

# Nanotechnology in the life sciences

## A FRONTIS LECTURE SERIES

organized by

**Pieter Stroeve**

**Department of Chemical Engineering and**

**Materials Science**

**University of California, Davis**

**Davis, CA 95616, USA**



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# Nanotechnology in the life sciences

February 13	13:30	Pieter Stroeve (UC Davis)- Size, measurement and sensing
	14:30	Mieke Kleijn (WUR)- Surface forces using AFM
<b>February 20</b>	<b>13:30</b>	<b>Pieter Stroeve- (Bio)materials</b>
	<b>14:30</b>	<b>Ernst Sudholter (WUR)- Hybrid organic semiconductor FETs</b>
February 27	13:30	Pieter Stroeve- Self assembling molecular structures
	14:30	Richard Schasfoort (U Twente)- Surface modification and microfabrication strategies
Friday, March 5	13:30	Pieter Stroeve- Environment
	14:30	Keurentje (TU Eindhoven)- Micellar systems for nanoscale engineering of reaction and separation processes
Friday, March 12	13:30	Pieter Stroeve- Life sciences and medicine
	14:30	Ton Visser (WUR)- Single-molecule fluorescence in microfluidic devices

# Nanotechnology: (bio)materials

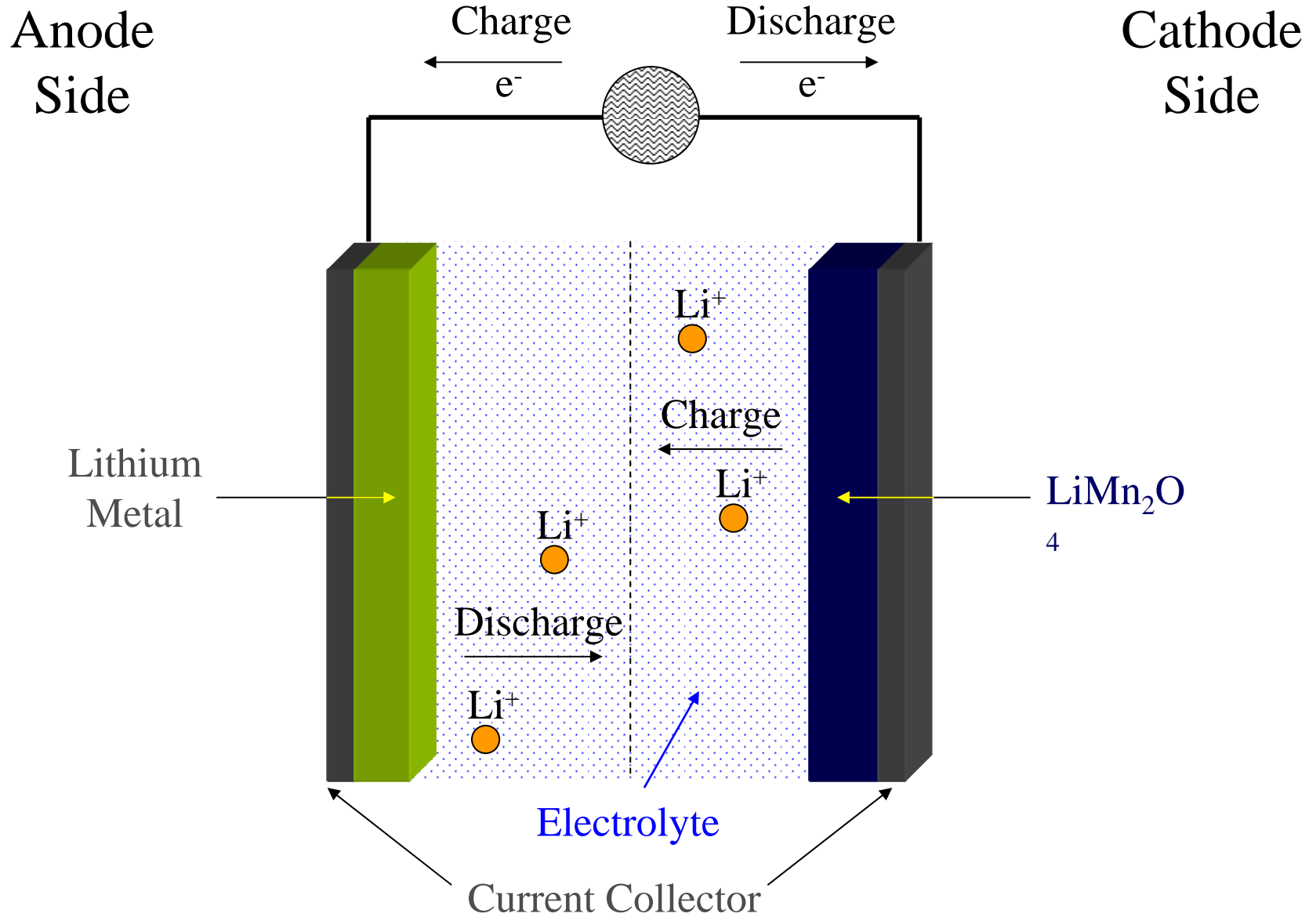
- **Rocket science?**
- **Engineered proteins for nanotechnology**
- **GeneChip**
- **Nanotubes, nanowires, nanocables**
- **Protein separation in nanopores**



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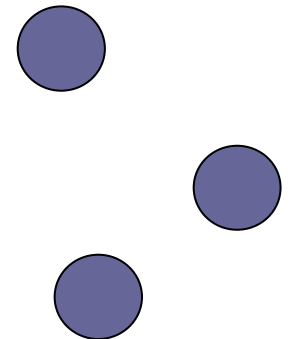
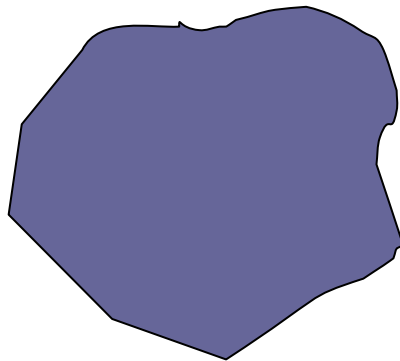
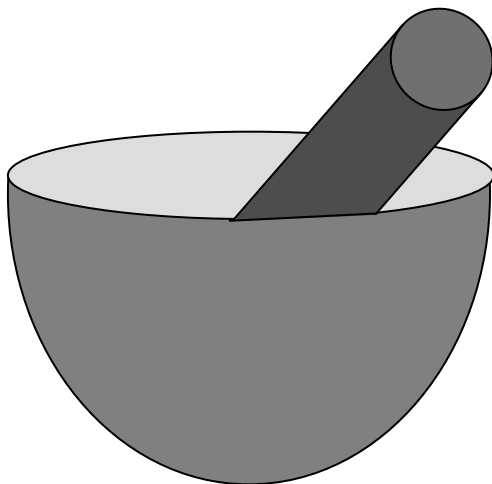
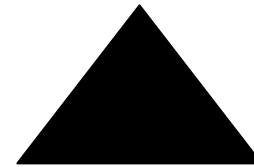
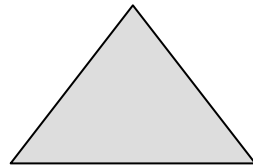
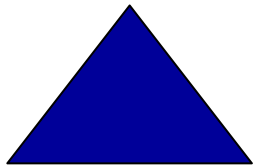
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# Lithium ion battery



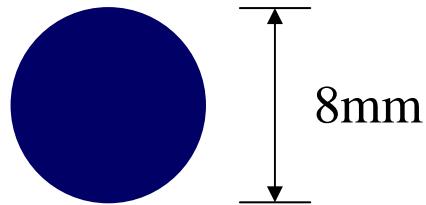
# Creation of $\text{LiMn}_2\text{O}_4$ Cathode

$\text{LiMn}_2\text{O}_4$  80 wt% (powder, active material)    PTFE 5 wt% (binder)    Acetylene Black 15 wt% (conductive material)



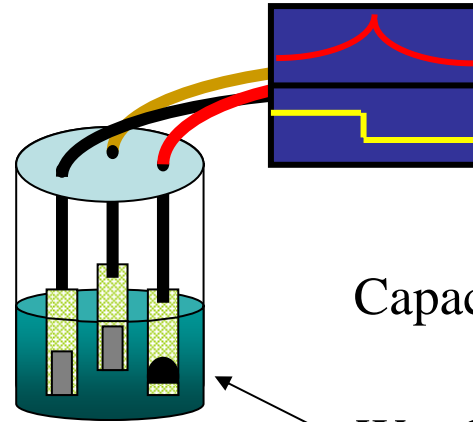
# Experimental stages

1



As Prepared

2

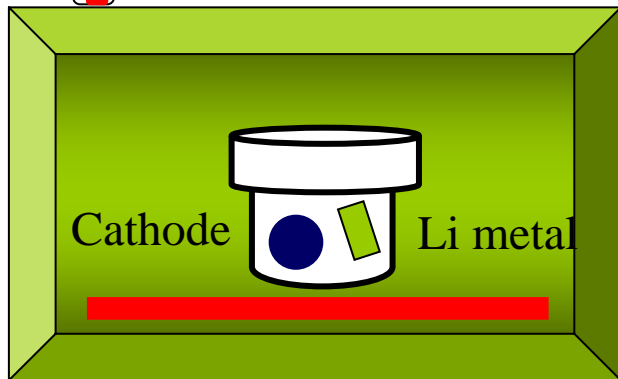
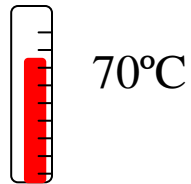


