Distribution and host range of the recently introduced willow sawfly, *Nematus oligospilus* Förster, on willows (*Salix* spp.) in southeast Australia.

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Summary A survey of pathogens and invertebrate fauna of willows (*Salix* L. spp., Salicaceae) was undertaken in southeast Australia. Rapid sampling techniques were employed to gather presence /absence data on the organisms inhabiting willows.

The willow sawfly, Nematus oligospilus Förster, is far more widespread than was previously thought and new occurrences were recorded for the Australian Capital Territory, New South Wales, Victoria and South Australia, as well as being detected in Tasmania for the first time. Willow species predominantly affected by sawfly included S. fragilis L. (crack willow), S. babylonica L., S. x sepucralis Simonk. var. sepucralis Simonk. and S. x sepucralis Simonk, (Dode) Meikle var. chrysocoma (Dode) Meikle (weeping willows) and S. alba L. var. vitellina (L.) Stokes (golden willow). It was also been found on (S. matsudana Koidz. 'Tortuosa' (tortured willow), S. humboldtiana Willd. 'Pyramidalis' (pencil willow) and S. cinerea L. (pussy willow). Occurrences on the S. cinerea, are the first records of sawfly on these species in the Southern Hemisphere.

Keywords. willows, Salix, sawfly, Nematus oligospilus.

INTRODUCTION

Willows occupy large areas of temperate Australia with extensive invasions in the southeast. They have deleterious impacts on water, agriculture, recreational, amenity and biodiversity values, but are considered by some to positively influence landscape values. Of the 32 willow taxa naturalised in Australia all, with the exception of *S. babylonica, S. x reichardtii* A. Kern and *S. x calodendron* Wimm., are designated Weeds of National Significance (ARMCANZ 2001). Despite this, there is no detailed knowledge of the indigenous and introduced organisms utilising willow taxa in Australia, or the effect that these organisms are having on willow growth and reproduction.

The willow sawfly, first detected in Canberra in 2004, was possibly introduced from New Zealand where it has been established since 1997 (Bruzzese and McFadyen 2006). The current distribution of

the sawfly encompasses disparate locations in Canberra, southern New South Wales (NSW), the Adelaide Hills and suburban Melbourne (Anon. 2005a, Bruzzese and McFadyen 2006). A major objective of this survey was to determine the current geographical and willow host range of *N. oligospilus* in Australia as it is reported to have a destructive effect on willows elsewhere in the southern hemisphere (Koch and Smith 2000)

MATERIALS AND METHODS

Following a survey of willow pathogens and invertebrate fauna undertaken in the summer of 2005-2006 in southeast Australia, the presence and absence of willow sawfly was recorded. The survey focussed on Victoria due to its extensive willow invasions (ARMCANZ 2001), but also included NSW, the Australian Capital Territory (ACT), South Australia and Tasmania. Tree willows (*Salix* subgenus *Salix*) and shrubs willows (*Salix* subgenus *Vetrix*) were included in the survey as encountered.

Site selection. A site consisted of one or more trees and was selected to cover as much as possible of the current distribution of willows in southeast Australia. Time constraints dictated that sites had to be spaced widely so there was little deviation from major road routes. At each site a detailed description was recorded along with GPS and altitude readings taken.

Tree sampling. Trees at each site were chosen randomly for sampling according to the following criteria: a basal diameter of at least 10 cm, the lowest branches were within reach of the long-handled tree loppers (about 5m) and within a cluster of willows; no two trees with overlapping canopies were sampled. Early in the survey two trees of each taxon were sampled per site, but this proved excessively time-consuming and later only one tree per taxon was sampled. Any obviously unhealthy or herbivore-damaged tree at a site was also sampled.

Rapid standardized sampling techniques were employed to collect presence/absence data only. Quantitative sampling to determine abundance was beyond the scope of this project. A branch sample of each tree was taken and pressed for identification. Each tree was tagged with an identification number, and details such as tree height, distance from and type of watercourse, tree habit, bark colour, fragility, leaf glaucousness and apparent herbivory were recorded.

