutherian mammals. Systematics Association Special Volume 35B:31-71.
- RF.ID, A. M. 1996. Review of the Peripatopsidae (Onychophora) in Australia, with comments on peripatopsid relationships. Invertebrate Taxonomy 10:663--936.
- REIG, O. A., J. A. W. KIRSCH, AND L. J. MARSHALL. 1987. Systematic relationships of the living and Neocenozoic American "opossum-like" marsupials (suborder Didelphimorphia), with comment on the classification of these and the Cretaceous and Paleogene New World and European metatherians. Pp: I- 89 in Possums and opossums: studies in evolution. Vol. 1 (M. Archer, ed.). Surrey Beatty and Sons, Sydney, New South Wales, Australia.
- RIDE, W. D. L. 1964. A review of Australian fossil marsupials. Journal of the Royal Society of Western Australia 47:97-13 1.
- ROUGIER, G. W., J. R. WIBLE, AND M. J. NOVACEK. 1998. Implications of *Deltatheridium* specimens for early marsupial history. Nature 369:459-463.
- SIMMONS, N. B. 1993. The importance of methods: archontan phylogeny and cladistic analysis of morphological data. Pp. 1-61 in Primates and their relatives in phylogenetic perspective (R. D. MacPhee, ed.). Plenum Publishing, New York.
- SPRINGER, M. S., J. A. W. KIRSCH, AND J. A. CASE. 1997. The chronicle of marsupial evolution. Pp. 129-161 in Molecular evolution and adaptive radiation (T. J. Givnish and K. J. Sytsma, eds.). Cambridge University Press, Cambridge, Massachusetts.
- SPRINGER, M. S., M. WESTERMAN, AND J. A. W. KIRSCH. 1994. Relationships among orders and families of marsupials based on 12S ribosomal DNA sequences and the timing of the marsupial radiation. Journal of Mammalian Evolution 2:85-115.
- SWOFFORD, D. L. 1993. PAUP: phylogenetic analysis using parsimony. Version 3. I. Illinois Natural History Survey, Chainpaign, Illinois.
- SZALAY, F. S. 1982. A new appraisal of marsupial phylogeny and classification. Pp. 621-640 in Carnivorous marsupials (M. Archer, ed.). Royal Zoological Society of New South Wales, Sydney, New South Wales, Australia.
- SZALAY, E S. 1994. Evolutionary history of the Marsupialia and an analysis of osteological characters. Cambridge University Press, New York.
- SZALAY, E S., AND B. A. TROFIMOV. 1996. The Mongolian Late Cretaceous *Asiatherium*, and the early phylogeny and paleobiogeography of Metatheria. Journal of Vertebrate Paleontology 16:474-".
- THOMAS, R. H., W. SCHAFFER, AND A. C. WILSON. 1989. DNA phylogeny of the extinct marsupial wolf. Nature 340:465-467.
- WIBLE, J. R., AND J. A. HOPSON. 1995. HOM010gieS Of the prootic canal in mammals and non-mammalian cynodonts Journal of Vertebrate Paleontology IS: 331-356.
- WOODBURNE, M. O., AND J. A. CASE. 1996. Dispersal, vicariance, and the Late Cretaceous to Early Tertiary land mammal biogeography from South America to Australia. Journal of Mammalian Evolution 3:121-161.

- WROE, S. 1996. *Muribacinus gadiyuli* (Thylacinidae, Marsupialia), a very plesiomorphic thylacinid from the Miocene of Riversleigh, Northwestern Queensland, and the problem of paraphyly for the Dasyuridae. Journal of Paleontology 70:1032-1044.
- WROE, S. 1997a. *Mayigriphus orbus,* a new genus and species of dasyuromorphian (Marsupialia) from the Miocene of Riversleigh, northwestern Queensland. Memoirs of the Queensland Museum 41:439-448.
- WROE, S. 1997b. A reexamination of proposed morphology-based synapomorphies for the families of Dasyuromorphia (Marsupialia): Part 1, Dasyuridae. Journal of Mammalian Evolution 4:19-52.
- WROE, S. 1998. A new species of 'bone-cracking' dasyurid (Marsupialia) from the Miocene of Riversleigh, northwestern Queensland. Alcheringa 22: 277-284.
- WROE, S. 1999a. The geologically oldest dasyurid (Marsupialia), from the Miocene Riversleigh, northwestern Queensland. Palaeontology 42:1-27.
- WROE, S. 1999b. Killer kangaroos and other murderous marsupials. Scientific American 280:68-74.
- WROE, S. In press. Australian marsupial carnivores: an overview of recent advances in palaeontology. In Predators with pouches (M. Jones, C. Dickman, and M. Archer, eds.). CSIRO Publishing, Melbourne, Victoria, Australia.
- WROE, S., J. BRAMMALL, AND B. N. COOKF. 1998. The skull of *Ekaltadeta ima* (Marsupialia: Hypsiprymnodontidae?): an analysis of some cranial features among marsupials and a re-investigation of propleopine phylogeny, with notes on the inference of carnivory in mammals. Journal of Paleontology 72: 738-751.

