ASTER YELLOWS



With the development of specialty cut flower production in Kansas, we are seeing some of the challenges that growers will face in production. The 1992 growing season will perhaps be remembered as the year it was difficult to grow statice and other flowers susceptible to aster yellows. Normally, each year low to moderate levels of aster yellows occur in flower production fields. However, 1992 losses throughout Kansas attributed to aster yellows ranged from 10–70 percent of certain flower plantings.

Aster yellows is caused by a mycoplasmalike organism, a microorganism intermediate between bacteria and viruses. While several strains of the organism have been reported, the eastern strain and the western or California strain are considered most common. The organism is located in the phloem tissues of infected plants and causes a variety of disorders such as distortion, discoloration, stunting and tissue proliferation. Eventually it may kill the plant.

Aster yellows has a broad host range (Table 1), encompassing more than 200 dicot plant species in more than 40 plant families worldwide. The disease is transmitted from plant to plant primarily by the aster leafhopper, *Macrosteles quadrilineatus* Forbes, (Homoptera: Cicadellidae). The highest plant infection rates are thought to be a result of infective aster leafhoppers migrating from southern states. Transmission by native aster leafhoppers acquiring the mycoplasmalike organism from local perennial or biennial plants is thought to account for only a small percentage of aster yellows infections.

Symptoms

Aster yellows has a range of characteristic symptoms which vary with the strain of aster yellows organism, timing of infection, plant species, temperature, age and/or size of the plant. Because of these variables, each plant infected with aster yellows may not display the same symptoms. This symptom variability, along with the broad host range and the habits of the insect vector (leafhopper), makes control difficult. Aster yellows can also be confused with symptoms of growth regulator herbicide damage and the occasional genetic disorder.

Initial symptoms of aster yellows usually appear as vein clearing, which spreads until the entire leaf becomes chlorotic. This is distinguished from nutrient deficiency, in which the veins remain green while the leaf blade becomes chlorotic. As the plant develops, the yellowing caused by aster yellows may spread, or it may remain restricted to one side or section of the plant, the remainder appearing healthy. Mature leaves generally do not change color, but new growth will be yellowishgreen. Infection early in the growing season will almost always cause stunting, shortened internodes, and dwarfed, deformed or lopsided flower heads. If infection occurs late in the season, the flowers will be deformed and remain yellowish-green regardless of the variety's normal color. With heavy infections, no flower production will occur.

One indicative symptom of aster yellows is adventitious shoot proliferation, which appears as a mass of leaves with a bushy or witch's broom effect in place of normal flower production. These leaves are yellowish-green and are smaller and thinner than normal. In late-season, the foliage may become red or bronze colored. Also, the root system of a plant with aster yellows is smaller than normal, predisposing the roots to field or storage diseases.

Aster Leafhoppers

Adult aster leafhoppers are 4mm, slender, wedge-shaped insects with wings held in a rooflike manner over their abdomens. Their nearly transparent wings have a slight tinge of color, ranging from straw-brown to light green. Their heads may have distinctive black spots, visible with the aid of a 10x hand lens. Or, there may not be any black spots. Individual leafhoppers are difficult to observe because of their small size. They are very skittish and fly away when approached. Observers report seeing "little, light-colored, gnat-sized bugs," not realizing they may have been seeing leafhoppers.

As is the case with all Homopterean insects, aster leafhoppers have piercing, sucking mouthparts (stylets), which they use to suck plant juices from deep phloem tissues. They spread salivary secretions while feeding. It is through this saliva that aster yellows is transmitted.

Aster Leafhopper Development. Leafhoppers are paurometabolous; their development is described as gradual or direct. They begin life in the egg stage. The nymphs which emerge from the eggs resemble the adult insects, but lack wings. Nymphs develop gradually, periodically shedding their outer skins as they grow. At maturity, nymphs shed their skins one more time, and pass directly to the adult stage.

This cycle begins when a female leafhopper uses her ovipositor to insert eggs into leaf tissues. Newly emerged nymphs exit the leaf and begin feeding. The duration of their five nymphal stages varies with the host plant and temperature, with more rapid development occurring at warmer temperatures. In general, development from egg to adult takes 21–35 days at temperatures from 68–82°F.

Migrant and Native Populations. Few specific details of aster leafhopper populations in Kansas have been documented. However, there are definitely two population types: migrant and native.

Migrants from south and southeast of Kansas in portions of Texas, Louisiana and Oklahoma ride southerly winds into Kansas perhaps as early as mid-March. The heaviest concentration of migrant leafhoppers typically arrives in mid- to late April. These migrants rest and feed before leaving Kansas as they continue their northward migration.

