# Genetic dissection of plant height by molecular markers using a population of recombinant inbred lines in maize 

Tang Ji-hua $\cdot$ Teng Wen-tao $\cdot$ Yan Jian-bing $\cdot$ Ma Xi-qing • Meng Yi-jiang • Dai Jin-rui Li Jian-Sheng

Received: 13 May 2006 / Accepted: 17 November 2006/Published online: 12 December 2006
© Springer Science+Business Media B.V. 2006


#### Abstract

Plant height is an important trait for maize breeding because it is related to planting density and lodging resistance. It is influenced by many qualitative genes and quantitative trait loci (QTL). In this study, the genetic basis of plant height and its related traits were dissected, using simple sequence repeat (SSR) markers with a maize population of 294 recombinant inbred lines (RIL). Correlation results showed that plant height had a significant positive correlation with leaf number, average internode length and internode number. Increased plant height was affected most by average internode length. Six QTL for plant height were detected, which were consistent with those reported in previous studies. Moreover, eight QTL for leaf number, seven for internode number and six for average internode length were identified. Four of six QTL detected for average internode length were located on the same chromosomal region as the QTL affecting


[^0]plant height and shared common molecular markers. This latter result strongly suggests that average internode length was the main contributor to plant height in maize.

Keywords Plant height • Average internode length • QTL • Maize (Zea mays L.)

## Abbreviations

QTL quantitative trait loci
RIL recombinant inbred line
SSR simple sequence repeats
LOD logarithm of odds
PH plant height
LN leaf number
IN internode number
AIL average internode length

## Introduction

Plant height is an important trait for maize breeding. It is significantly correlated to grain yield, and is affected by many qualitative genes and quantitative trait loci (QTL; Beavis et al. 1991). In order to increase grain yield, the planting density of maize is increasing gradually in the major areas of maize production throughout the world, but some hybrids with tall plants
are often susceptible to lodging, which is always accompanied by reduction of grain output. To avoid plant lodging, the main breeding strategy is to decrease plant height, enhance stalk strength, and/or increase brace roots.

Biologically, plant height is comprised of two main components: internode number and average internode length. With the development of molecular genetics, it was widely used as a model trait for dissecting the genetic basis of QTL. Beavis et al. (1991) reported that most QTL detected for plant height were in close proximity to mapped qualitative genetic loci for the trait. Plant height was associated with 11 chromosomal regions in the study by Berke and Rocheford (1995). To date, numerous QTL for plant height were identified in a number of studies (Schön et al. 1993; Koester et al. 1993; Veldboom and Lee 1994, 1996; Lance and Lee 1996; Kozumplik et al. 1996; Lübberstedt et al. 1997; Austin et al. 2001, Yan et al. 2003). In these reports, only the total plant height in the mature stage of development was used as criteria for genetic analysis while few reports have analyzed the components contributing to plant height in maize.

In this study, the main objectives were to characterize QTL for plant height, and its components with a population of recombinant inbred lines (RIL) using molecular markers, and to analyze the relationship of plant height to its components.

## Materials and methods

## Materials

A population of 294 inbred lines (RIL) was selected for analysis. It was derived from a cross between two inbred lines, Z3 and 871, using a single seed descent method until the F8 generation. The cross was an elite hybrid cultivar name Yuyu22, which was planted in about 1.7 million hectares per year during 2002-2004 in China. The inbred line $\mathrm{Z} 3\left(\mathrm{P}_{1}\right)$ is dent, selected from a synthetic population with Chinese domestic germplasm; the other parental line, $871\left(\mathrm{P}_{2}\right)$, is flint, selected from an exotic germplasm.

Field evaluation
The RIL population, as well as the two parents, was evaluated on the agronomy farm at China Agricultural University (Beijing, China) over 3 years (2002-2004). The field experiment followed a randomized complete block design with three replications in each year. Each plot included one row that was $4-\mathrm{m}$ long and $0.67-\mathrm{m}$ wide with a density of 45,000 plants per hectare.