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Appendix I

Character analysis

1. Upper incisor number.-5 (0); 4 (1). See Archer(1976b), Marshall et al. (1990), and Wroe (1999a).

2. Shape of upper incisors.-Peg shaped (0); spatulate (1). Szalay (1994) proposed the presence of spatulate incisors as an australidelphian synapomorphy. In their character state matrix dealing with American taxa, Reig et al. (1987) score only microbiotheriids as showing spatulate upper incisor morphology. However, Archer (1976b) noted that both spatulate and peg- shaped upper incisors are found in representatives of Australidelphia and Ameridelphia. Thus, apomorphic reversal to a plesiomorphic state must have occurred within both clades regard- less of the polarity accepted. Nevertheless, each of these authors and Wroe (1999a) agree that

peglike morphology is plesiomorphic for marsupials.

3. Height of 11.-Not hypsodont (0); hypsodont (1). See Archer (1976b), Reig et al. (1987), and Wroe (1999a).

4. Morphology of Cl.--Caniform (0); premolariform (1). See Archer (1976b).

5. Height of P3.-Higher crowned than P2 (0); lower crowned than P2 (1); absent (2). See Archer (1976b), Reig et al. (1987) and Wroe (1997b, 1999a).

6. Shape of P3.-Laterally compressed in occlussal view (0); bulbous and ovate in occlussal view (1). See Wroe (1998).

7. Posterolingual cuspule present or absent on P3.-Absent (0); present (1). See Wroe (1996).

8. Relative size of paracone and metacone M1-3.-Paracone and metacone equal or almost equal in size to metacone (0); metacone much larger than paracone (1). See Cifelli (1993), Godthelp et al. (1999), and Szalay and Trofimov (1996).

9. Metacone on M4.-Present and distinct from metastylar corner of tooth (0); present but not distinct from metastylar corner of tooth (1); absent (2). See Wroe (1999a).

10. Shape and orientation of the centrocrista.-Centrocrista straight, with apex of postparacrista and premetacrista oriented dorsoventrally and terminating dorsally at or almost level with the talon basin (0); apex of centrocrista oriented buccally and well above talon basin, with an acute angle evident between the postparacrista and premetacrista and a distinct V shape in occlusal view (1); apex of centrocrista oriented buccally and well above protocone basin, with an oblique angle apparent between the postparacrista and premetacrista (2); apex of centrocrista positioned well above protocone basin with linear centrocrista (3).

The character states centrocrista straight or linear versus V-shaped and predilambdodont versus dilambdodont have been used synonymously by some authors (Marshll et al. 1990; Reig et al. 1987), although there is a distinction (Cifelli 1993; Johanson 1996). In practical terms, this is an academic point with regard to the investigations concerned because in the taxa under consideration a V-shaped centrocrista is intrinsic to dilambdodonty sensu Crochet (1980; i.e., preparacrista + centrocrista + postmetacrista form a W in occlussal view). Johanson (1996) provided a very useful and more thorough de-