Native aster leafhopper populations arise from overwintered eggs deposited the previous fall in winter grains—mainly wheat. These eggs do not hatch until late spring, after the first wave of migrants arrive in Kansas. The leafhopper nymphs begin their development on wheat plants. As the wheat matures, they seek alternate host plants flower crops—upon which to complete development. There is a complete second generation in Kansas. It is not known whether the second generation or possibly a third generation deposits overwintering eggs.

Aster Yellows Transmission

Once an aster leafhopper feeds on and acquires the mycoplasmalike organism from an infected plant, 3 weeks will elapse before that leafhopper can transmit the disease to another plant. During this incubation period, the mycoplasmalike organism multiplies within the leafhopper and moves to the salivary glands, from which it will be injected into healthy plants as the leafhopper feeds. Some of the migrants which acquired the aster yellows organism in their overwintering area may have passed through the 3-week incubation period by the time they reach Kansas. Any plants in the field, likely to be small at this time, are thus susceptible to infection when the leafhoppers rest and feed before resuming their northward migration.

In Kansas, native leafhoppers are probably less important in aster yellows transmission. In wheat fields, first generation nymphs emerging during late spring do not feed on infected plants during their development. As adults, they would have to encounter an inoculum source outside the grain field and then pass through the 3-week incubation period before becoming infective. Because flowering plants are larger later in the season, the initial amount of mycoplasmalike organism injected into the plant might not be sufficient to cause symptoms immediately. The organism would have to propagate to a high enough concentration in the plant to adversely affect flower production. Similarly, second-generation leafhoppers are of minor importance in transmitting aster yellows. The mycoplasmalike organism is not transmitted from parent to offspring through the egg. Thus secondgeneration nymphs or adults would have to feed on infected plants to acquire the organism. By the time they would become infective, most annuals used as specialty cut flowers already would have been harvested.

Infectivity Rates of Migrant Aster Leafhoppers

University of Wisconsin entomologists have developed an Aster Yellows Index on which vegetable producers in Wisconsin base their decisions to spray for aster leafhoppers. Decisions are based on a crop's particular level of susceptibility to aster yellows, on the number of leafhoppers present as determined by sweep net sampling, and on the percent infectivity rate. Generally, infectivity rates are less than 1 percent. In some years, infectivity rates may be 4-5 percent. The infectivity rate in 1992 was 10 percent, the highest in 30 years. This index might be useful to cut flower growers in northern states who would have information on infectivity rates well before the arrival of the migrant aster leafhoppers. Cut flower growers in southern regions such as Kansas are too close to the source of migrant leafhoppers to use the information.

Treatment Options

The variable nature of the disease and the sporadic way it spreads make control of aster yellows difficult. Control requires an integrated management approach, combining early detection and monitoring, eradication of infected plant material, and control of the aster leafhopper.

Monitor the flower crop and surrounding areas at regular intervals during the growing season, examining the crop for disease symptoms and aster leafhoppers. Rogue infected plants and remove piles of infected plant material from the field. Control weeds in and around the field to prevent infection of an alternate host and overwintering of the disease. Avoid planting annuals near susceptible perennial or biennial plants in the field.

A preventive strategy is to apply the systemic insecticide DI-SYSTON 15% Granular at planting time. If plants are seeded directly into the ground, apply the insecticide as an over-the-row band treatment. If plants are transplanted into the field, apply the insecticide as a side-dress treatment. This treatment should kill aster leafhoppers for 45-60 days, before they can successfully transmit the aster yellows mycoplasma. Treatment and residual effectiveness depend on moisture availability in the soil; if the soil becomes too dry, the insecticide will not be in solution for uptake into the plant via the root system. Irrigate periodically to better ensure effectiveness. As noted on the label, when applying DI-SYSTON 15% Granular by hand, wear heavy rubber or latex gloves with no holes or tears, protective clothing and footwear.

Some foliar sprays registered for use against leafhoppers on ornamentals in 1993 include formulations with the following active ingredients: azinophosmethyl (Guthion), bifenthrin (Talstar), carbaryl (Sevin), chlorpyrifos (Dursban and Pageant), cyfluthrin (Tempo and Decathelon), diazinon, ethyl parathion (Parathion 25WP), fluvalinate (Mavrik) and permethrin (Ambush and Pounce). Apply treatments when leafhoppers appear in the field, and reapply as needed. Read labels to determine whether specific products are registered for use against leafhoppers on target ornamentals. Follow label directions on use rates and phytotoxicity.