Sampling involved three methods: visual inspection of whole trees, beating branches using a beating tray and laboratory examination of cut branches. Each tree was visually inspected from the ground for a total of ten minutes, with the entire circumference of the tree inspected if possible. Invertebrates found during this search were collected with a pooter and foliage with sessile insects and / or diseased leaves or twigs was stored in plastic bags for later identification. Two randomly selected branches, on opposite sides of the tree, were beaten with a rod for 30 seconds each and a beating tray (84cm square) was used to collect fallen invertebrates. A further two branches were cut down, again on opposite sides of the tree, using long handled tree loppers to cut about 5m above the ground. Branches between 0.5m and 1m were immediately placed in plastic bags for subsequent microscope examination in the laboratory.

All invertebrate material collected was preserved in 70% ethanol except any material likely to be damaged by this method of preservation such as moths, which were placed in a jar containing ethyl acetate. Plant material was kept refrigerated at between 10-15°C until examined. All samples were inspected for the presence of willow sawfly.

RESULTS

A total of 214 sites were visited and a total of 336 trees sampled (see Fig. 1). Willow taxa sampled included the tree willows (Salix subgenus Salix) such as S. alba L. var. caerulea (Sm.) Sm. (cricket bat willow), S. alba var. vitellina, S. alba x matsudana, S. babylonica, S x pendulina, S. x sepulcralis var. sepucralis, S. x sepulcralis var. chrysocoma, S. fragilis, S. matsudana 'Tortuosa', S. humboldtiana, S. x rubens Schrank (basket willow), S. nigra Marshall (black willow) and a number of unusual hybrid species such as S. alba var. vitellina x S. matsudana 'Tortuosa'. (golden tortured willow). Also sampled were the shrub willows (Salix subgenus Vetrix) including S. cinerea L. and S. x reichardtii (pussy willows), S. viminalis L. (osier) and S. purpurea L. (purple osier). Nomenclature follows Cremer (1995) with updates from G. Carr (pers. comm.).

The predominant tree species affected by *N. oligospilus* was identified as *S. fragilis*. However, willows readily hybridize causing some difficulty in identification. In the field, *S. fragilis* and *S. x rubens* (hybrid between *S. alba* and *S. fragilis*) can be easily confused (Kennedy *et al.* 2003). Closer examination of specimens may reveal that trees referred to as *S. fragilis* may have been *S. x rubens*.

Other prominent taxa of willows affected were the S. alba var. vitellina, S. babylonica, S. sepulcralis var. sepulcralis, S. sepulcralis var. chrysocoma. There were also some occurrences of N. oligospilus on S. alba x matusdana, S. humboldtiana and S. cinerea. There were no occurrences on S. viminalis, S. nigra or S. purpurea. In the ACT, N. oligospilus occurred predominantly on S. fragilis, but also on S. babylonica, S. sepucralis var. sepucralis, S. x sepucralis var. chrysocoma, S. alba var. vitellina, S. matsudana 'Tortuosa' and S. x rubens. The sawfly was not found on any shrub willows sampled. Most of the sampling sites were in Canberra, where the insect was first recorded in Australia (Bruzzese and McFadyen 2006), although new occurrences were found in the Brindabella.

In NSW occurrences mirrored that of the ACT *N. oligospilus*. Existing distribution records of *N. oligospilus* in NSW were not specific and referred only to surrounding areas of Canberra such as Queanbeyan, Braidwood and Cooma, southeast and southwest NSW (Anon 2005a,b), although the latter is probably a typographical error. New distribution records found in this study include Gundagai, Tumut, Mullengandra, Deniliquin and the Snowy Mountains region (Adaminaby, Jindabyne, and Tumbarumba).

In South Australia, *N. oligospilus* was found primarily on the weepers; *S. babylonica, S.* x *sepucralis* var. *sepucralis* and *S. x sepucralis* var. *chrysocoma*) and to a lesser extent on *S. fragilis* and *S. alba* var. *vitellina*. The Adelaide Hills were the only previously reported location for *N. oligospilus* (Anon 2005b), but this study has found populations from Victor Harbour, sites east to Murray Bridge and north to Tanunda in the Barossa Valley.