Capacity Measurement

Wet Cell

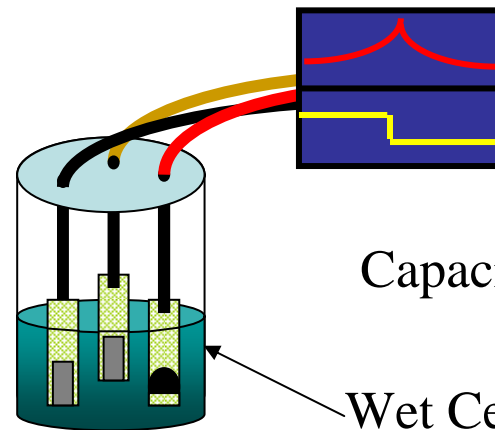
Before Storage

3



Store in oven 5 Days

4

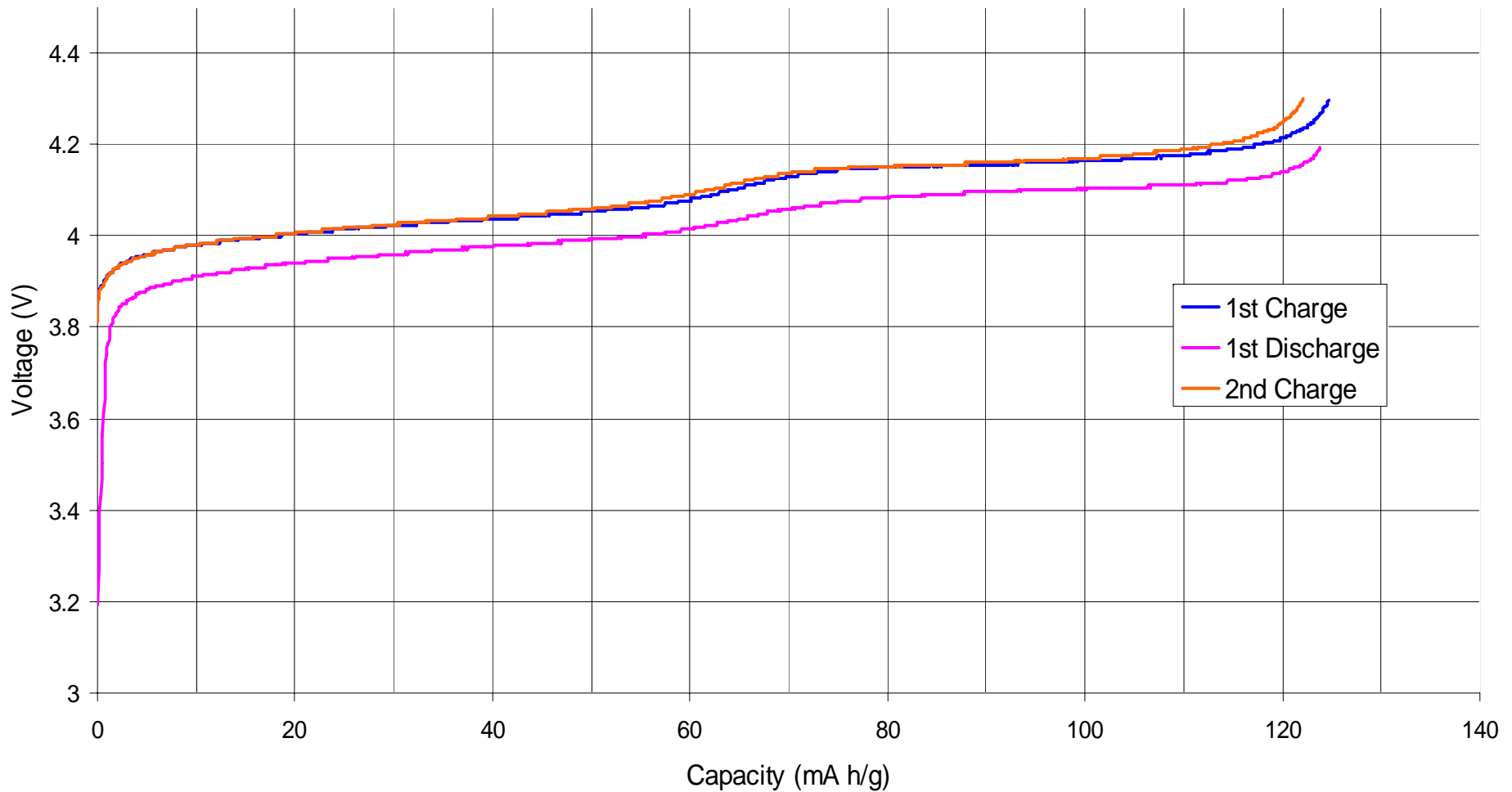


Capacity Measurement

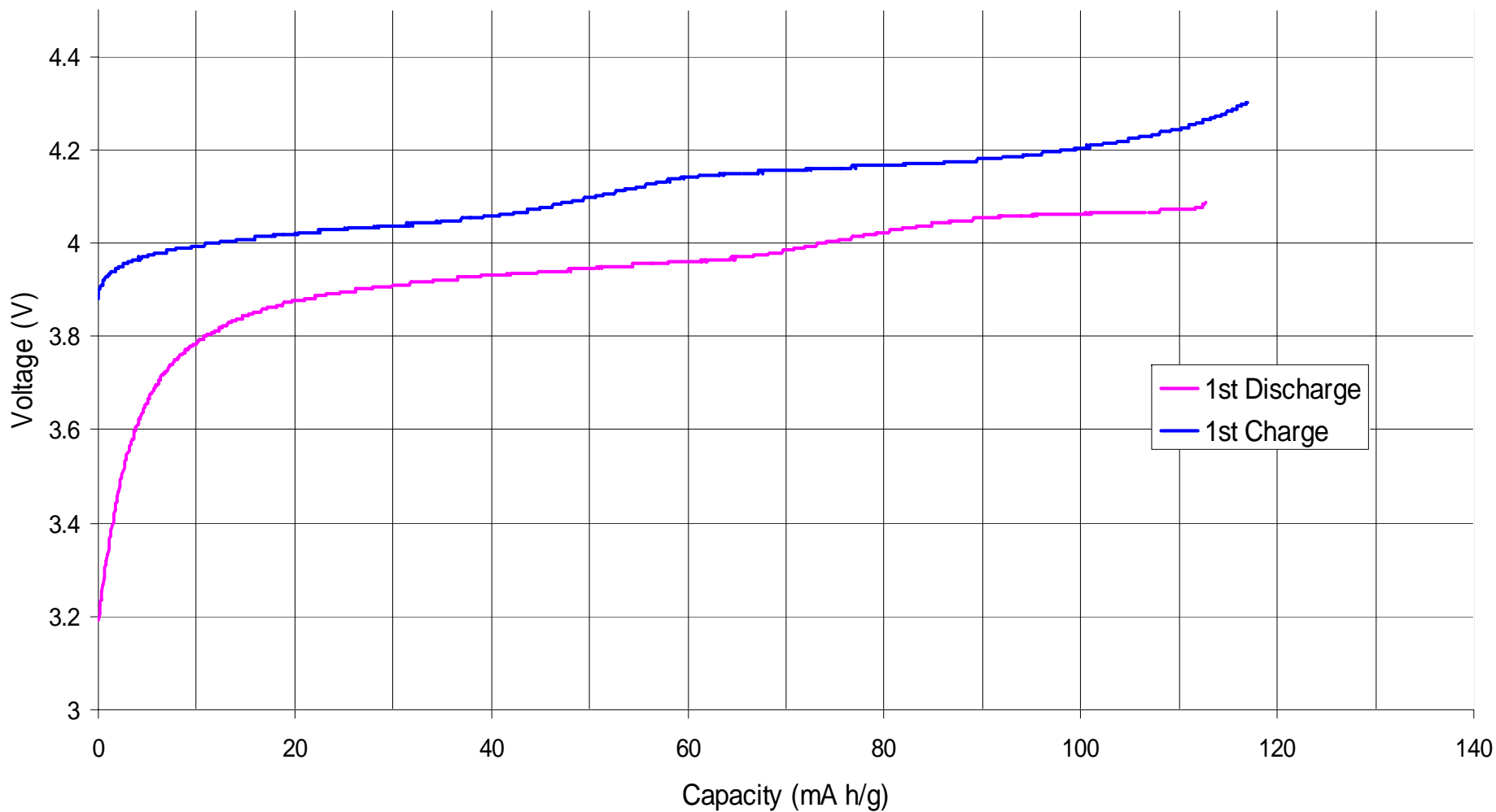
Wet Cell

After Storage

# Capacity Measurement of $\text{LiMn}_2\text{O}_4$ Before Storage



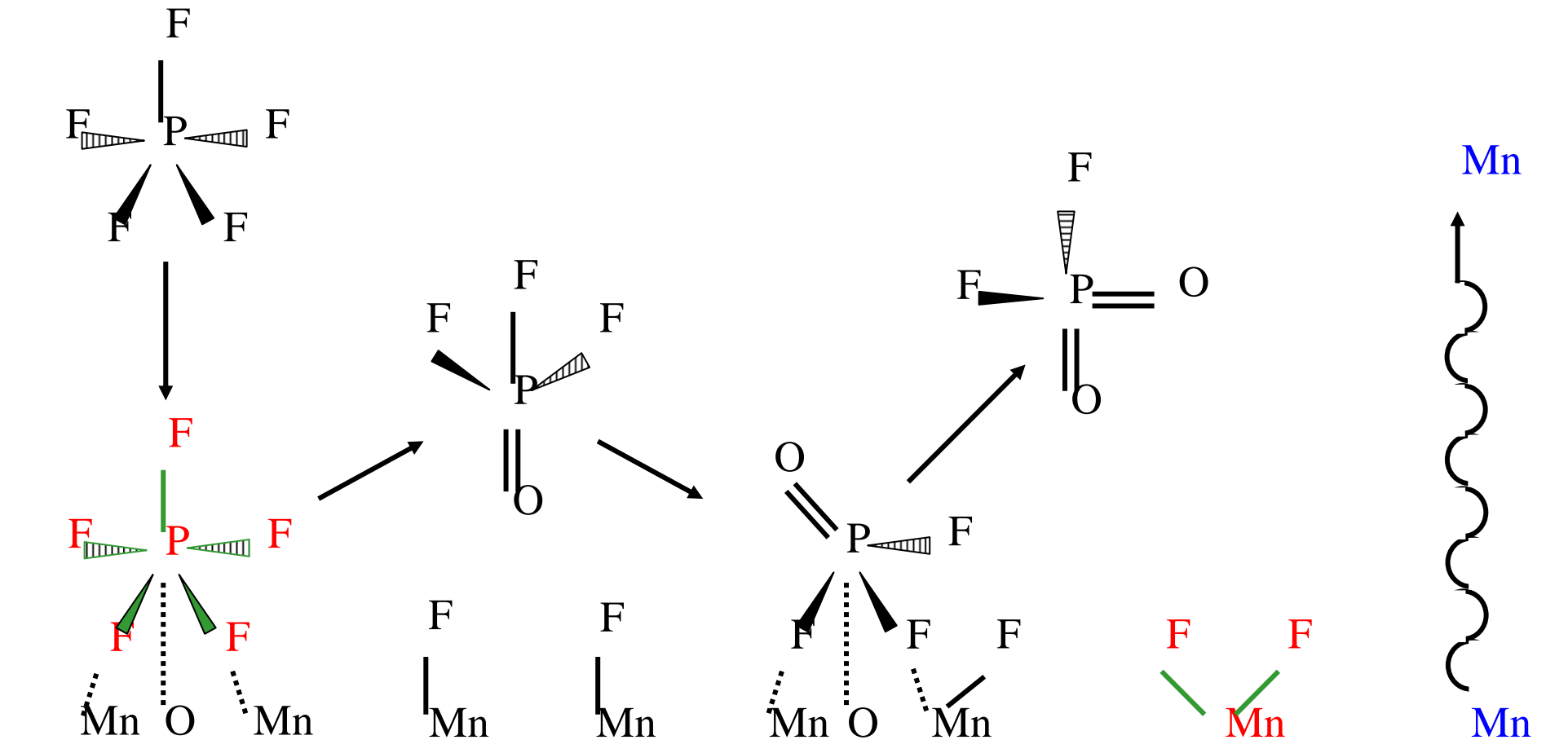
# Degradation: Capacity Measurement of $\text{LiMn}_2\text{O}_4$ After Storage at 70 C