of dilambdodonty scription predilambdodonty, aspects of which are incorporated here. But for the range of taxa included in the present study, further refinement is required because a mosaic of features evident in some carnivorous marsupials precludes their consideration as dilambdodont either strictly or predilambdodont (sensu Johanson 1996). In potential outgroup taxa to Marsupialia (e.g., *Deltatheridium)*, some generalized marsupials of uncertain affinity (e.g., Aenigmadelphys archeri), and most peradectids, a linear centrocrista is always concurrent with both the presence of an apex of the centrocrista, which is directed dorsally, and either very slight or no clear distinction in height between the tri- gon and talon basins. This combination of features was included under the category predilambdodonty by Johanson (1996). However, in peradectids and generalized australidelphian and ameridelphian shaped marsupicarnivore taxa, a Vcentrocrista is always accompanied by other features incorporated under the category dilambdodonty by Johanson, i.e., presence of a bucally oriented apex and a clear distinction in height between the trigon and talonon basins. In derived carnivorous marsupials (e.g., Thylacinus, Sarcophilus, Borhyaena), the centrocrista is linear in occlusal view but the trigon and talon basins are not of nearly equal height, in contrast to the condition in generalized centrocrista taxa with linear (e.g., peradectids). In thylacinids, a distinct morphocline is evident supporting the notion that the linear centrocrista in Thylacinus was secondarily derived from a dilambdodont condition as is present in basal thylacinid taxa (e.g., Muribacinus). The centrocrista in Muribacinus forms a distinct V shape; however, the inner angle of the V is obtuse in contrast to that of generalized ameridelphians and australidelphians. The size of this angle increases among other thylacinid taxa, culminating in the linear centrocrista of Thylacinus.

1 *11. Proximity of apex of centrocrista to ectoloph in dilambdodont* taxa.-Does not approach or breech ectoloph (0); approaches or breeches ectoloph (1). See Muirhead and Filan (1995).

12. Orientation of preparacrista on M1.-M 1 preparacrista forms a nearly perpendicular angle with respect to the long axis of the tooth (0); M1 preparacrista oriented anterobuccally relative to long axis of the tooth (1); M1 preparacrista runs posterobuccally relative to long axis

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of the tooth (2); M 1 preparacrista absent (3). See Muirhead and Wroe (1998).

13. Relative lengths of M3 and M4 preparacristae.-M4 preparacrista shorter than or equal to that of M3 (0); M4 preparacrista longer than that of M3 (1). See Godthelp et al. (1999).

14. Protoconule present or absent.-Present (0); absent (1). See Archer (1976b), Godthelp et al. (1999), Reig et al. (1987), and Wroe (1999a).

15. Metaconule present or absent.-Present (0); absent (1). See Archer (1976b), Godthelp et al. (1999), and Reig et al. (1987).

16. Stylar cusp A distinct or indistinct from parastylar corner of tooth.-Distinct (0); indistinct (1). See Archer (1976b).

17. Size of stylar cusp B on M3.-Large (0); small (1); absent (2). See Wroe (1996).

18. Size of stylar cusp D on M3.-Large (0); small (1); absent (2). See Wroe (1996) and Wroe (1997b).

19. Relative size of stylar cusp B and stylar cusp D on M2.-Stylar cusp B > stylar cusp D (0); stylar cusp D > stylar cusp B > (1). See Wroe (1996).

20. Relationship of stylar cusp D to metacone.-Not oppressed (0); oppressed (1). See Ride (1964) and Wroe (1996).

21. Presence or absence of stylar cusp C.-Stylar cusp C absent (0); stylar cusp C present (1). See Godthelp et al. (1999), Marshall et al. (1990), and Wroe (1999a).

22. 'Central cusp'.-Absent (0); present (1). See Godthelp et al. (1999).

23. Twinned cusps in the C position.-Absent (0); present (1). See Godthelp et al. (1999).

24. Presence or absence of anterior cingulum on M1.-Present (0); absent (1). See Muirhead and Wroe (1998).

25. Presence or absence of posterior cingulum on *M1.*-Present (0); absent (1). See Muirhead and Wroe (1998).

26. Lower incisor number.-4 (0); 3 (1). See Archer (1976b), Marshall and Muizon (1995), and Wroe (1999a).

27. i3 bilobed or not bilobed.-Not bilobed (0); bilobed (1). See Archer (1976b) and Muirhead and Filan (1995).

28. Position of hypoconulid.-positioned posterolingual to entoconid (0); positioned posterior to entoconid (1). See Muirhead and Filan (1995).

29. i3 staggered or not staggered.-Not stag-

gered (0); staggered (1). See Hershkovitz (1995).