Fifteen plants were planted in each plot, among which, ten consecutive plants starting from the third plant (i.e., plants $3-12$ in the row) were used for trait measurements. In seedling stage, the fifth and tenth leaf of sampled plants were designated as markers for evaluating leaf number in the field. After pollen shedding, the same ten plants, which were used to measure leaf numbers, were evaluated for plant height, internode numbers and average internode length. Plant height was measured from the ground to the top of tassels. The internode number was defined as the number of elongated above-ground internodes. The average internode length was characterized as average length of the above-ground internodes, and was calculated as plant height divided by internode numbers. The average performance data over 3 years was employed as input data for further analyses. Estimates of means and variances for the measured traits were conducted using SAS software (SAS Institute 1996). The broad-sense heritabilities and confidence intervals of measured traits were computed according to Knapp et al. (1985), the estimates of $\sigma_{\mathrm{g}}^{2}, \sigma_{\mathrm{g} \times \mathrm{y}}^{2}$, and $\sigma_{\mathrm{e}}^{2}$ were obtained from the linear model in a two-way analysis of variance (ANOVA).

Construction of molecular linkage maps and QTL analysis

A total of 846 pairs of simple sequence repeat (SSR) markers were selected from the maize genome database (www.maizegdb.org) in order to screen polymorphisms between two parents. Of these, 285 SSR markers had distinct polymorphisms between two parents and were used to amplify the DNA of 294 RILs. Molecular linkage maps were constructed using Mapmakers 3.0
(Lander et al. 1987) at a LOD threshold greater than 3.0.

The composite interval mapping method, Mapmaker 3.0 (Zeng 1994) was employed for mapping QTL of measured traits using the average values of 3 years by the software QTL Cartographer (Wang et al. 1994). Model 6 of the Zmapqtl module was used to identify QTL, scanning intervals of 2 cM between markers and putative QTL with a window of 10 cM . The number of marker cofactors for background control was set by forward-backward stepwise regression with five controlling markers. A genome-wide critical threshold value for the experiment wise type I error rate $(\alpha=0.05)$ was set for each trait independently by randomly permuting 1,000 times. For main QTL effects, positive and negative signs of the estimates indicated that $\mathrm{Z3}$ and 871 contributed toward higher value alleles for the traits.

## Results

Performance of plant height and its related traits

Among the four traits measured, some differences between the two parents were identified (Table 1). Parent Z3 ( $\mathrm{P}_{1}$ ) had fewer internodes than parent $871\left(\mathrm{P}_{2}\right)$ and $\mathrm{P}_{1}$ was 9.6 cm shorter
than $\mathrm{P}_{2}$. The values of the measured traits among the RIL population varied widely: ranges for plant height were $128-242 \mathrm{~cm}$, for leaf number 16-23, for internode numbers 10.9-17.1 and for average internode length $9-19 \mathrm{~cm}$. The averages for each of these traits were 19.8, 14.3, 13.4, and 190.7, respectively (Fig. 1; Table 1). The broad heritability for leaf numbers, internode numbers, average internode length, plant height reached $94,92.8,92.9$, and $92.8 \%$, respectively (Table 1). The interaction between genotype and year for the measured traits was not significant (data not shown).

The four measured traits were phenotypically correlated with each other (Table 2). Plant height had the strongest positive correlation to average internode length ( $r=0.77$ ) and was weakly correlated to internode $(r=0.38)$ and leaf $(r=0.22)$ number ( $p<0.01$ for each). Leaf number had a very strong positive correlation to internode number ( $r=0.92$ ) and a weak one with average internode length ( $r=-0.39 ; p<0.01$ for each). Internode number and average internode length ( $r=-0.28$ ) were weakly correlated ( $p<0.01$ ).

## QTL analysis

A genetic linkage map was constructed using 263 SSR markers that covered the whole genome of maize, spanning $2,361 \mathrm{cM}$ length with an average interval of 9 cM between markers. These charac-