# Proposed Surface and Solvent Degradation

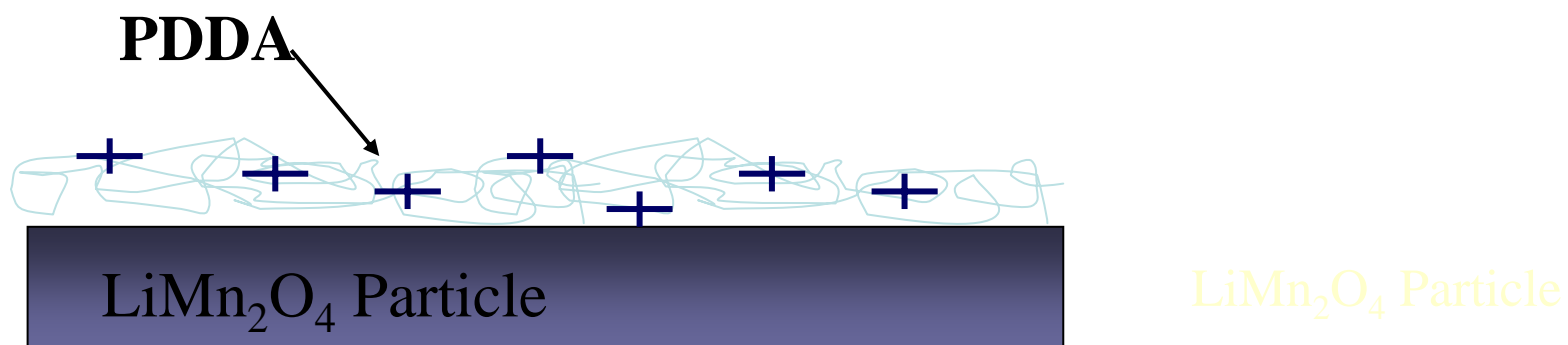
F. Quinlan et al., Chem. Mat. 2002



**Cathode Surface**

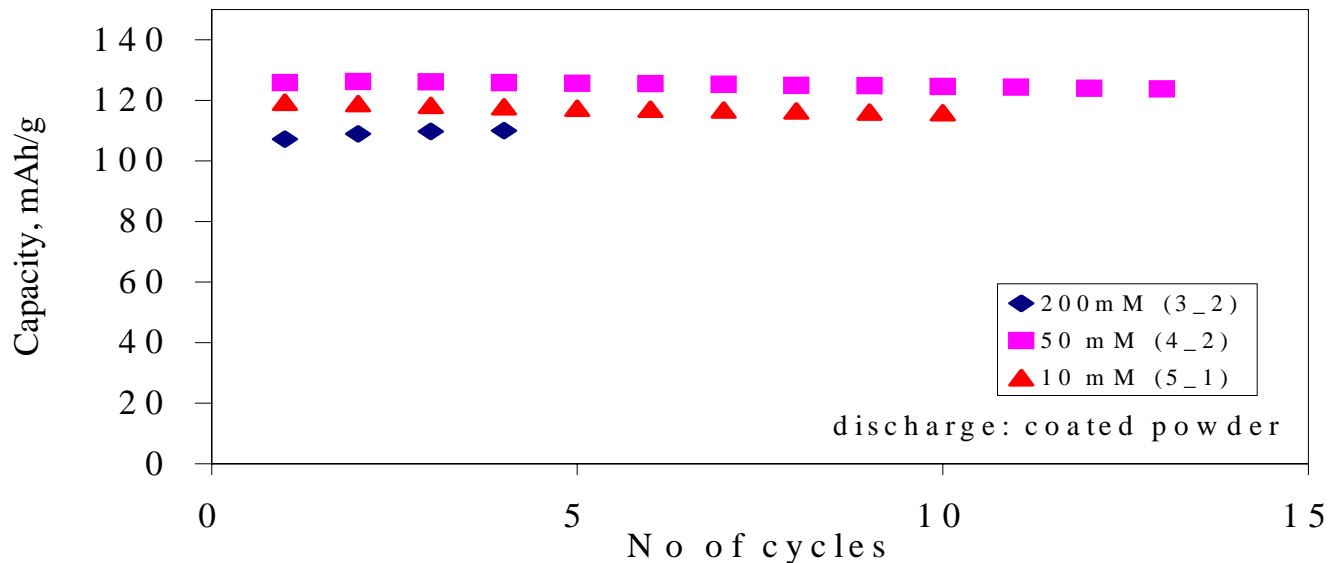
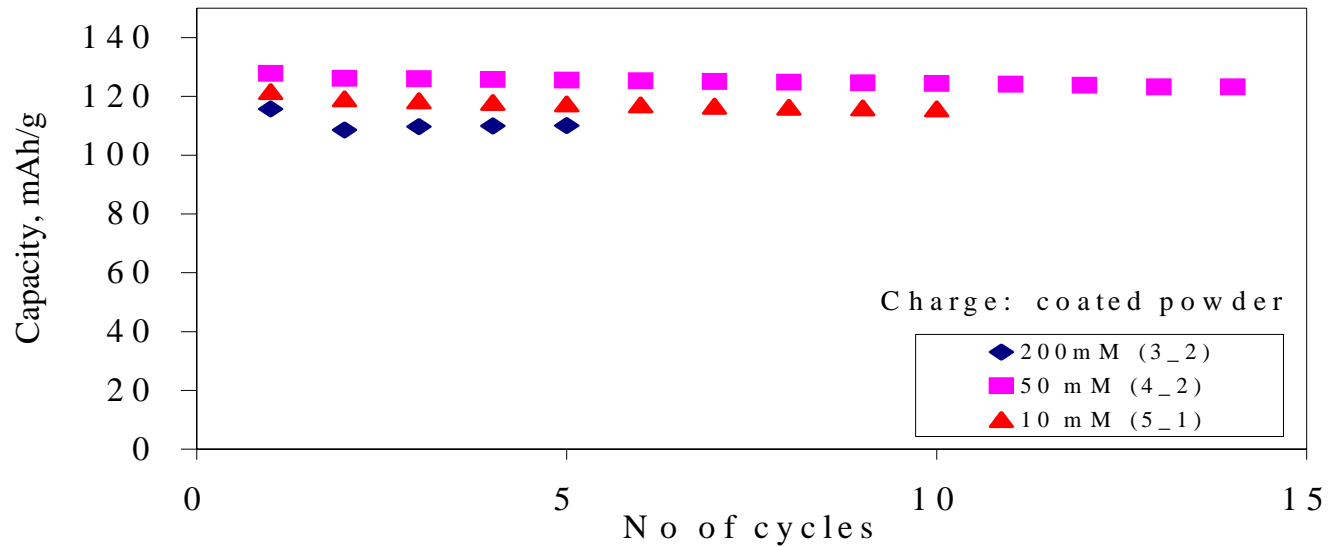


# Surface coating (adsorption) with ~1 nm PDDA on $\text{LiMn}_2\text{O}_4$ to inhibit surface degradation



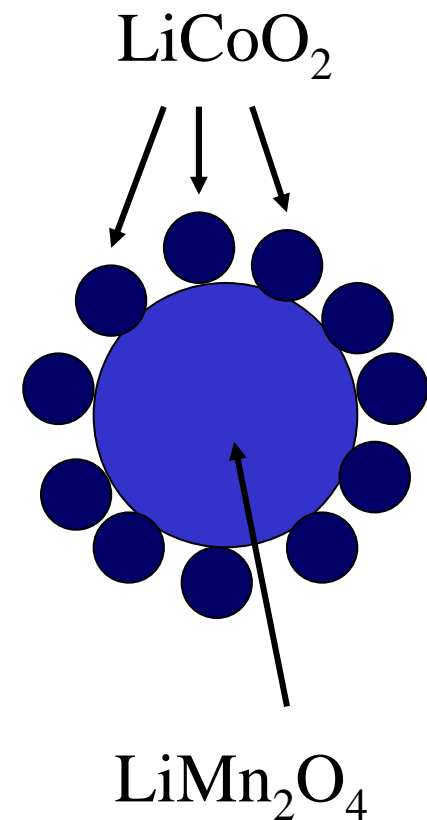
# Treated cathodes show better capacity.

The variation of the measured capacity with the number of cycles for the cathodes obtained from the PDDA coated powder (Vidu and Stroeve, in press).



# Another approach: coating $\text{LiMn}_2\text{O}_4$ powder with $\text{LiCoO}_2$ nanoparticles

- Objective – to stop surface degradation of the cathode material
- Use  $\text{LiCoO}_2$  which does not degrade at elevated temperatures
- Protect the  $\text{LiMn}_2\text{O}_4$  material with the  $\text{LiCoO}_2$  coating



# Why $\text{LiCoO}_2$ over $\text{LiMn}_2\text{O}_4$ ?

- No significant capacity fading of  $\text{LiCoO}_2$  at elevated temperatures

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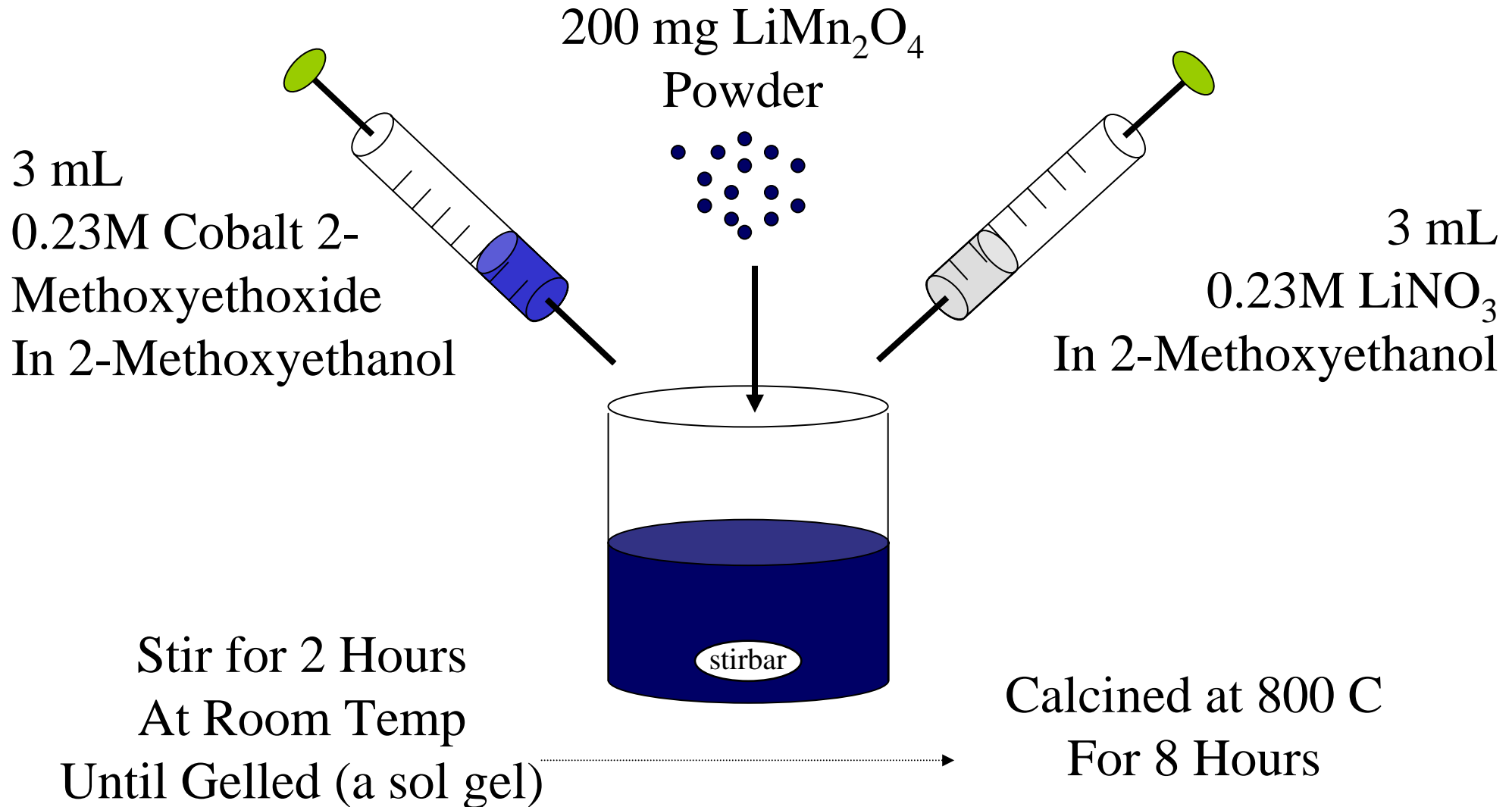
# Why $\text{LiMn}_2\text{O}_4$ over $\text{LiCoO}_2$ ?

- Less Toxic
- Less Expensive
- Easily synthesized

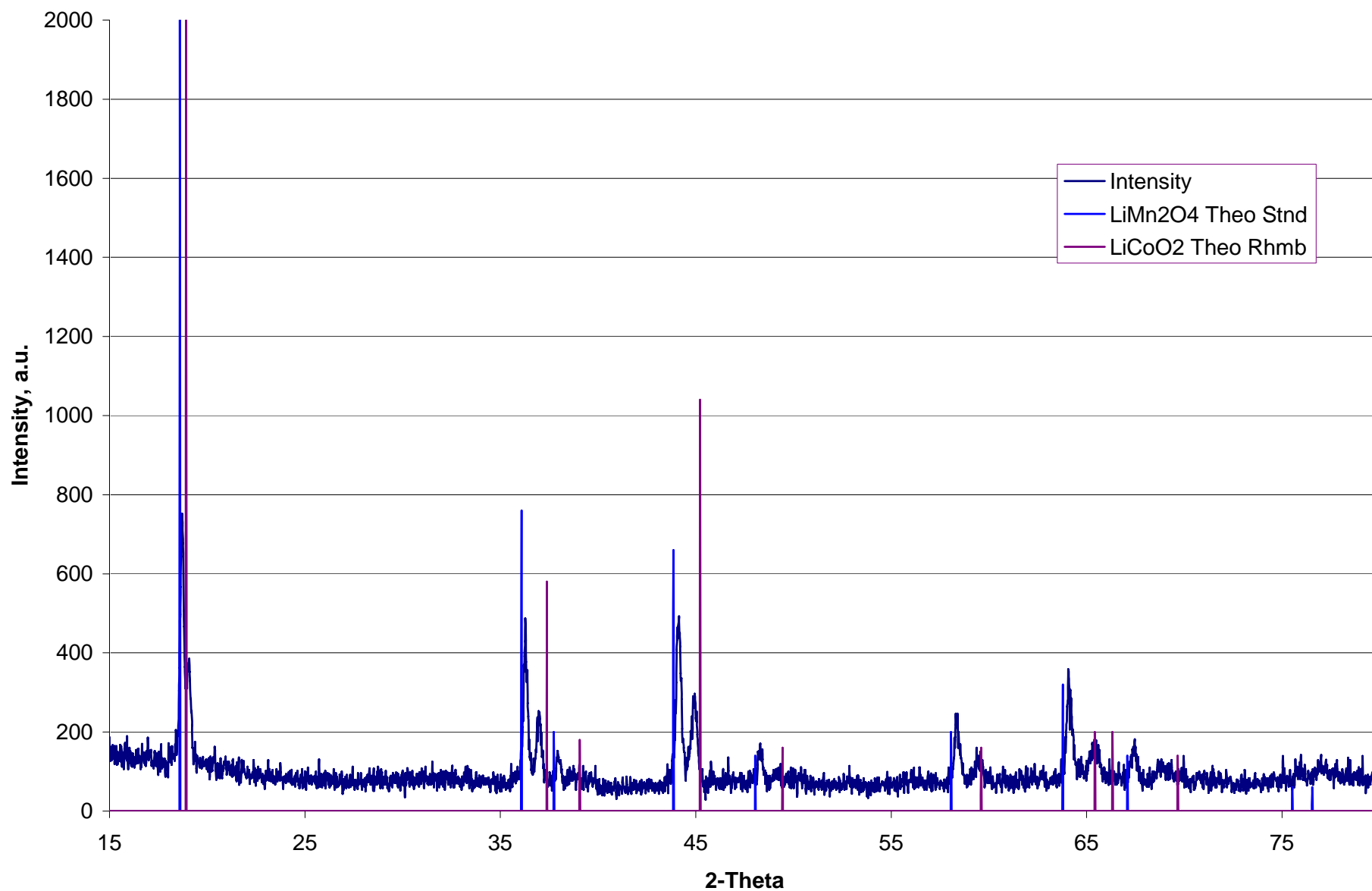
# Coating of $\text{LiMn}_2\text{O}_4$ Powder With $\text{LiCoO}_2$ nanoparticles

F.

