## TCID 50 protocol

1. Put A6 cells in 96 well plates at 5000 cells $/$ well in $\mathrm{ASF}+10 \%$ FCS (100ul total) for about 2 days.
2. In a separate V bottom 96 well plate, do dilutions of virus:
A. 1:25- Add 8ul of virus in 192ul of ASF-A6 or ASF
B. 1:50- Add 60 ul of 1:25 in 60 ul ASF-A6 or ASF
C. 1:500- Add 12ul of 1:50 in 108ul ASF-A6 or ASF
D. 1:5000- Add 12 ul of 1:500 in 108ul media and so on until dilution H. $5 \times 10^{7}$
*Note: Make sure you resuspend the wells very well before going to the next dilution!!
3. Transfer 100 ul of each dilution to the plate with A6 to make a total of 200 ul per well.
*Remember to leave columns 1 and 2 without virus, these are the control.
4. Leave in incubator at 37 C with CO 2 for approx. 5 days.
5. Check daily.

## TCID 50 Calculation

Example


1. Calculate Proportionate Distance (PD) between the two dilutions in between 50\% death: (\% next above 50\%)- 50\% / (\% next above 50\%) - (\% next below 50\%)

Example above:

$$
\mathrm{PD}=80 \%-50 \% / 80 \%-0 \%=30 / 80=.375
$$

2. Calculate $50 \%$ end point. Log lower dilution= dilution in which position is next above 50\%

Example above:
Log lower $=10^{-6}$ or -6

## 3. Add PD and Log lower dilution

Example above: $-6+.375=-6.375$
Log TCID50 $=10-6.375$ or $1 / 2.37 \times 10^{6}$
4. Calculate TCID 50/ml. Divide by the ml of viral innoculum added to row A

Example above: according to our protocol $=.008 \mathrm{ml}$
TCID $50 / \mathrm{ml}=2.37 \times 10{ }^{6} .008=2.9 \times 10^{8}$
5. Calculate PFU/ml. Divide by constant.

Example above: $2.37 \times 10^{6} \times .69=2.0 \times 10^{8} \mathrm{PFU} / \mathrm{ml}$