In Tasmania willow species attacked included S. fragilis, S. fragilis crosses with S. babylonica or S. x sepucralis var. sepucralis and a golden New Zealand hybrid species (S. alba x matsudana x. alba var. vitellina). All occurrences constitute new distribution records.

In Victoria there were relatively few occurrences for the number of sites sampled.

However, N. oligospilus was found on S. fragilis, S. babylonica, S. sepucralis var. sepucralis, S. x sepucralis var. chrysocoma, S. alba. var. vitellina and an usual cross between S. x rubens and S. matsudana 'Tortuosa'. There was also one occurrence each S. humboldtiana and S. cinerea. The only previously reported occurrence of N. oligospilus in Victoria was in suburban Melbourne (Bruzzese and McFayden 2006).

Table 1 shows the number of sites per state and the areas in each state/territory where sawfly were found. Figure 1 represents survey samples sites with occurrences of sawfly marked.

Of the other tenthredinid species associated with willows in Australia the gall-inducing *Pontania* proxima (Lepeletier) was found predominantly on *S. fragilis* in Tasmania, where it is currently endemic (Naumann et al 2002). *Amauronematus viduatus* (Zetterstedt), currently known only from southeast NSW (Naumann et al. 2002), was found on weeping willows and *S. fragilis* in this area and scattered locations throughout Victoria.

DISCUSSION

The host preference of sawfly for tree willow species in Australia concurs with New Zealand research that showed it has a marked oviposition preference for tree over shrub willows (Charles *et al.* 1998).

The low incidence on *S. humboldtiana* in this survey is not representative of the South American situation where it has been reported numerous times (Koch and Smith 2000).

There are records of *S. nigra* as hosts for *N*.

oligospilus in New Zealand (Charles et al. 1998) and South America (Koch and Smith 2000) and

only very recently (April 2006) there have been reports of extensive infestations of sawfly on *S. nigra* from the Ovens River near Wangaratta, Vic (A. Briggs pers. *comm.*). The apparent absence of sawfly on *S. nigra* in this survey is probably a result of the relatively few trees of this species sampled. Our study has recorded for the first time sawfly feeding on pussy willow in Australia, although there is anecdotal evidence of feeding on this species in New Zealand (Harman 2004). It must be borne in mind, however, that host range may differ from country to country given that sawfly may be considered a complex of closely related species (Koch and Smith 2000).

The level of defoliation from sawfly was particularly high on *S. fragilis* and its hybrids. Occurrences on shrub willows usually constituted only a few individuals and were in places where *S. fragilis* was not present; perhaps indicating that feeding on these species occurred not by choice but necessity.

The distribution of sawfly in southeast Australia is more widespread than previously thought. The current disparate occurrences, many kilometres apart, probably indicate recent spread. In the Kiewa Valley (Victoria) the unsightly appearance of defoliated trees caused residents to contact the Catchment Management Authority (Bugelly 2006). It is possible that sawfly had been present in low densities around Kiewa for some time but the public response in early 2006 strongly suggests it had not previously achieved a significant population there.

In most instances, new detections of sawfly have consisted of only a few individuals indicating it may have recently spread to new areas but not yet built up in numbers. Given that the

Trees with sawfly State No. sites New distribution records for N. oligospilus present / No. present/trees sites sampled sampled ACT 9/12 21/25 Brindabella NSW 15/37 22/56 Adaminaby, Deniliquin, Jindabyne, Mullengandra, Tumbarumba, SA 10/24 14/29 Mount Compass, Murray Bridge, Strathalbyn, Tanunda, Victor Harbour, Williamstown, TAS 4/40 5/56 Deloraine, Devonport, Longford, Trevallyn (Launceston). Beechworth, Bendoc, Geelong, Kergunyah, Kiewa, Melton, VIC 8/101 14/170Rosedale, Tambo Crossing, Wodonga, Yea.