Quinlan et al., submitted



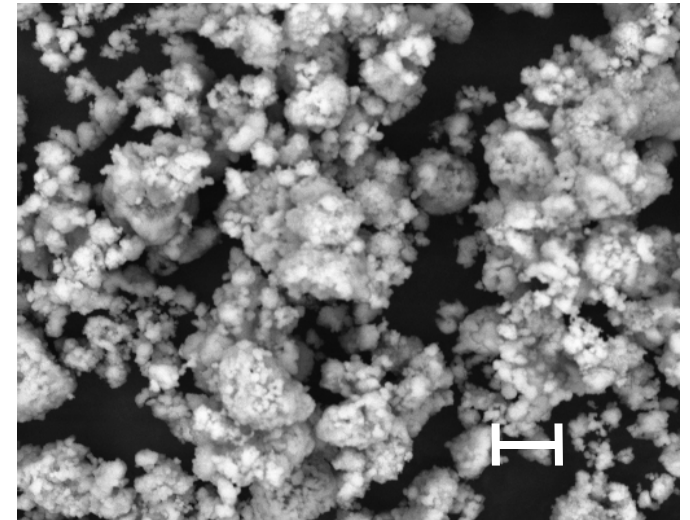
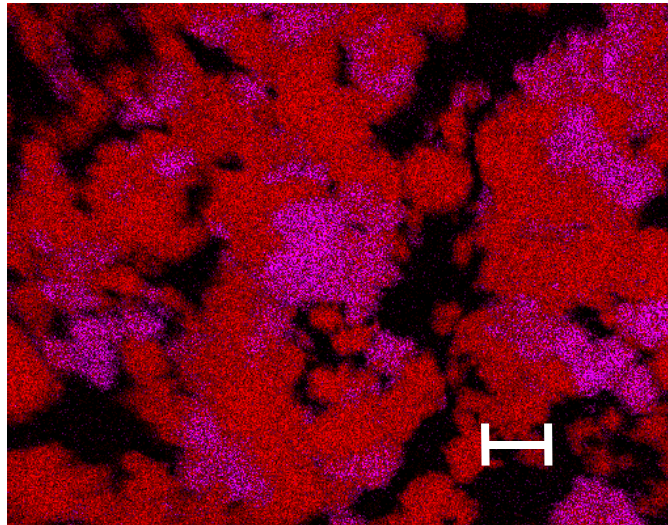
# XRD of Coated $\text{LiMn}_2\text{O}_4$ Powder With $\text{LiCoO}_2$



# SEM elemental mapping of coated Powder

Co

Mn



**SEM compositional mapping (left panel) of nanoparticles on  $\text{LiMn}_2\text{O}_4$  powder particles with Mn shown in red and Co in pink. The SEM micrograph for the identical sample location is shown in the right panel. Nanoparticles are 100 nm.**



**Surface modification on the nanoscale can lead to large changes in the macroscopic behavior of systems. Surface modification can be done in a standard laboratory**



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# Biomaterial: biological mortar as a glue for composites

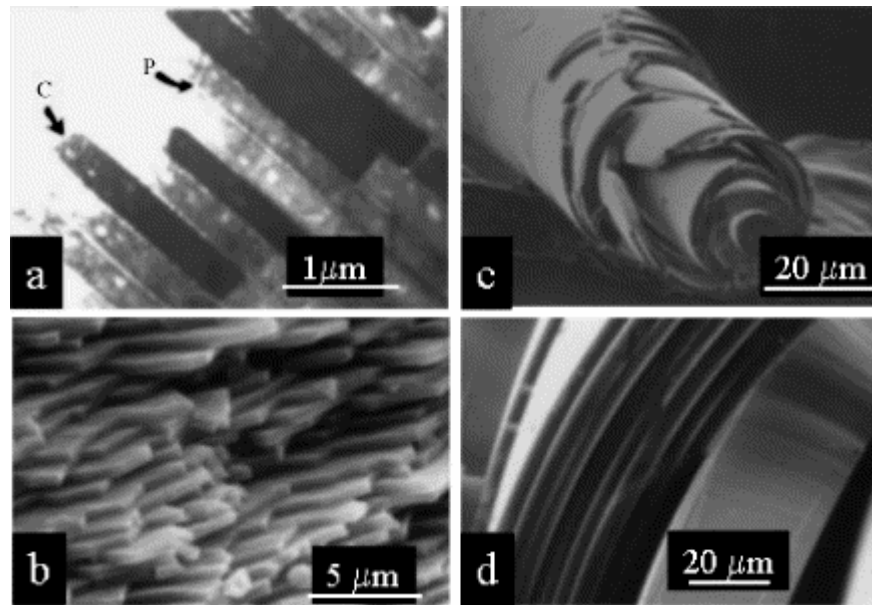


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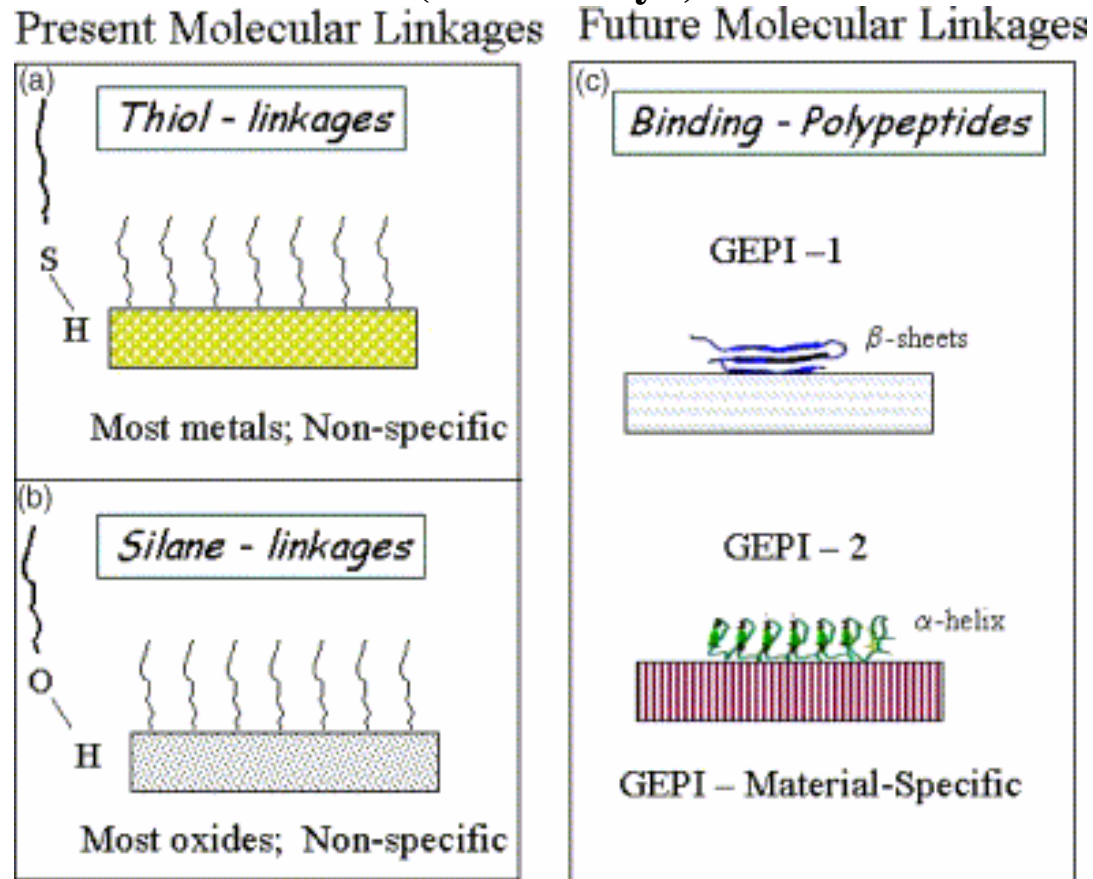
# Peptide glue

**Biological hard tissues: (a) TEM cross-section and (b) SEM fractured surface of nacre (mother-of-pearl) showing brick-mortar microarchitecture of the biocomposite, where the mortar is mostly composed of proteins. (c) and (d) SEM images of layered structure of sponge spicules in which layers are separated by an organic substance (M. Sarikaya).**



# Biomimetic molecular coatings using engineered proteins

Thiol- and silane-based self-assembled monolayers are chemically linked onto noble metals or oxides, respectively. Engineered proteins could bind to a given material specifically. While (a) thiol- and (b) silane-linkages are non-specific, (c) genetically engineered polypeptides could be material specific with a variety of conformational architectures (M. Sarikaya).

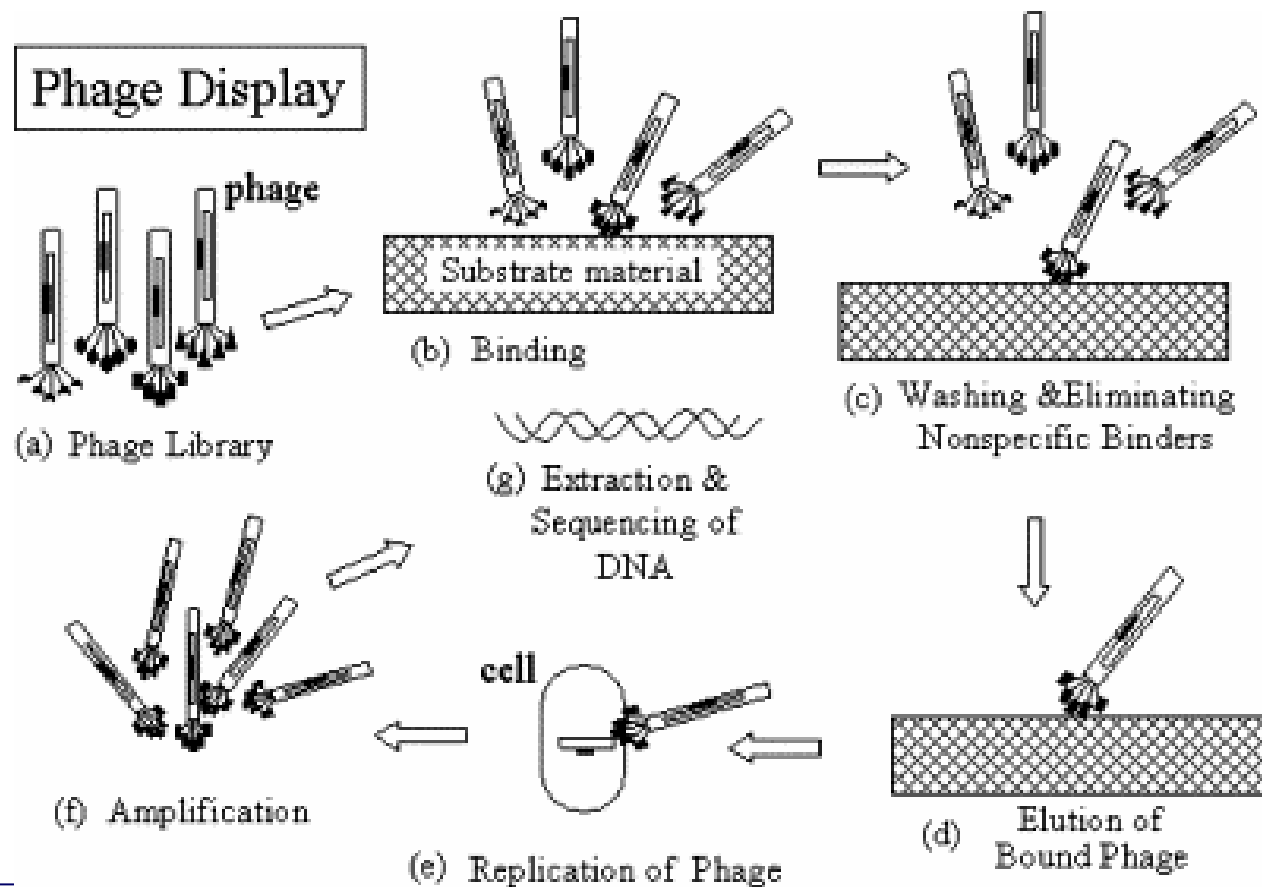


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# Biomimetic molecular coatings using engineered proteins

A flowchart of phage display protocol (from M. Sarikaya)