Aster Yellows		
Family: Genus/species	Common name	
Amaranthaceae		
Amaranthus retroflexus	Rough pigweed	
Apiaceae		
Anethium graveolens	Dill	
Apium graveolens	Celery	
Apium graveolens rapaceum	Celeriac	
Coriandrum sativum	Coriander	
Daucus carota	Carrot	
Pastinaca sativa	Parsnip	
Apocynaceae		
Catharanthus roseus	Periwinkle	
Asclepiadaceae		
Asclepias nivea	Common milkweed	
Asteraceae Compositae		
Anthemis cotula	Mayweed	

 Table 1. A Partial List of Plants that can be Infected with

 Aster Yellows

Family: Genus/species

> Aphanostephus humilis Bidens frondosa Bidens pilosa Brachycome iberidifolia Calendula officinalis Centaurea americana Centaurea cyanus Chrysanthemum carinatum Chrysanthemum cinerariifolium Chrysanthemum coronarium *Chrysanthemum frutescens* Chrysanthemum segetum Conyza canadensis Cichorium endivia Cichorium intybus Coreopsis grandiflora Coreopsis lanceolata Cosmos bipinnatus Dyssodia wrightii Erigeron canadensis Erigeron linifolius Erigeron philadelphicus Gaillardia pulchella Galinsoga parviflora Gnaphalium decurrens Gnaphalium ramosissimum Helenium autumnale

Helenium latifolium

Asteraceae

Helenium nudiflorum Helenium puberulum Helianthus annus Helichrysum bracteatum Hemizonia corumbosa Lactuca spp. (altaica, canadensis, floridana, graminifolia, indica, perenis, muralis, raddeana, saligna, spicata, squarrosa, virosa) Lactuca sative Lactuca scariola var. integrata Leontodon autumnalis Matricaria suareolens Parthenium hysterophorus Lazy daisy Beggar-ticks Hairy bur marigold Swan River daisy Pot marigold Basket flower Cornflower, Bachelor's button Tricolor chrysanthemum

Common name

Pyrethrum Crown daisy Marguerite daisy Corn chrysanthemum Horseweed Endive Common chicory Tickseed Tickseed Cosmos Fetid marigold Horseweed Flax-leaved fleabane Philadelphia fleabane Annual blanket flower Small flower galinsoga California everlasting Pink everlasting Common sneezeweed, Helen's flower Sneezeweed

Purple sneezeweed Rosilla Common sunflower Strawflower Coast tarweed

Lettuce Garden lettuce Prickly lettuce Fall dandelion Pineapple weed Santa Maria

Family: Genus/species Common name Picris echioides Bristly oxtongue False dandelion Pyrrhopappus multicaulis Rudbeckia hirta Hairy coneflower, Black-eyed Susan Scorzonera hispanica Black salsify Senecio vulgaris Common groundsel Sonchus oleraceus Common sowthistle Tagetes erecta African or American marigold Tagetes patula French marigold Tragopogon dubius Western salsify Tragopogon porrifolius Oyster plant Verbesina enceliodes Crownbeard Zinnia elegans Zinnia Begoniaceae Begonia semperflorens Wax begonia Boraginaceae Myosotis scorpiodes Forget-me-not Brassicaceae (Cruciferae) Armoracia rusticana Horseradish Common yellow mustard Brassica campestris Brassica oleracea var. botrytis Cauliflower Brassica oleracea var. capitata Cabbage Broccoli Brassica oleracea var. italica Capsella bursa-pastoris Shepherd's purse Wallflower Cheiranthus cheiri Raphanus sativus Radish Rorippa curvisiliqua Western yellow cress Sisymbrium irio Mustard Campanulaceae Lobelia erinus var. compacta Edging lobelia Caricaceae Carica papaya Papaya Caryophyllaceae Dianthus barbatus Sweet William Dianthus caryophyllus Carnation Gysophila paniculata Baby's breath Spergula arvensis Corn spurry