Table 1 Plant height and its related traits in the RIL population and its parents

| Population | Trait |  | LN | IN | AIL (cm) | PH (cm) |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- |
| $\mathrm{P}_{1}^{\mathrm{a}}$ |  |  | 18.0 | 12.8 | 14.4 | 179.5 |
| $\mathrm{P}_{2}^{\mathrm{a}}$ |  | 19.2 | 14.0 | 13.5 | 189.1 |  |
| RIL population |  | Average | $19.8 \pm 1.2$ | $14.3 \pm 1.1$ | $13.4 \pm 1.5$ | $190.7 \pm 22.5$ |
|  | Ranges | $15.8-23.1$ | $10.9-17.1$ | $9.4-18.5$ | $127.5-242.2$ |  |
|  | $\sigma_{\mathrm{g}}^{2} \mathrm{~b}$ | 1.28 | 0.98 | 1.94 | 415.23 |  |
|  | $\sigma_{\mathrm{og}_{\mathrm{b}} \mathrm{b}}^{2}$ | 0.16 | 0.13 | 0.32 | 80.07 |  |
|  | $\sigma_{\mathrm{b}}$ | 0.25 | 0.29 | 0.39 | 50.90 |  |
|  | $\mathrm{H}_{\mathrm{B}}^{2} \mathrm{c}$ | 94.00 | 92.80 | 92.90 | 92.80 |  |
|  |  | $92.9-94.9$ | $91.4-0.93 .8$ | $91.6-94.0$ | $91.4-93.9$ |  |

[^1]

B: IN


**

## C:AIL



D:PH


Fig. 1 Frequency distributions of a leaf number, b internode number, c average internode number, and d plant height in RIL population. a $L N$ leaf number, b $I N$ internode number, c $A I L$ average internode length, d $P H$ plant height

Table 2 Correlation coefficients between plant height and its related traits in RIL population

| Trait | IN | AIL | PH |
| :--- | :--- | :--- | :--- |
| LN | $0.92^{*}$ | $-0.39^{*}$ | $0.22^{*}$ |
| IN |  | $-0.28^{*}$ | $0.38^{*}$ |
| AIL |  |  | $0.77^{*}$ |

$L N$ leaf number, $I N$ internode number, $A I L$ average internode length, $P H$ plant height

* $p<0.01$
teristics were consistent with linkage maps published in the maize genome database (www.maizegdb.org).

As shown in Table 3, 27 QTL were detected for the four measured traits using the composite interval mapping method. For plant height, six QTL were detected, located on chromosomes 1, 2, 3, and 5. The QTL, qPH1a, located on chromosome 1, between markers umc2151-bnlg1556, had the highest additive effect with a value of 11.34 cm . It accounted for $24.53 \%$ of the phenotypic variance. Another QTL, $q P H 2$, explained $16 \%$ of the phenotypic variance. The inbred line of 871 alleles at QTL $q P H 1 a, q P H 1 b, q P H 2, q P H 5 a, q P H 5 b$ and the alleles from $\mathrm{Z3}$ at QTL $q P H 3$ were associated with increased plant height. The total QTL effects detected for plant height accounted for $71 \%$ of the phenotypic variance.

Eight QTL were detected for leaf number, and were located on chromosomes $1,3,4$, and 9 . The QTL, qLN9a and $q L N 9 b$, accounted for 14 and $11 \%$ of the phenotypic variance, respectively. Among the mapped QTL, six alleles derived from inbred line 871 were associated with increased leaf number. Only two alleles from Z 3 were associated with increased leaf number. The total QTL effects detected for leaf number explained $69 \%$ of the phenotypic variance.

For internode numbers, seven QTL were detected, five of which were located on chromosome 1 and the others on chromosomes 6 and 7. The QTL, qIN1e and qIN7 had relatively large contribution rates ( $R^{2}$ ), accounting for 11 and $10 \%$ of the phenotypic variance, respectively. Six alleles from the line 871 were associated with increased internode numbers and only one allele derived from Z3 was related to increased internode numbers. In all, $53 \%$ of the phenotypic

Table 3 Codes, locations, genetic effects, and phenotypic variation $\left(R^{2}\right)$ of putative QTL for plant height and its related traits