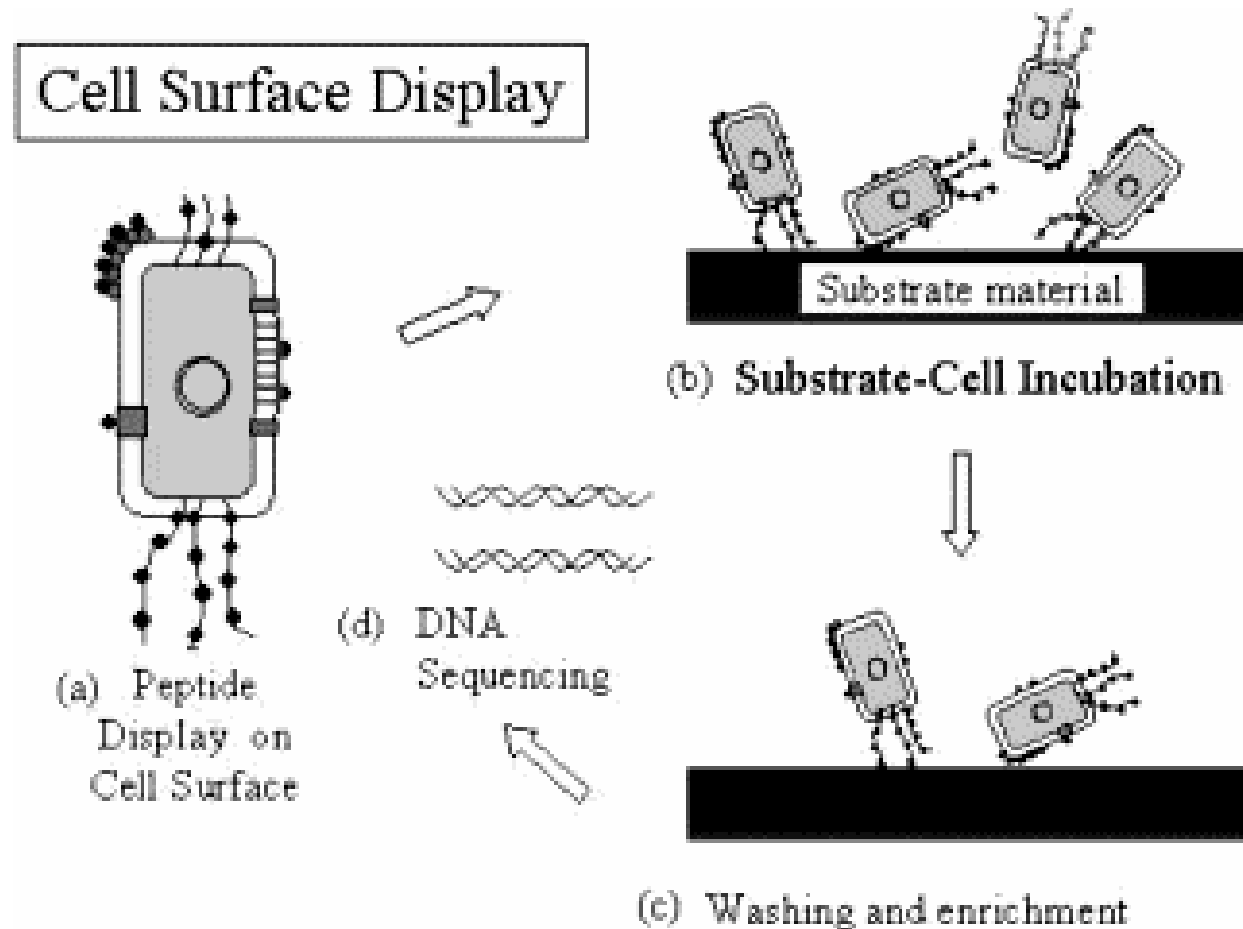


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# Biomimetic molecular coatings using engineered proteins

## Flowchart of cell surface display protocol (from M. Sarikaya)



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# Biomimetic molecular coatings using engineered proteins

Some Examples to Inorganic Binding Polypeptides	
Nobel Metals	Metal Oxides
<p><b>Au Binders:</b><sup>21,23</sup></p> <p>Au<sup>0</sup>BP1: MHGKTQATSGTIQS (14 AA)</p> <p>Au<sup>0</sup>BP2: SKTSLGQSGASLQGSEKLTNG(21AA)</p> <p>Au<sup>0</sup>BP3: QATSEKLVKGMGASLHPAKT(21 AA)</p> <p><b>Ag-Binders:</b><sup>29</sup></p> <p>Ag<sup>0</sup>BP1: AYSSGAPPMPPF (12 AA)</p> <p>Ag<sup>0</sup>BP2: NPSSLFRYLPSD (12 AA)</p> <p>Ag<sup>0</sup>BP3: SLATQPPRTPPV (12 AA)</p> <p><b>Pt-Binders:</b><sup>31</sup></p> <p>Pt<sup>0</sup>BP1: DRTSTWR (7 AA)</p> <p>Pt<sup>0</sup>BP2: TSPGQKQ (7 AA)</p> <p>Pt<sup>0</sup>BP3: IGSSLKP (7 AA)</p> <p><b>Pd-Binders:</b><sup>31</sup></p> <p>Pd<sup>0</sup>BP1: SAGRLSA (7 AA)</p> <p>Pd<sup>0</sup>BP2: TLPNHTV (7 AA)</p> <p>Pd<sup>0</sup>BP3: HTSKLGI (7 AA)</p>	<p><b>Silica-Binders:</b><sup>25</sup></p> <p>Si4-1 : MSPHPHPRHHHT (12 AA)</p> <p>Si4-10: RGRRRRLSCRL (12 AA)</p> <p>Si3-8 : KPSHHHHHTGAN (12 AA)</p> <p><b>ZnO Binders:</b><sup>30</sup></p> <p>pJKS9: <sub>RS</sub>NTRMTARQH<sub>RS</sub> ANHKSTQR<sub>ARS</sub></p> <p>pJKS15: <sub>RS</sub>YDSRSMRPH<sub>RS</sub> (9 AA)</p> <p><b>Cr<sub>2</sub>O<sub>3</sub> Binders:</b><sup>22</sup></p> <p>pKKJ62: <sub>RS</sub>VVRPKAATN<sub>RS</sub> (9 AA)</p> <p>pKKJ66: <sub>RS</sub>RIRHRLVGQ<sub>RS</sub> (9 AA)</p> <p><b>CoO Binders:</b><sup>22</sup></p> <p>pKKJ75: <sub>RS</sub>GRMQRRVAH<sub>RS</sub> (9 AA)</p> <p>pKKJ76: <sub>RS</sub>LGKDRPHF<sub>RS</sub> (9 AA)</p>
<p>Uncharged polar side chains: STNQYC</p> <p>Charged polar side chains: KRHDE</p> <p>Non-polar side chains: AVGLIMPFW</p>	<p>Hydrophobic: AGVPMILWF</p> <p>Acidic: DE</p> <p>Basic: RKH</p> <p>Hydroxyl : STY</p>



# Nanotechnology: (bio)materials

- **Rocket science?**
- **Engineered proteins for nanotechnology**
- **GeneChips**
- **Nanotubes, nanowires, nanocables**
- **Protein separation in nanopores**



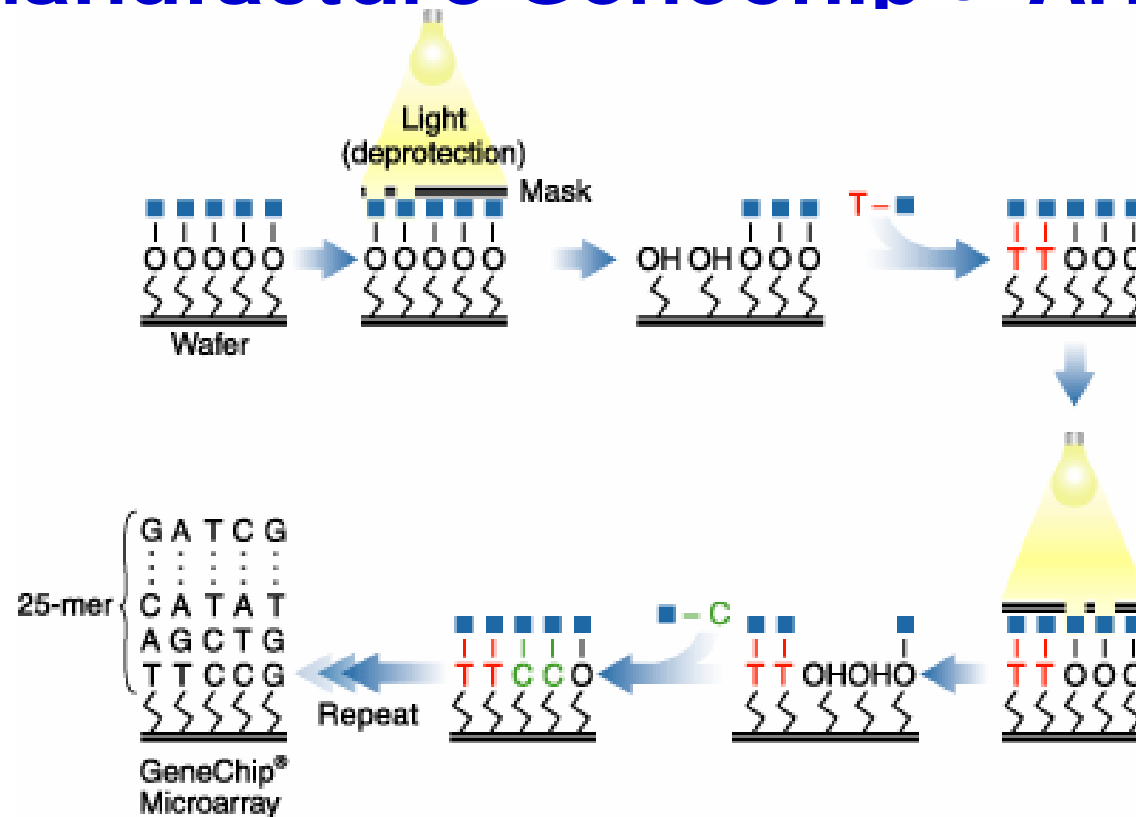
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# Nanotechnology in the life sciences

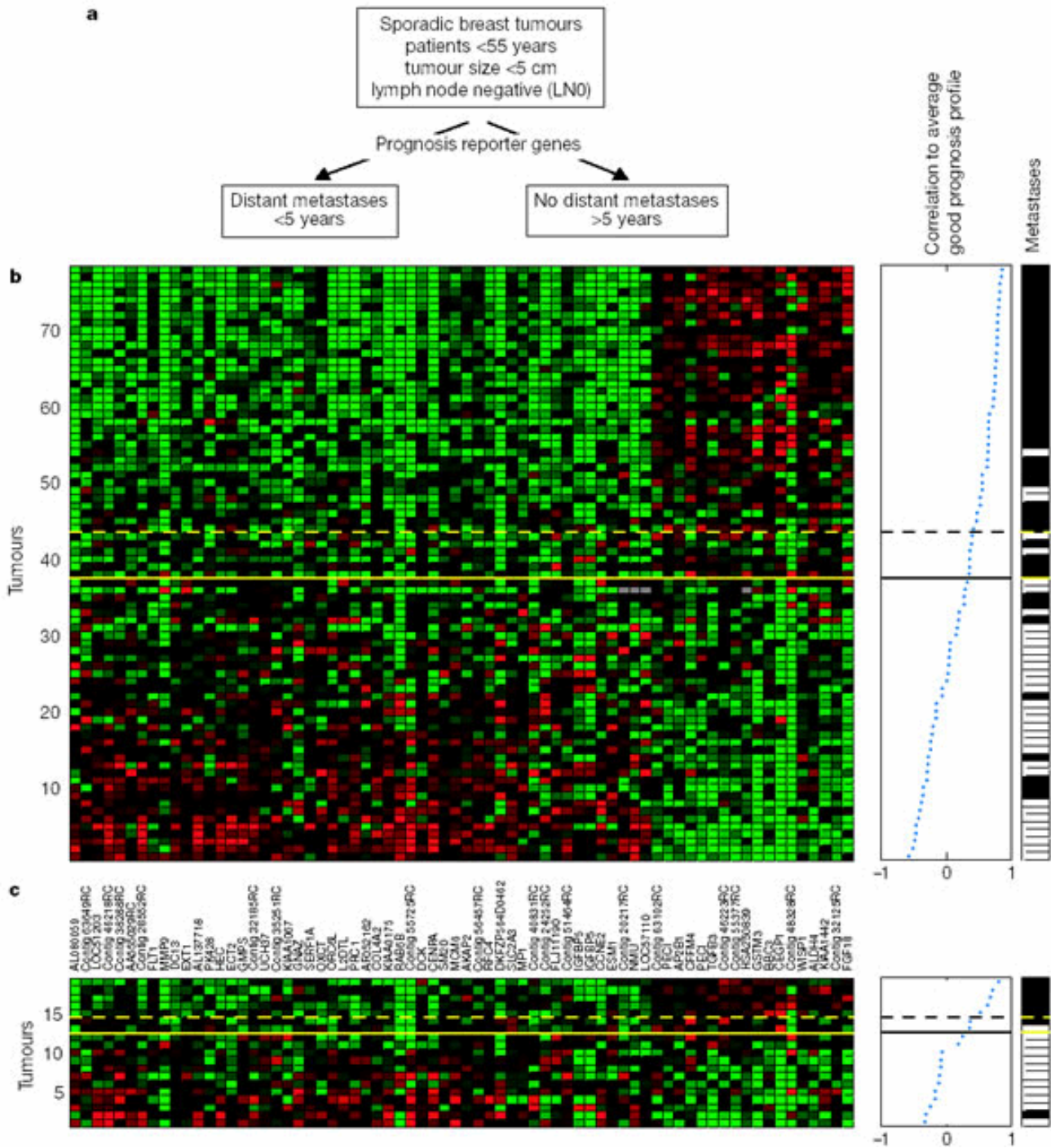
## Affymetrix GeneChip® Arrays

# Affymetrix uses a unique combination of photolithography and combinatorial chemistry to manufacture GeneChip® Arrays



# Gene expression profiling predicts clinical outcome of breast cancer. Data from 78 tumors and 70 genes.

Nature **415**, 530 - 536 (31 Jan 2002)



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# Nanowires, nanotubes, nanocables

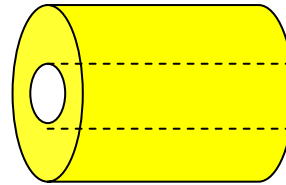
## Nanowires



Material: Cu, Fe, CdTe....