Table 1: Occurrence of the willow sawfly, N. oligospilus, in southeast Australia

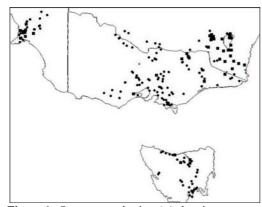


Figure 1 Survey sample sites (•) showing distribution of the willow sawfly *N. oligospilus* (•).

distribution has been extensively revised from only one season's survey, the rate of spread seems to be following the New Zealand example of up to 300km a year (Charles and Allen 2000). At the time of writing, there have been further unconfirmed reports of sawfly present in Armidale, Orange and Woomargama in NSW, and the King River Valley in Victoria (P. Ash, A. Briggs, S. Holland-Clift, *pers. comm.*).

Although we cannot yet report data on the incidence of other organisms associated with willows in Australia, the overwhelming impression is that the general level of herbivory is extremely low which is in direct contrast with *Salix* spp. from their country of origin (Sagliocco and Bruzzese 2001). The arrival of sawfly, therefore, constitutes a considerable change for willows in Australia as it may be the first time they are subjected to substantial herbivore injury. Given the very large sums expended in willow removal the likely impact of the sawfly requires further investigation.

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REFERENCES

Agriculture and Resource Management Council of Australian and New Zealand (ARMCANZ), (2001). 'Weeds of National Significance. National Strategy. Willows (*Salix* taxa)'.

- Anon. (2005a).Commonwealth of Australia, Office of the Chief Plant Protection Officer. 'Willow sawfly incident'. http://www.affa.gov.au/, 03/04/06.
- Anon. (2005b). 'CSIRO Entomology About Insects. Willow sawfly, *Nematus oligospilus*'. http://www.ento.csiro.au/about_insects/willows_ sawfly.html, 03/04/06).
- Bruzzese, E. and McFadyen, R. (2006). Arrival of the leaf-feeding willow sawfly *Nematus oligospilus* Förster in Australia–pest or beneficial? *Plant Protection Quarterly* 21, 43-4.
- Bugelly, C. (2006). Sawfly is savaging willow trees. Border Mail 21/04/06.
- Charles, J.G. and Allan, D.J. (2000). Development of the willow sawfly, *Nematus oligospilus*, at different temperatures, and an estimation of voltinism throughout New Zealand. *New Zealand Journal of Zoology* 27, 197-200.
- Charles, J.G., Allan, D.J. and Fung, L. (1988).
 Susceptibility of willows to oviposition by the willow sawfly, *Nematus oligospilus*.
 Proceedings of the 51st Plant Protection Conference, Hamilton, 11-13 Aug 1998. pp. 230-4.
- Cremer, K.W. (1996). Willow identification for river management in Australia. Technical Paper No. 3, pp 21 (CSIRO Division of Forestry, Canberra).
- Harman, H.M. (2004). Feasibility of biological control of grey willow Salix cinerea. DOC Science Internal Series 183, 29.
- Kennedy, S.A., Ganf, G.G. and Walker, K.F. (2003). Does salinity influence the distribution of exotic willows (*Salix* spp.) along the Lower River Murray? *Marine and Freshwater Research* 54, 825-31.
- Koch, F. and Smith, D.R. (2000). Nematus oligospilus Förster (Hymenoptera: Tenthredinidae), an introduced willow sawfly in the southern Hemisphere. Proceedings of the Entomological Society of Washington 102, 292-300.
- Naumann, I.D., Williams, M.A. and Schmidt, S. (2002). Synopsis of the Tenthredinidae (Hymenoptera) in Australia, including two newly recorded, introduced sawfly species associated with willows (*Salix* spp.) Australian Journal of Entomology 41, 1-16.
- Sagliocco, J-L and Bruzzese, E. (2001). 'Biological control of willows in Australia'. pp 61 (DNRE, Keith Turnbull Research Institute, Frankston).