

A Convenient Synthesis of Antibacterial Linezolid from (S)-Glyceraldehyde Acetonide

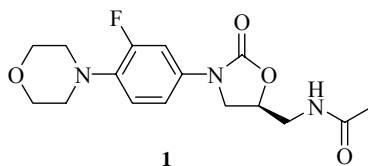
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Abstract: A convenient synthesis of oxazolidinone antibacterial linezolid from readily available L-ascorbic acid is described. The key steps include reductive amination of arylamine and (S)-glyceraldehyde acetonide in the presence of NaBH₄ and 4Å sieve, followed by hydrolysis and regioselective cyclization.

Keywords: Linezolid, antibacterial, synthesis, ascorbic acid.

Oxazolidinones as a new class of synthetic antimicrobial agents are active against numerous multidrug-resistant Gram-positive organisms¹. Linezolid **1** as the promising candidate of this family works effectively against numerous serious Gram-positive human pathogens as MRSA and VRE². Recently linezolid has been marketed in the USA and is available in oral and i.v. formulations.



Several methods are available for the synthesis of linezolid^{2,3}. Brickner² reported a route to linezolid with good yield. However, in this method for the synthetic key step to oxazolidinone ring, severe conditions with a low temperature (-78°C) and air-sensitive base (*n*-BuLi) were requested, which limit a large scale of production in industry. Lohray³ offered another possibility to linezolid *via* asymmetric bis-epoxide using D-mannitol as a starting material. However, the synthetic route was very long and without optical data.

We wish to report herein a very convenient and efficient synthesis of linezolid from readily available (S)-glyceraldehyde acetonide **3**. The synthetic route was outlined in **Scheme 1**. Reductive amination of fluoroaryamine **2**² with L-(S)-glyceraldehyde acetonide in the presence of NaBH₄ and 4Å molecular sieve in methanol afforded

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2 $\xrightarrow[\text{a } 76-78\%]{\text{3}}$ **4** $\xrightarrow[\text{b } 74-80\%]{\text{b}}$ **6** $\xrightarrow[\text{c } 88\%]{\text{c}}$ **7** $\xrightarrow[\text{e } 51\%]{\text{2 steps}}$ **1**

5a Y=OH
5b Y=OCOC1

dioxane **4**⁴ in good yields. (*S*)-Glyceraldehyde acetonide was easily obtained from L-ascorbic acid (vitamin C) *via* hydrogenation⁵, acetonation and oxidative cleavage⁶. The yield was considerably poor, when Pd/C or Raney Ni was used as catalyst in reductive amination step, due to the fact that unstable (*S*)-glyceraldehyde acetonide may polymerize under the condition of long reaction time. The use of NaBH₄/molecular sieve system can shorten the reaction time and minimize the polymerization of glyceraldehyde acetonide. After hydrolysis of dioxane **4** in aq. HCl/methanol, the regioselective cyclization with triphosgene in the presence of K₂CO₃ afforded oxazolidinone **5**⁷. However, if NaOH or triethylamine was used as acid scavenger, the byproduct **5b** can be formed. The key intermediate **5a** after side-chain manipulation² provided the targeted antibacterial linezolid⁸ in good yields.