Applications: electronic devices, FED

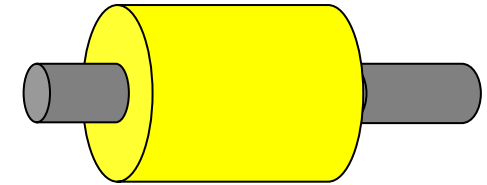
## Nanotubes



Material: C, Au, Polymer....

Applications: biosensor, FED

## Nanocables



Materials: Ag/SiO<sub>2</sub>, Zn/ZnS

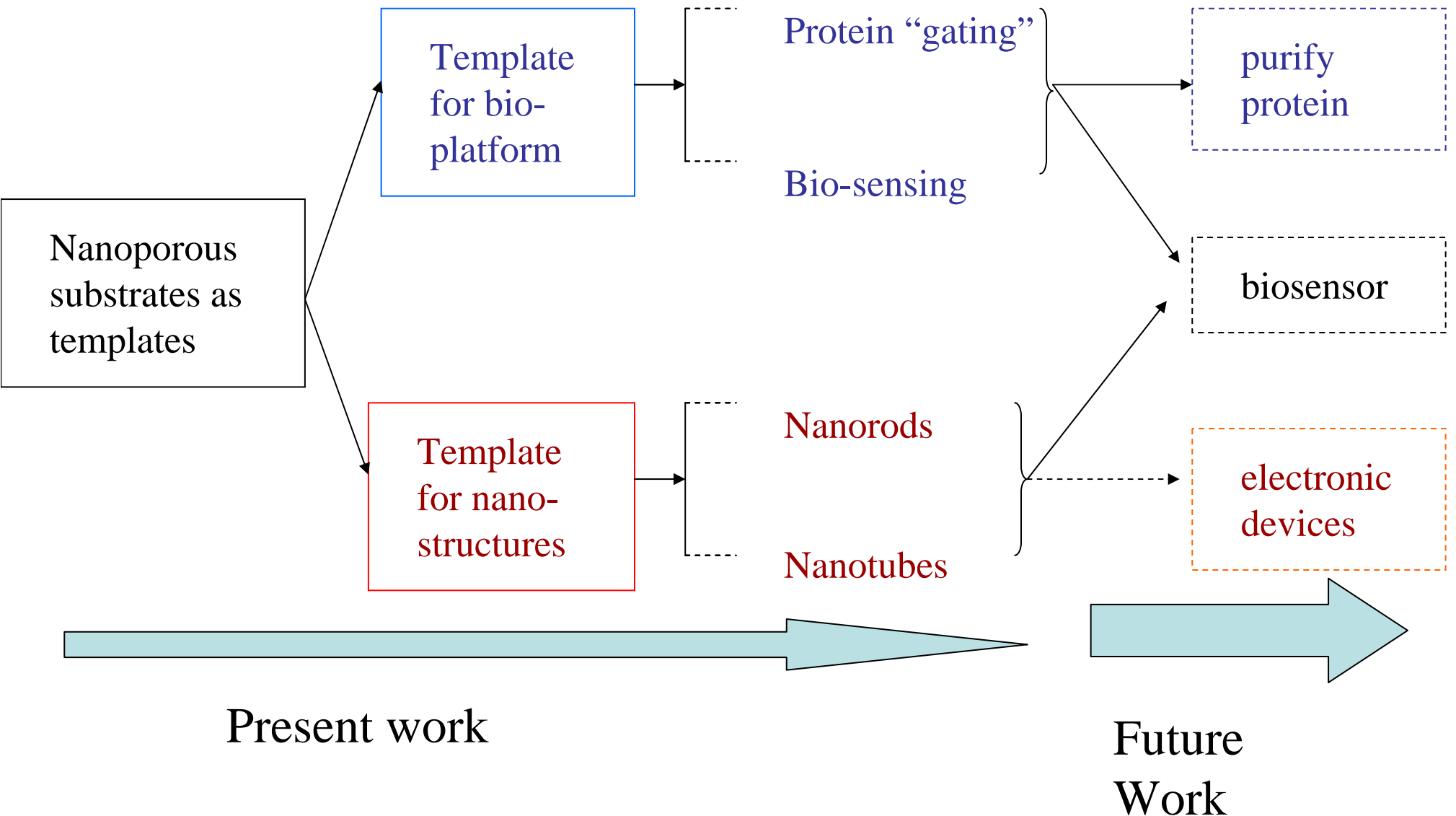
Applications: electronic devices



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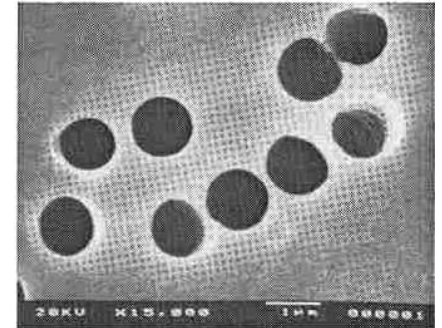
# Nanotubes



# Nanoporous Substrates

## 1. PCTE (Polycarbonate track-etching membranes)

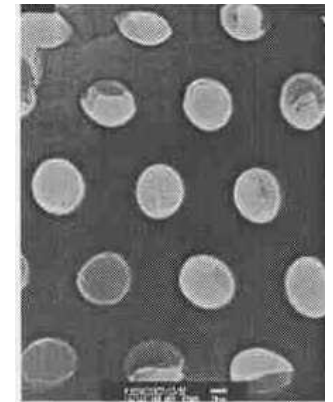
- Mono-disperse pore and narrow pore size distribution
- Random distribution
- Uniform diameters (10 nm ~ 20  $\mu\text{m}$ , commercially available)
- Pore density  $\sim 10^9$  pore/cm<sup>2</sup>



SEM image of PCTE membrane with 1  $\mu\text{m}$  diameter pores. Ref: J. C. Hulteen and C. R. Martin, *J. Mater. Chem.*, 1997 7(7), 1075.

## 2. AAO (Anodic Aluminum Oxide membranes)

- Mono-disperse pore and narrow pore size distribution
- Hexagonal array
- Uniform diameters (limited pore size, commercially available)
- Pore density  $\sim 10^{11}$  pore/cm<sup>2</sup>



TEM image of AAO membrane with 70 nm diameter pores. Ref: J. C. Hulteen and C. R. Martin, *J. Mater. Chem.*, 1997 7(7), 1075.

## 3. Porous silicon

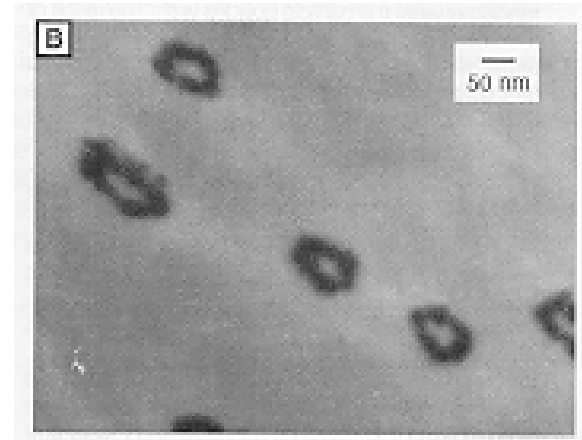
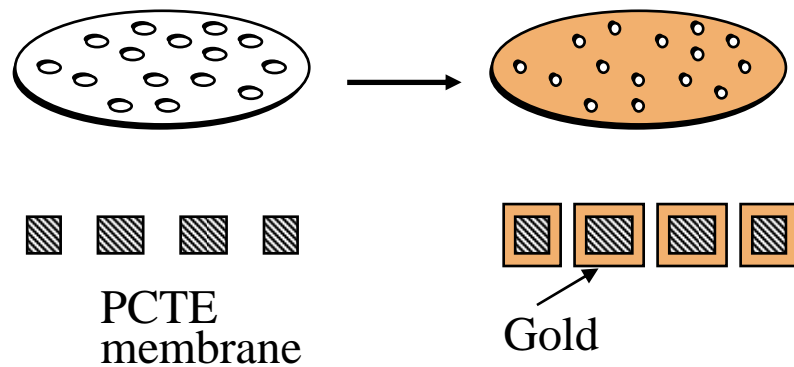
- 20 nm uniform pores



# Electroless Deposition: A Technique to Deposit Metals on Complex Surfaces

Nishizawa et al., *Science*, 1995:

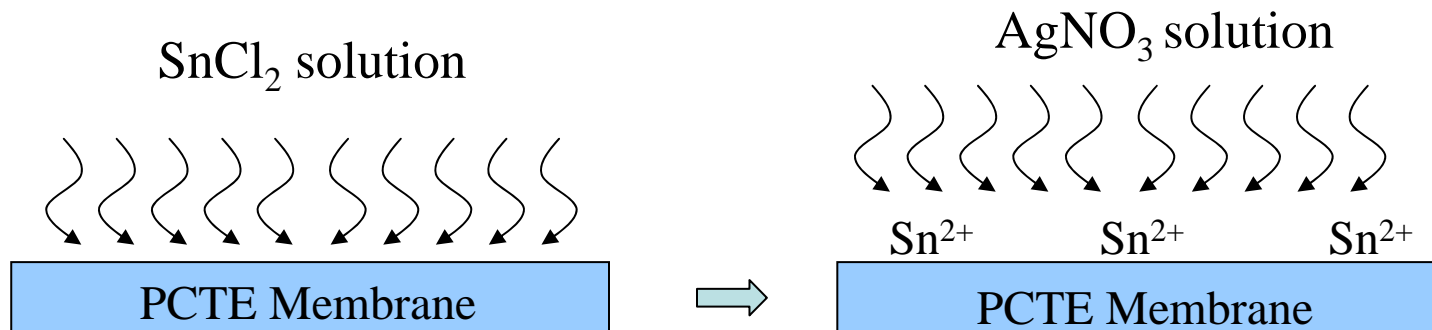
Electroless gold deposition in polycarbonate track-etched (PCTE) membranes



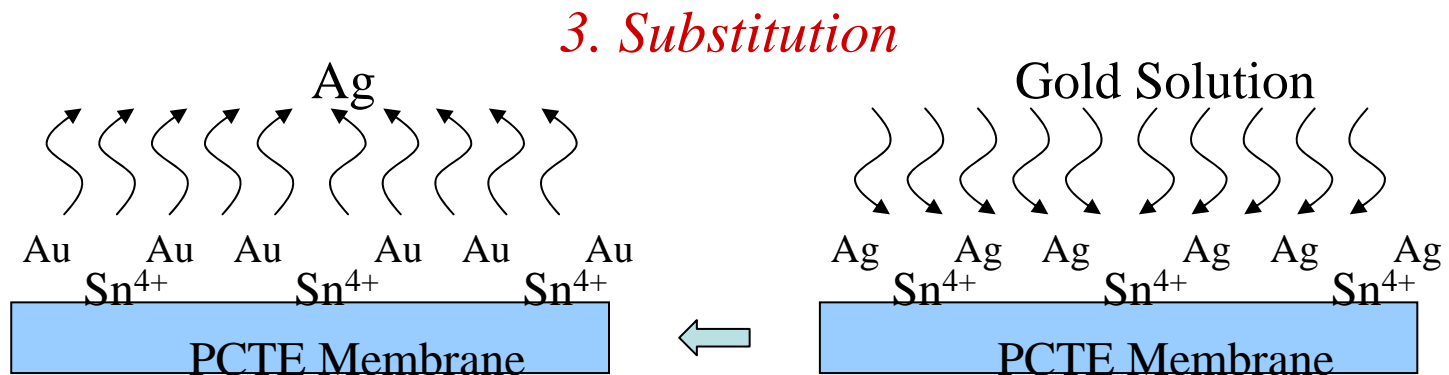
Charge-selectivity can be reversibly controlled by electrical potentials applied to gold