30. Presence or absence of hypoconulid notch.-Present (0); absent (1). See Wroe (1997a, 1997b).

31. Presence or absence of well-developed sulcus formed by anterior cingulid.-Absent (0); present (1). See Wroe (1996).

32. Size of metaconid on m1 relative to that of m2-4.-Metaconid in ml not reduced relative to m2-4 (0); metaconid reduced relative to m2- 4 (1). See Wroe (1996).

33. Size of metaconid in m2-4.-Large (0); reduced (1); absent (2). See Wroe (1996).

34. Size of paraconid in m1.-Large (0); reduced (1); absent (2). See Archer (1976b) and Wroe (1999a).

35. Orientation of postprotocristid with respect to long axis of dentary.-Transverse (0); oblique (1). See Archer (1976b), Wroe (1999a).

36. Size of posterior cingulid in m1-3.-Posterior cingulid present and well developed (0); posterior cingulid reduced (1); posterior cingulid absent (2). See Muirhead and Wroe (1998).

37. Posterior cingulid in m4 present or absent.-Present (0); absent (1). This feature has been treated as separate from character 37 be- cause loss of a posterior cingulid on m4 occurs in many taxa without concomitant reduction or loss of the posterior cingulid in m1-3.

38. Presence or absence of 'carnassial notch' in cristid obliqua.-Absent (0); present (1). See Muirhead and Wroe (1998).

39. Morphology of postprotocristid in m3-4.-Postprotocristid continuous with metacristid (0); postprotocristid oriented posteriorly and almost continuous with cristid obliqua (1); postprotocristid oriented posteriorly and continuous with cristid obliqua (2). See Muirhead and Wroe (1998).

40. Anterior point of termination of the cristid obliqua in m3 with respect to carnassial notch formed by postprotocristid and metacristid.- Beneath carnassial notch (0); lingual to carnassial notch (1); buccal to carnassial notch (2). See Archer (1976b) and Godthelp et al. (1999).

41. Entoconid size.-Large (0); reduced (1); absent (2). See Archer (1976b) and Wroe (1999a).

42. Anteroposterior dimension of m4 relative to that of m3.-m4 < m3 (0); m4 > m3 (1). See Muirhead and Wroe (1998).

43. Height of p3 relative to p2.--p3 higher

crowned than p2 (0); p3 smaller than p2 (1); p3 absent (2). See Archer (1976b) and Wroe (1996, 1997b).

*44. Number of cusps on m4 talonid.-*3 cusps (0); 2 cusps (1); 1 cusp (2). See Archer (1976b).

45. Presence or absence of orbital crest.-Absent (0); present (1). See Muirhead and Wroe (1998).

46. Morphology of fossa for the lower canine.-Bordered by anterolateral process of maxilla (0); bordered by anterolateral processes of premaxilla and maxilla (1); bordered by anterolateral process of premaxilia (2); no lateral process (3). See Muizon (1998).

47. Presence or absence of palatal vacuities.-Absent (0); present (1). See Marshall and Muizon (1995).

48. Presence or absence of posterolateral palatine foramen.-Present (0); absent (1). See Archer (1984) and Wroe (1997b).

49. Presence or absence of accessory posterolateral palatine foramen.-Present (0); absent (1). See Wroe (1997b).

50. Contribution of alisphenoid and periotic to primary foramen ovale.-Delimited by alisphenoid anteriorly and periotic part of the petrosal posteriorly (0); delimited by alisphenoid only (1). See Gaudin et al. (1996) and Wroe (1997b).

51. Presence or absence of secondary foramen ovale formed by anteriorly directed strut of alisphenoid tympanic wing.-Absent (0); present but incomplete (1); present and complete (2). See Wroe (1997b).

52. Presence or absence of secondary foramen ovale formed by mesial fold in alisphenoid tympanic wing.-Absent (0); present but incomplete (1); present and complete (2). See Wroe (1997b).

53. Presence or absence of contribution to secondary foramen ovale by posteriorly directed strut in alisphenoid. Absent (0); present but in- complete (1); present and complete (2). See Wroe (1997b).

54. Morphology of ventral facial nerve canal-Absent (0); present but completed anteriorly by squamosal only (1-); present and enclosed within periotic with squamosal contribution to ventral rim (2); present and enclosed wholly within periotic (3). See Archer (1976a) and Wroe (1999a).