Chenopodiaceae Spinacia oleracea

Stellaria media

Spinach

Common chickweed

Family: Genus/species	Common name	Family: Genus/species	Common name		
Cistaceae		Onagraceae	Onagraceae		
Helianthemum chamaecistus	Rockrose	Clarkia concinna	Red ribbons		
Cucurbitaceae		Clarkia unguiculata	Clarkia, farewell-to-spring godetis		
Cucurbita muschata	Musley cound	Epilobium californicum	California willow herb		
	Musky gourd	Epilobium paniculatum	Panicled willow herb		
Cucurbita pepo	Pumpkin	Gaura lindheimeri	White guara		
Datiscaceae					
Datisca cannabina	Akalbir	Papaveraceae Eschscholzia californica	California poppy		
Dipsacaceae					
Dipsacus fullonum	Fuller's teasal	Plantaginaceae			
Scabiosa atropurpea	Pincushion flower,	Plantago major	Great plantain		
Fabaceae (Leguminosae)	sweet scabious	Plumbaginaceae			
	Deep alassa	Limonium sinuatum	Annual statice		
Medicago hispida	Bur-clover	Limonium sinuatum			
Trifolium fragiferum	Strawberry clover				
Trifolium hybridum	Alsike clover	Poaceae (Gramineae)	_		
Trifolium pratense	Red clover White clover	Avena sativa	Oats		
Trifolium repens	white clover	Delement			
Geraniaceae		Polemoniaceae	Claba -: 1: -		
		Gilia capitata	Globe gilia		
Erodium cicutarium Erodium moschatum	Redstem filaree Whitestem filaree	Phlox drummondii	Annual phlox		
		Polygonaceae			
Gesneriaceae		Polygonum convolvulus	Black bindweed		
Didymocarpus horsfeldii		Rumex acetosella	Sheep sorrel		
Iridaceae		Portulacaceae			
Gladiolus x hortulanus	Gladiolus	Calandrinia grandiflora	Rock purslane		
Gradionas x normannas	Gladiolas	Portulaca oleracea	Purslane		
Labiatae (Lamiaceae)			T distance		
Lamium amplexicaule	Dead henbit nettle	Primulaceae			
Salvia azurea	Azure sage	Anagallis arvensis	Scarlet pimpernel		
	6	Primula polyantha	Primula		
Liliaceae		r - y			
Allium ascalonicum	Shallot	Ranunculaceae			
Allium cepa	Onion	Anemone coronaria	Poppy anemone		
····· · · · · · ·		Consolida (Delphinium) ajacis	Rocket larkspur		
Loasaceae		Delphinium x cultorum	Hybrid larkspur		
Blumenbachia hieronymii		Nigella damascena	Love-in-a-mist		
Cajophora lateritia		Ranunculus asiaticus	Persian buttercup		
Malvaceae		Rosaceae			
Malva parviflora	Little mallow	Fragaria x ananassa	Garden strawberry		
	Little manow	1 / againa n ananassa	Cardon Suurborry		

Family:			References	References	References
Genus/species	Common name				Agrios, George N. 1988. Plar
Scrophulariaceae					Edition. Academic Press, Inc.
Linaria bipartita	Clover-lip toad flax				Beirne, B.P. 1952. The Nearti Macrosteles (Homoptera: Cicade
Linaria canadensis	Oldfield toad flax				Entomologist. Vol. 84 No. 7.
Mimulus cardinalis	Scarlet monkey-flower				Daughtrey, Margery, and A.
Mimulus guttatus	Common monkey-flower				Field Guide to Diseases of Green
Veronica americana	American speedwell			tals. Ball Publishing.	
Veronica buxbaumii	Byzantine speedwell			e	Fordberg, Junius L. 1975. Dis
Solanaceae					tal Plants. University of Illinois F
Lycopersicon esculentum			,	,	Horst, R. Kenneth. 1990. We
(L. lycopersicum)	Tomato				Disease Handbook. Fifth Edition
Nicotiana rustica	Wild tobacco		Reinhold.		
Petunia x hybrida	Garden petunia				Migration of the Six-Spotted Macrosteles fascifrous. 1965. Resea
Salpiglossis sinuata	Painted-tongue				University of Wisconsin, Madiso
Solanum nigrum	Black nightshade				Pirone, Pascal P. 1978. Disea
Solanum tuberosum	Potato				Ornamental Plants. Fifth Edition
	1 0 0000		Sons.		
Tropaeolaceae					Wallis, R.L. 1960. Host Plant
Tropaeolum majus	Garden nasturtium				Leafhopper and the Aster Yellow
1 5					Vectors of the Virus. USDA-ARS
Urticaceae					Whitcomb, Robert F., and Jo
Urtica californica	Nettle				The Mycoplasmas, Volume V Sp
·					Acholeplasmas, and Mycoplasm
			Arthropods	Arthropods. Academic	Arthropods. Academic Press, Inc
					Brand names appearing in this p
			·	1	product identification. No endorseme
					criticism of similar products not men
					such products assume responsibility i accordance with current label direction
		turer.			

About the authors:

Judith O'Mara is an Extension Diagnostician, Plant Pathology; Robert Bauernfeind is an Extension Specialist, Entomology; Alan Stevens is an Extension Specialist, Floriculture and Ornamental Horticulture; Karen L.B. Gast is Extension Specialist, Post Harvest and Marketing; and Susan Stevens is a Research Assistant, Horticulture.



Cooperative Extension Service, Manhattan, Kansas

AF-1086 October 1993	MF
ssued in furtherance of Cooperative Extension Work, acts of May 8 and June 30, 1914, as amended. Kansas State University, County	Issue
extension Councils, and United States Department of Agriculture Cooperating, Richard D. Wootton, Associate Director. All	Exte
ducational programs and materials available without discrimination on the basis of race, color, national origin, sex, age, or disability.	educ
ile Code: Horticulture and Landscaping (Commercial) JH10-93–2M	File