# Procedure of Electroless Gold Plating on PCTE Membranes

## 1. Sensitization



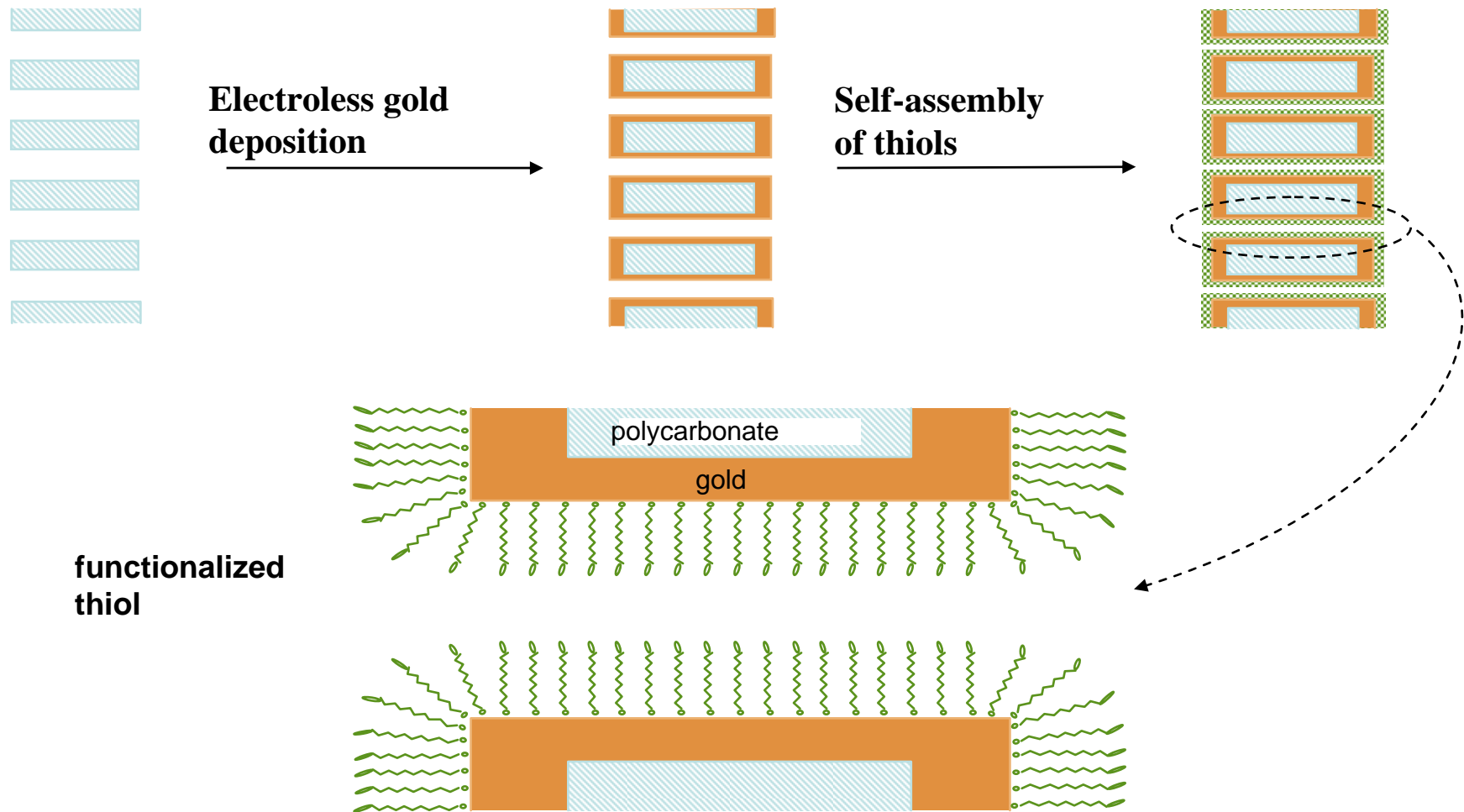
## 2. Adsorption



## 3. Substitution

Requirements : Control the pH(10~10.5) and temperature (~1°C) in the gold solution

# Modification of PCTE Membranes



**Pore radii of bare PCTE membranes: 12 - 28 nm**

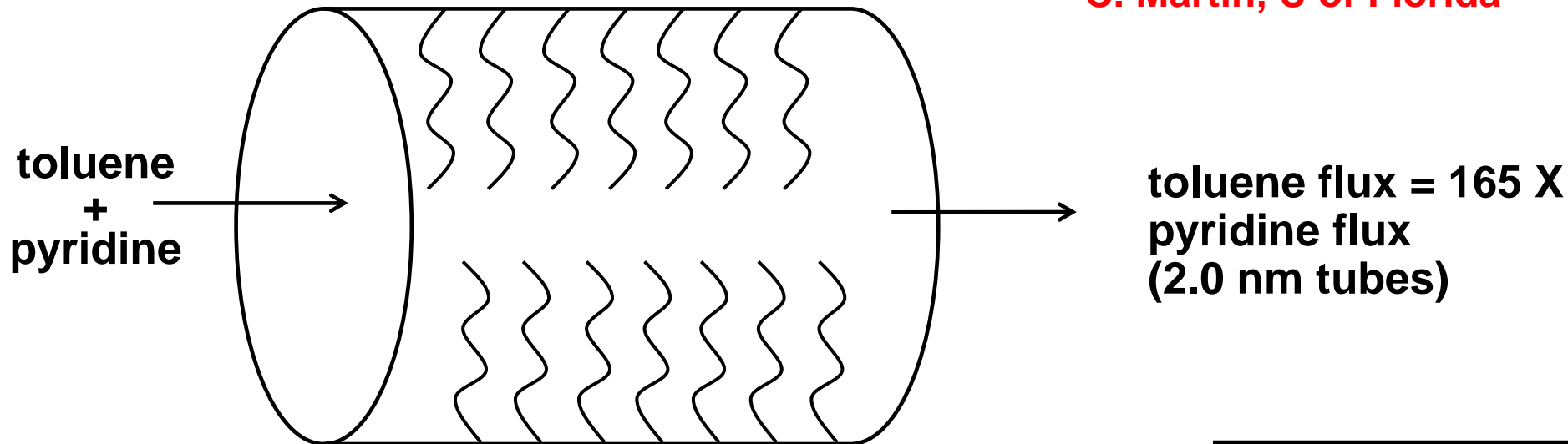
**Pore radii of after gold deposition: 4 - 15 nm**

**Hou & Stroeve, Langmuir 2000; Chun & Stroeve, Langmuir 2001, 2002; Ku & Stroeve  
Langmuir 2004**

# Functionalized nanotubes with alkane thiol chains enables separation of hydrophobic from hydrophilic species

$C_3$ ,  $C_{10}$  and  $C_{16}$  } alkane thiols 

C. Martin, U of Florida



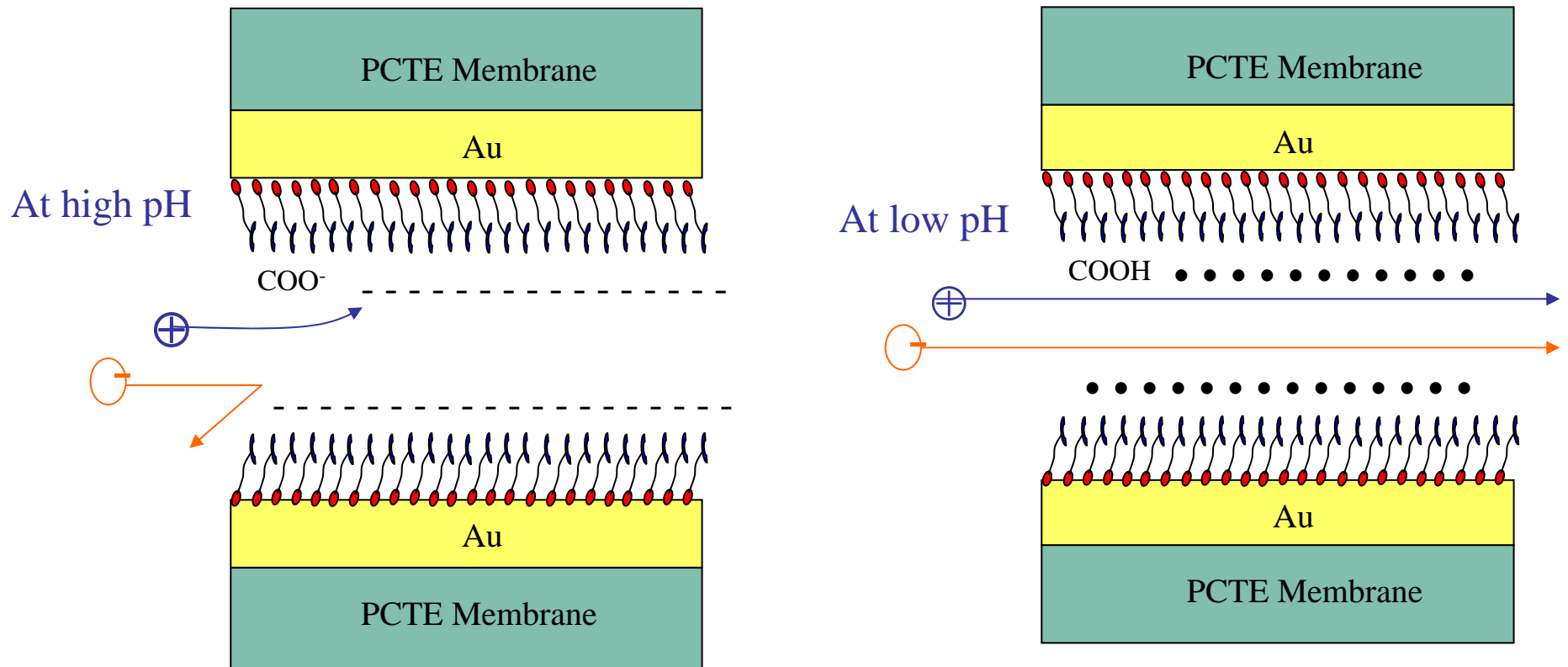
Factor of 15 X  
for 3.0 nm  
tubes

toluene: hydrophobic

pyridine: hydrophilic

# Transport Mechanism - General Idea

## pH-Responsive Nanomembranes

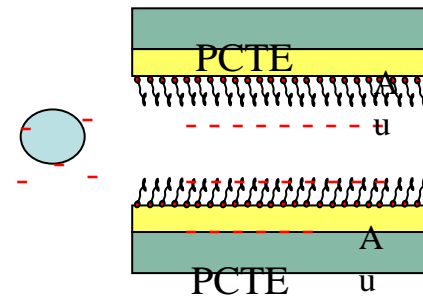


# External control factors

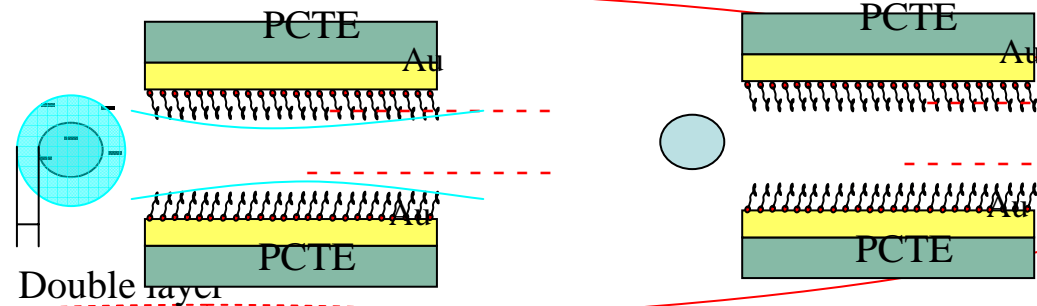
## External Control Factors

### 1. pH (Ref.:K. Y. Chun and P. Stroeve, Langmuir,

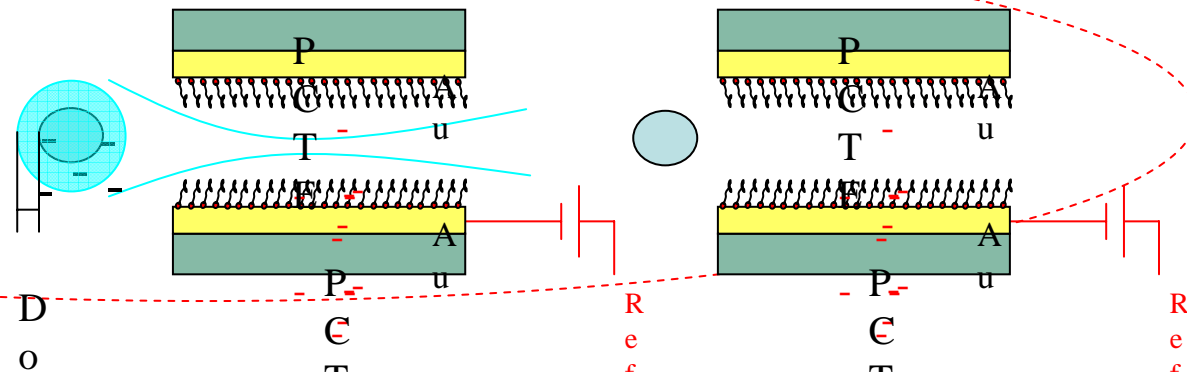
18, 4653-4658, 2002)