55. Size of squamosal epitympanic sinus.-

Absent (0); present (1). See Archer (1976a), Wroe (1999a), and Wroe et a]. (1998).

56. Alisphenoid hypotympanic sinus.-Absent (0); present with periotic component (1); present wholly within alisphenoid, i.e., separated from alisphenoid hypotympanic sinus by distinct petrosal ridge (2); present with alisphenoid, periotic, and squamosal contributions (3). See Archer (1976a).

57. Size of alisphenoid tympanic wing.-Absent (0); poorly developed, i.e., with alisphenoid hypotympanic sinus visible in external view (1); well developed, i.e., extends posteriorly to reach posterior limit of alisphenoid contribution of alisphenoid hypotympanic sinus in ventral view (2). See Archer (1976a), Wroe (1999a), and Wroe et al. (1998).

58. Shape of ectotympanic.-Simple U shape (0); laterally extensive, but simple U shape in lateral view (1); laterally extensive, complex saddle shape (2). See Archer (1976a) and Wroe et al. (1998).

59. Medial process of the squamosal-Absent (0); present (1). See Muizon (1998) and Muizon et al. (1997).

60. Length of the internal jugular canal-Does not extend anteriorly to basisphenoid (0); extends to basisphenoid (1). See Archer (1976a) and Wroe (1999a).

61. Presence or absence of a well-developed posteroventral lip formed by a mesially directed process in the pars petrosa, enclosing the internal jugular ventrally.-Absent (0); present (1). See Archer (1976a) and Wroe (1999a).

62. Dorsal enclosure of internal jugular canal-Not fully enclosed dorsally (0); fully enclosed dorsally (1). See Wroe (1999a).

63. Presence or absence of transverse canal.-Absent (0); present (1). See Archer (1976a) and Wroe (1999a).

64. Frontal-squamosal or alisphenoid-parietal contact-Alisphenoid-parietal contact (0); frontal-squamosal contact (1). See Muizon @1998) and Wroe (1999a).

65. Morphology of tympanic wing of periotic.-Absent (0); present but does not contact alisphenoid tympanic wing (1); present and con- tacts alisphenoid tympanic wing but does not form periotic hypotympanic sinus (2); present and contacts alisphenoid tympanic wing and floors periotic hypotympanic sinus (3). See Archer (1976a) and Wroe (1999a).

66. Morphology of paroccipital tympanic pro-

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cess.-Absent (0); present but does not contact any other tympanic elements (1); present and contacts the mastoid tympanic process laterally (2); present and fused to the pars petrosa anteriorly and mastoid tympanic process laterally (3). See Archer (1976a) and Wroe (1999a).

67. Morphology of mastoid tympanic process.-Absent (0); present (1); present and con- tacts pars petrosa (2); present and contacts alisphenoid tympanic wing (3). See Archer (1976a) and Wroe (1999a).

68. Presence or absence of mastoid contribution to occiput.-Present (0); absent (1). See Muizon (1998).

69. Morphology of foramen lacerum medium.-Distinct from primary foramen ovale (0); confluent with primary foramen ovale (1). See Marshall and Muizon (1995), Muizon (1998), and Wroe et al. (1998).

70. Morphology of tubal foramen.-Absent (0); present but slitlike and incomplete anteroventrally (1); present and ovoid, with sulcus in pars petrosa (2); present, with sulcus in alisphenoid tympanic wing (3). See Wroe (1999a).

71. Shape of nasals.-Posteriorly expanded (0); not posteriorly expanded (1). See Archer (1981, 1982a).

72. Maxilla-nasal contact.-Maxilla-nasal contact longer than premaxilla-nasal contact (0); premaxilla-nasal contact longer than maxilla- nasal contact (1). See Muirhead (1994).

73. Posterior extension of nasals.-Extend posteriorly beyond the anterior rim of the orbit (0); do not extend posteriorly beyond level of the orbit (1). See Muirhead (1994).

74. Frontal-maxillary contact.-Present *(0);* absent (1). See Archer (1982b).

75. Morphology of jugal-Jugal not Y-shaped (0); jugal Y-shaped (1). See Muirhead (1994).

76. Morphology of antorbital fossa.-Antorbital fossa absent (0); antorbital fossa small or incomplete (1); antorbital fossa deep (2). See Muirhead (1994).