| Traits | QTL | Flanking markers | Bin | Logarithm rate (LOD) | $\mathrm{A}^{\text {a }}$ | $R^{2 \mathrm{~b}}$ | $\mathrm{LOD}_{0.05}^{\mathrm{c}}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Plant height | $q$ PH1a | umc2151-bnlg1556 | 1.07 | 13.52 | -11.3 | 24.53 | 3.3 |
|  | qPH1b | umc2029-phi011 | 1.08 | 3.35 | -6.06 | 7.17 |  |
|  | qPH2 | umc2372-umc1497 | 2.07 | 5.11 | -9.11 | 16.2 |  |
|  | qPH3 | umc2166-umc1311 | 3.05 | 5.72 | 6.41 | 8.11 |  |
|  | qPH5a | umc1692-umc2373 | 5.03 | 5.14 | -5.79 | 6.59 |  |
|  | $q$ PH5b | umc1155-bnlg1237 | 5.05 | 3.94 | -6.58 | 8.56 |  |
| Leaf number | qLN1a | umc1590-umc1035 | 1.06 | 3.82 | -0.34 | 7.44 | 3.1 |
|  | $q L N 1 b$ | umc1335-umc1122 | 1.07 | 4.58 | -0.36 | 8.71 |  |
|  | qLN1c | bnlg1643-bnlg1597 | 1.09 | 3.45 | 0.31 | 6.54 |  |
|  | qLN3a | umc1223-umc1773 | 3.04 | 4.9 | -0.31 | 6.73 |  |
|  | $q L N 3 b$ | umc2002-bnlg1035 | 3.04 | 5.27 | -0.31 | 6.32 |  |
|  | $q L N 4$ | phi213984-umc2082 | 4.02 | 4.45 | 0.34 | 7.95 |  |
|  | qLN9a | bnlg127-bnlg 1208 | 9.03 | 6.89 | -0.46 | 14.29 |  |
|  | $q L N 9 b$ | bnlg1208-umc1771 | 9.04 | 7.6 | -0.41 | 11.25 |  |
| Internode number | qIN1a | phi427913-phi001 | 1.02 | 4.36 | -0.27 | 6.25 | 3.1 |
|  | qIN1b | phi001-bnlg1484 | 1.03 | 4.72 | -0.32 | 8.76 |  |
|  | qIN1c | umc2112-umc1689 | 1.05 | 3.31 | -0.23 | 4.11 |  |
|  | qIN1d | umc1590-umc1035 | 1.06 | 3.57 | -0.32 | 7.45 |  |
|  | qIN1e | umc2151-umc1122 | 1.07 | 5.06 | -0.37 | 10.82 |  |
|  | qIN6 | umc1020-phi299852 | 6.07 | 4.47 | 0.26 | 6.13 |  |
|  | qIN7 | bnlg1792-mmc0411 | 7.02 | 4.97 | -0.34 | 9.91 |  |
| Average internode length | qAIL1 | umc2029-phi011 | 1.09 | 6.95 | -0.62 | 15.96 | 3.2 |
|  | qAIL3a | umc1773-bnlg1035 | 3.04 | 5.7 | 0.39 | 6.5 |  |
|  | qAIL3b | umc2127-umc2166 | 3.05 | 6.4 | 0.51 | 11.29 |  |
|  | qAIL5a | umc1692-umc2373 | 5.03 | 3.77 | -0.33 | 4.31 |  |
|  | qAIL5b | umc1155-umc1019 | 5.05 | 4.91 | -0.49 | 10.4 |  |
|  | qAIL9 | bnlg1209-umc1771 | 9.04 | 4.45 | 0.38 | 6.05 |  |

${ }^{\text {a }}$ Additive effect: positive values indicate that the Z 3 alleles are increasing the traits and vice versa for negative values
${ }^{\mathrm{b}} R^{2}$ contribution rate
${ }^{\text {c }} \mathrm{LOD}_{0.05}$ logarithm of odds at $p<0.05$
variance for the internode number accounted was explained by detected QTL.

Six QTL were detected for average internode length and were located on chromosomes $1,3,5$, and 9. The QTL qAIL1, qAIL3b, and qAIL5b had relatively large contribution rates $\left(R^{2}\right)$, accounting for 16,11 , and $10 \%$ of the phenotypic variance, respectively. The alleles from the line 871 at QTL $q A I L 1, q A I L 5 a$, and $q A I L 5 b$ were associated with increased average internode length. The total QTL effects detected for average internode length explained $55 \%$ of the phenotypic variance.