### 2. Ionic Strength (Debye length)



### 3. Applied Potential

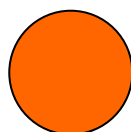


# pH responsive nanoporous membranes for protein Separation: proteins with similar molecular size and weight

Chung, Ku, Stroeve in Langmuir, 2002, 2004

**BHb** (Bovine Hemoglobin)

Spherical



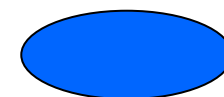
6.4 x 5.5 x 5 nm

Molecular weight = 65 kD

**pI = 7.0**

**BSA** (Bovine Serum Albumin)

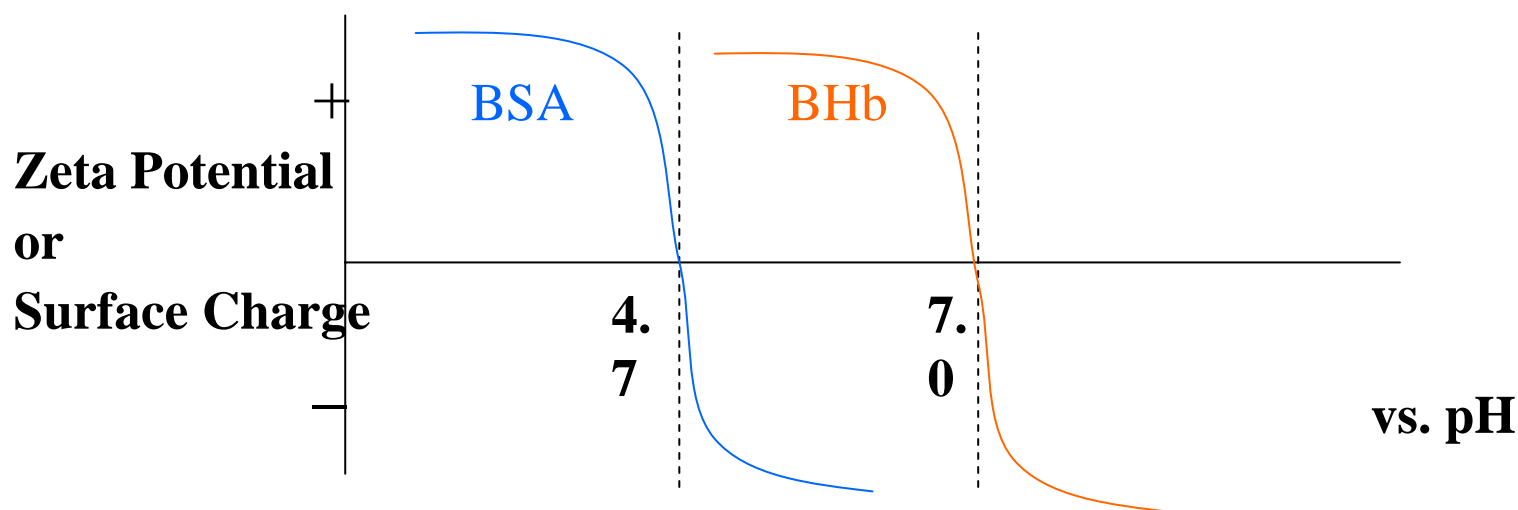
Globular ellipsoid



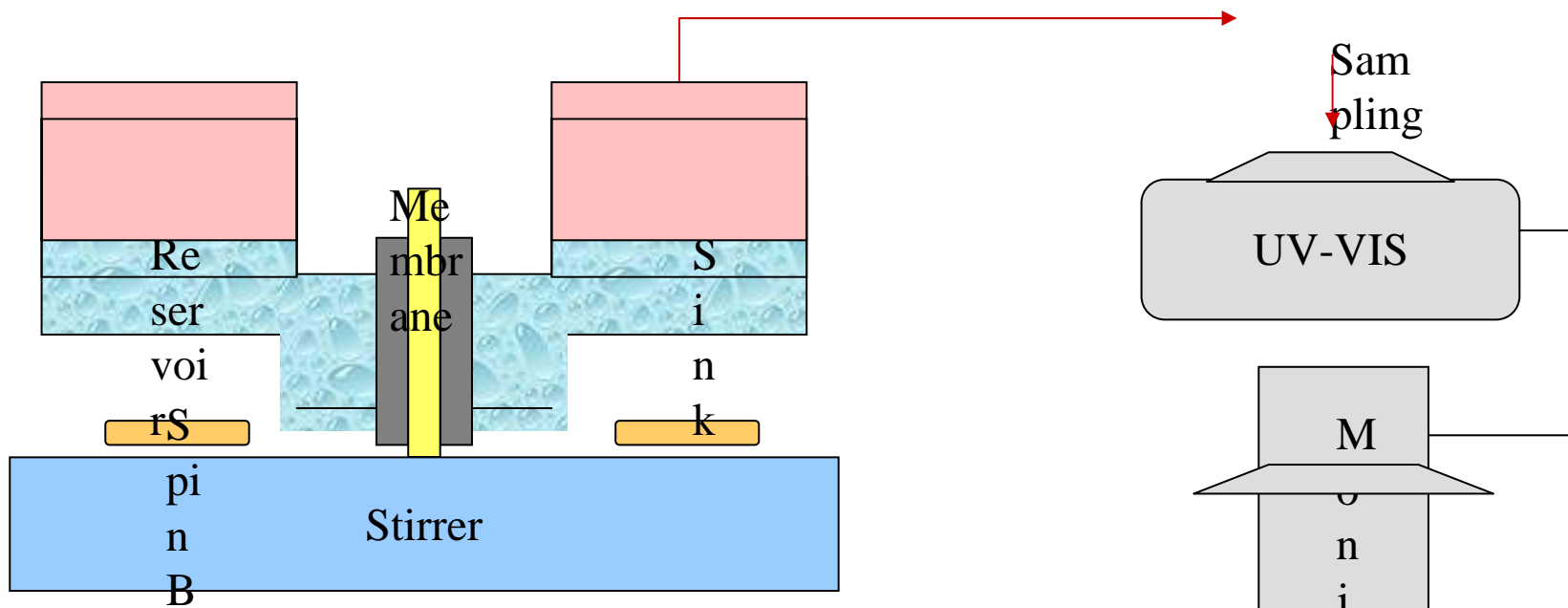
14 x 3.8 x 3.8 nm

Molecular weight = 66 kD

**pI = 4.7**



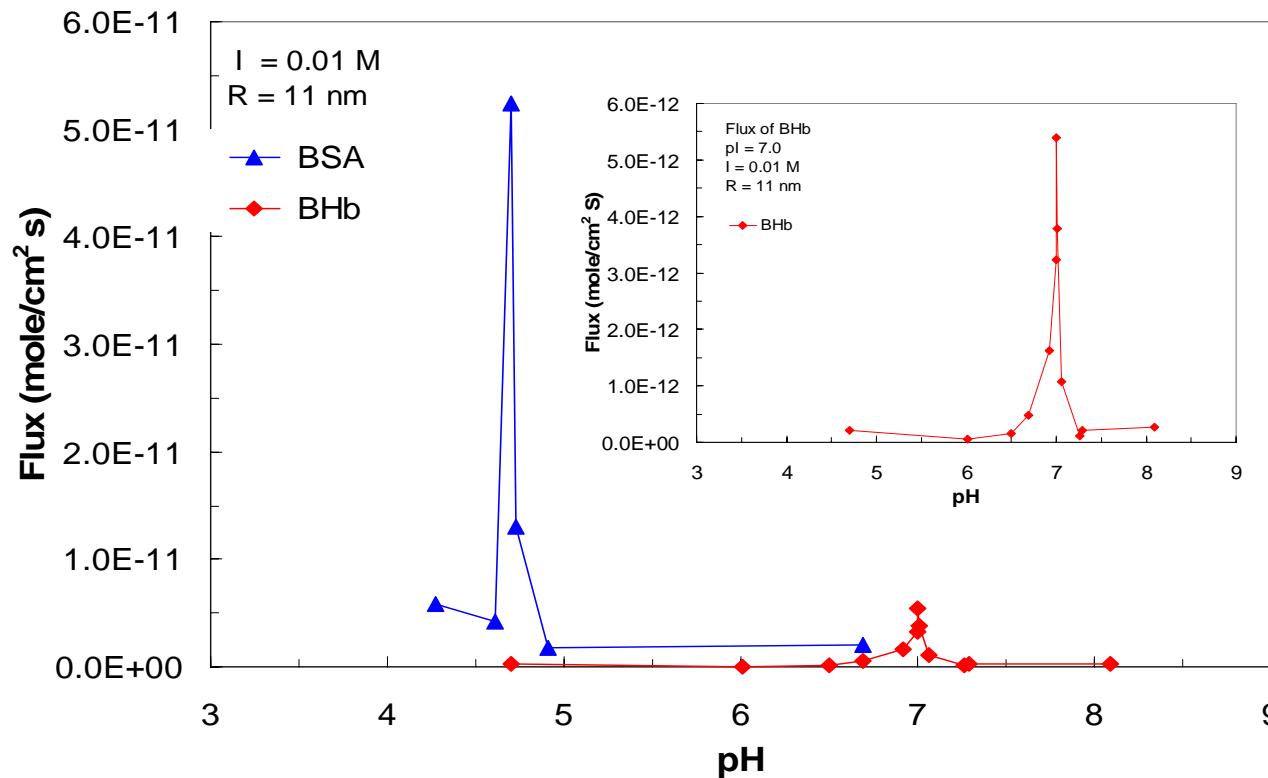
# Experiment Set-up - Diffusion Measurement



Parameters : External pH (4 ~ 8.3)  
Ionic strength (0.1M and 0.01M)  
Protein concentration in reservoir is  
(1mg/ml ~ 0.000015M)



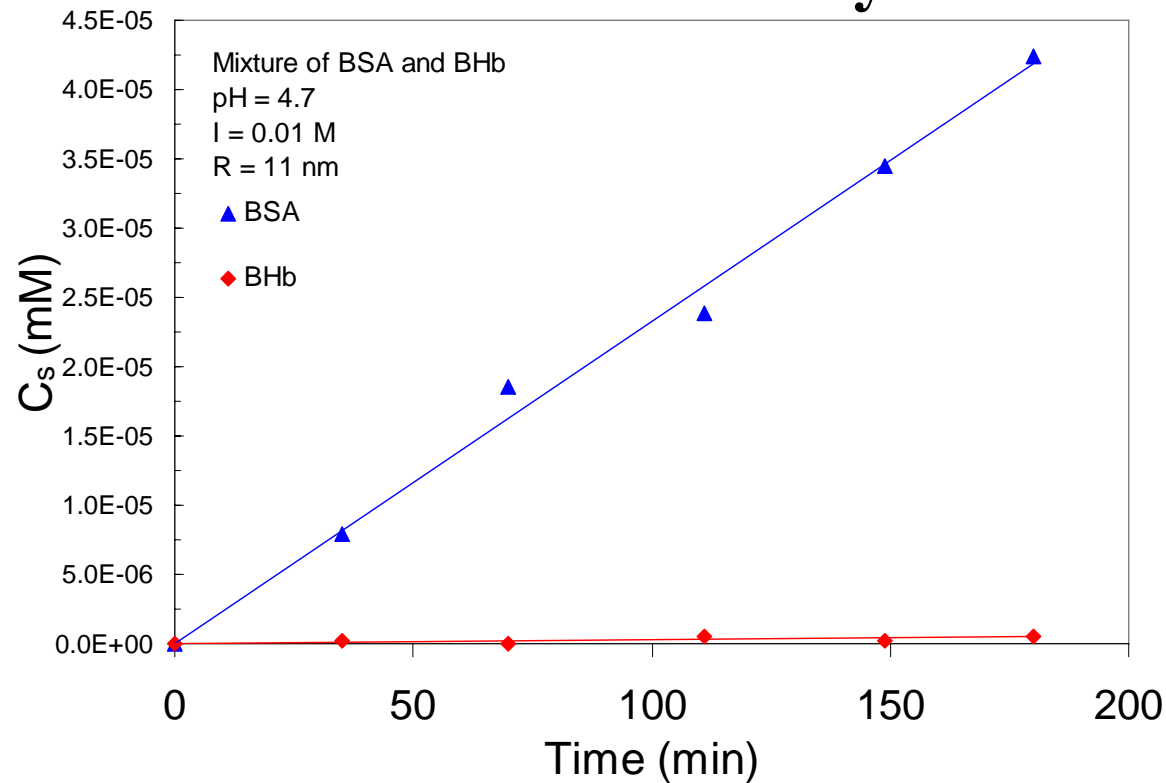
# Low ionic strength, $I = 0.01$ M, shows “molecular gating phenomena” at pI



**Comparison of fluxes versus pH for single BSA and single BHb transport experiments for  $I = 0.01$  M. The insert shows the enlarged BHb results. Ku and Stroeve, Langmuir, 2004**

# Mixture of BHb+BSA at low ionic strength and pI of BSA

The BSA/BHb selectivity > **67**.



Sink concentrations versus time for a mixture of BSA and BHb at pH = 4.7, I = 0.01 M, R = 11 nm. Ku and Stroeve, Langmuir 2004.

# Protein transport in nanoporous membranes

- **At low ionic strength, protein transport occurs mainly at the pI of the protein leading to a pseudo “molecular gating effect”.**
- **The selectivity of BSA over BHb is improved at the pI of BSA when the buffer solution is at a low ionic strength.**

# Nanotechnology in the life sciences: (bio)materials

## Closing comments:

**Nanotechnology has produced new materials with interesting properties. Nano-materials offer unique applications in biology. Surfaces play a dominant role in nanosystems.**