77. Presence or absence of prootic canal.-Present (0); absent (1). See Wible and Hopson (1995) and Muizon et al. (1997). A prootic canal was observed in 2 dasyurid specimens, *Antechinus* (AMNH 183455) and *M. longicaudata* (AMNH 198721), by J. R. Wible (pers. commun.). Presence of this feature is certainly rare among dasyurids and has not been observed in any other specimens studied by J. R. Wible (pers. comm.), Archer (1976a), or us.

Appendix II

Character state changes for nodes 1-22 in 1 of the 64 equally parsimonious trees

Node 1.-C59 0 ---> 1. *Node 2.-*C9 0 ---> 1; C42 0 ---) 1; C56 0 ---> 1; C65 0 ---> 1. Node 3.-C21 1 ---> 0; C31 0 ---> 1; C36 1 2; C47 0 ---> 1; C57 0 ---> 1; C63 0 ---> 1; C69 1 ---> 0. Node 4.-C3 0 ---> 1; C42 1 ---> 0., C45 1 ---> 0; C52 1 ---> 2; C56 1--> 2. *Node 5.-*C 14 0-->1; C19 0-->1; C44 0 ----> 1; C57 1 --- > 2; C67 0 ---> 1. Node 6.-C2 0--> 1; C12 0--> 1; C26 0--> 1; C46 0 ---> 3., C53 1--> 0; C69 0-->1; C77 0--> 1. Node 7.-CI2 1 ----> 2; C27 0 ---> 1; C28 0 ---> 1; C30 0 --.> 1; C 56 2 --> 0; C57 2 ---> 1; C75 0--> 1; C76 0 ---> 1. *Node 8.*-C4 0 ---> I., C9 1 --> 0; C11 0 ---> 1; C15 1 --> 0; C25 1 ---> 0; C40 2 ---> 1; C64 0 1; C71 0 --> 1; C72 0 ---> 1; C73 0 ---> 1. Node 9.-CI7 0 --> 2; C18 0--> 2; C29 1-->0; C31 1 ---> 0; C52 2 --> 0. *Node 10.-*C1 0-->1; C2 1 ---> 0; C16 0 ---> 1; C24 0 --> 1; C33 0-->1; C35 0-->1; C45 0 --- > 1; C46 3 --> 1; C55 0 --> 1. Node 11.-C29 0 ---> 1; C36 2-->1; C48 0--> 1, C52 0 --> 1; C56 2 ---> 1; C58 0 ---> 1. Node 12.--C3 1 ---> 0; C12 1 ---> 0; C14 1 0; C15 1 ---> 0; C17 2--> 0; C18 2 --> 0; C24 1 --> 0; C25 1 ---> 0; C43 0 ---> 1. Node 13.-C33 1 ---> 0; C52 1 ---> 0; C58 1--> 0: C61 0 ---> 1: C65 1 --> 3: C66 0 --> 2: C70 0--> 2 Node 14.-C54 0 ---> 1; C56 1 ---.> 2; C60 0--> 1; C62 0 ---> 1; C66 2 ---> 3; C67 1 ---> 2. Node 15.-C5 0 ---> 2; C12 0 ---> 3; C32 0 1; C34 0 ---> 1; C36 0 ---> 1; C43 1 ---> 2. Node 16.-C3 0-->1; C9 1 --- > 2; C10 1 ---> 2; C20 0--> 1; C25 0--> 1; C33 0 ---> 1; C36 1 --> 2; C38 0 ---> 1; C44 1 --..> 2; C51 0 --> 1; C52 0-->1; C58 0 --> 1. Node 17.-C44 0 ---> 1; C45 1 ---> 0. *Node 18.-*C2 0 ---> 1; C71 0 --> 1. Node 19.-C54 1 --- > 2. Node 20.-C44 0 -->2; C46 1 ----> 2; C72 0 ---1 *Node 21.-*C25 0-->1; C35 1--> 0; C46 2-->3; C64 0 ---> 1. Node 22.-CI4 0-->1; C17 0-->1; C18 0--> 1; C41 0 ---> 2.