Acknowledgments

References and Notes

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2. S. J. Brickner, D. K. Hutchinson, M. R. Barbachyn, *et al.*, *J. Med. Chem.*, **1996**, 39(3), 673.
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4. Spectral data: **4** ^1H NMR (CDCl_3 , δ_{ppm}): 6.84 (t, 1H, $J=8.2$ Hz), 6.39 (m, 2H), 4.35 (m, 8lines, 1H), 4.09 (dd, 1H, $J=7.1$, 8.3 Hz), 3.84 (t, 4H, $J=4.7$ Hz), 3.75 (dd, 1H, $J=6.0$, 8.2 Hz), 3.24 (dd, 1H, $J=3.7$, 12.3 Hz), 3.13 (dd, 1H, $J=6.6$, 12.5 Hz), 2.95 (brs, 4H), 1.44 (s, 3H), 1.37 (s, 3H). ^{13}C NMR (CDCl_3): δ 25.24, 26.86, 46.92, 51.73, 67.07, 67.12, 74.32, 101.70 (d, $J=24.35$ Hz), 108.50, 109.50, 120.25, 130.99, 144.7. MS: m/z 310 (M^+ , 58), 209 (100), 151 (16). $[\alpha]_{\text{D}}^{20}$ -1.7 (c 4.54, CHCl_3).
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7. **5a**: white powder (EtOAc/Hex.), m.p. 114–116°C, lit.² m.p. 112–114°C, Spectral data: ^1H NMR (CDCl_3 , δ_{ppm}): 7.47 (dd, 1H, $J=2.1$, 14.4 Hz), 7.12 (dt, 1H, $J=2.4$, 8.9 Hz), 7.00 (brs, 1H), 4.75 (m, 1H), 3.95 (m, overlapping, 3H), 3.88 (t, 4H, $J=4.7$ Hz), 3.76 (d, 1H, $J=13.1$ Hz), 3.08 (t, 4H, $J=4.2$ Hz), 2.35 (brs, 1H). MS: m/z 296 (M^+ , 100), 238 (55), 149 (20), 57 (36). Anal. Calcd for $\text{C}_{14}\text{H}_{17}\text{FN}_2\text{O}_4$: C, 56.75; H, 5.78; N, 9.45. Found: C, 57.02; H, 5.78; N, 9.28. $[\alpha]_{\text{D}}^{20}$ -53 (c 0.69, CHCl_3), lit.² $[\alpha]_{\text{D}}^{20}$ -54 (c 0.990, CHCl_3). **5b**: ^1H NMR (CDCl_3 , δ_{ppm}): δ 7.01 (m, 3H), 5.05 (m, 1H), 4.59 (t, 1H, $J=8.7$ Hz), 4.16 (dd, 1H, $J=6.6$, 9.1 Hz), 4.07 (dd, 1H, $J=3.6$, 14.7 Hz), 3.86 (m, 5H), 3.14 (t, 4H, $J=4.8$ Hz). MS: m/z 358 (M^+ , 100), 300 (72), 213 (52), 208 (26), 164 (28), 150 (57). Anal. Calcd. for $\text{C}_{15}\text{H}_{16}\text{ClFN}_2\text{O}_5$: C, 50.22; H, 4.50; N, 7.81. Found: C, 50.39; H, 4.50; N, 7.57. $[\alpha]_{\text{D}}^{20}$ -32 (c 0.650, CHCl_3).
8. **1**: white needles (EtOAc/Hex.), m.p. 179–180.5°C, lit.² m.p. 181.5–182.5°C. Spectral data: ^1H NMR (CDCl_3 , δ_{ppm}): 7.46 (dd, 1H, $J=2.2$, 14.6 Hz), 7.08 (dd, 1H, $J=1.8$, 9.1 Hz), 6.96 (t, 1H, $J=9.1$ Hz), 6.05 (t, 1H, $J=6.0$ Hz), 4.77 (m, 1H), 4.02 (t, 1H, $J=8.9$ Hz), 3.88 (t, 4H, $J=4.5$ Hz), 3.73 (dd, 1H, $J=1.8$, 7.0 Hz), 3.69 (dd, 1H, $J=3.2$, 5.9 Hz), 3.61 (dt, 1H, $J=5.5$, 8.5 Hz), 3.07 (t, 4H, $J=4.6$ Hz), 2.02 (s, 3H). MS: m/z 337 (M^+ , 27), 293 (25), 234 (20), 209 (26), 149 (50), 91 (100). Anal. Calcd. for $\text{C}_{16}\text{H}_{20}\text{FN}_3\text{O}_4$: C, 56.97; H, 5.98; N, 12.46. Found: C, 57.16; H, 6.03; N, 12.22. $[\alpha]_{\text{D}}^{20}$ -9 (c 1.52, CHCl_3). lit.² $[\alpha]_{\text{D}}^{20}$ -9 (c 0.919, CHCl_3).

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