## Discussion

Compared with previous reports (Table 4), all six QTL for plant height detected in this study
seemed to have similar, or the same, chromosomal locations with different mapping populations. Lübberstedt et al. (1997) identified 20 QTL for plant height using two testcross population of $\mathrm{F}_{3}$ lines, including the six QTL detected in this study. Four QTL detected by Melchinger et al. (1998) were situated at the same chromosomal loci as the QTL reported in this study. One QTL, $q P H 5 a$, is located in the same chromosomal region as many QTL for plant height identified in other independent studies with at least seven different mapping populations in maize (Beavis et al. 1991; Ajmone-Marsan et al. 1994; Schǒn et al. 1994; Berke and Rocheford 1995; Kozumplik et al. 1996; Lübberstedt et al. 1997; Melchinger et al. 1998). In other studies, $q P H 1 a$ and $q P H 1 b$ (Schǒn et al. 1994), and $q P H 3$ and $q P H 5 b$ (Abler et al. 1991; Ajmone-Marsan et al. 1994) were also

Table 4 Comparison of QTL locations for plant height between this study and previous studies

| QTL ${ }^{\text {a }}$ | Chromosomal bin ${ }^{\text {a }}$ | Linked markers ${ }^{\text {b }}$ | Reference |
| :---: | :---: | :---: | :---: |
| qPH1a | 1.07 | umc23a-umc58 | Schǒn et al. (1994) |
|  |  |  | Lübberstedt et al. (1997) |
|  |  |  | Melchinger et al. (1998) |
| $q P H 1 b$ | 1.08 | umc83a | Beavis et al. (1991) |
|  |  | bnl15.18-umc37a | Schǒn et al. (1994) |
|  |  | isu18-umc37a | Veldboom and Lee (1994) |
|  |  | an1 anther ear1 | Veldboom and Lee (1996) |
|  |  | an1 anther ear1 | Veldboom and Lee (1996) |
|  |  |  | Lübberstedt et al. (1997) |
| $q \mathrm{PH} 2$ | 2.07 | php20005-umc131 | Beavis et al. (1994) |
|  |  |  | Lübberstedt et al. (1997) |
|  |  |  | Melchinger et al. (1998) |
| $q \mathrm{PH} 3$ | 3.05 | bn15.37a-csu154b(eif5A) umc26a-umc42b | Abler et al. (1991) |
|  |  |  | Ajmone-Marsan et al. (1994) |
|  |  |  | Kozumplik et al. (1996) |
|  |  |  | Lübberstedt et al. (1997) |
|  |  |  | Melchinger et al. (1998) |
| $q P H 5 a$ | 5.03 | bnl7.56 <br> bnl5.71a-umc1 umc27a-umc90 | Beavis et al. (1991) |
|  |  |  | Ajmone-Marsan et al. (1994) |
|  |  |  | Schǒn et al. (1994) |
|  |  |  | Berke and Rocheford (1995) |
|  |  |  | Kozumplik et al. (1996) |
|  |  |  | Lübberstedt et al. (1997) |
|  |  |  | Melchinger et al. (1998) |
| $q P H 5 b$ | 5.05 | bnl5.71a-umc1 | Ajmone-Marsan et al. (1994) |
|  |  |  | Lübberstedt et al. (1997) |

[^2]identified. These demonstrate that a large number of QTL influence plant height in maize and some of them may have a common origin.

Based on correlation results (Table 2), plant height increases significantly as the average internode length increases. It also has a weak correlation with increased number of leaves, and internodes. Thus, the above-ground internode length contributes largely to taller plants while the numbers of leaves and internodes have a lesser influence. Interestingly, four of six QTL detected for plant height had the same chromosomal regions as the QTL for average internode length, sharing common molecular markers, such as $q P H 1 b$ versus $q A I L 1 ; q P H 3$ versus $q A I L 3 b$; $q P H 5 a$ versus $q A I L 5 a$, and $q P H 15 b$ versus $q A I L 5 b$. In contrast, only one QTL detected for plant height was located on the same chromosomal regions as the QTL for internode number. These results suggest that average internode length was the main contributor to plant height
genetically. Therefore, in order to understand the molecular mechanisms contributing to plant height, average internode length should be the primary focus of further investigations.

To date, a number of qualitative mutant genes controlling plant height have been mapped (Coe Jr 1980; Touzet et al. 1995; Helentjaris et al. 1993; Rhoades and Dempsey 1954; Robertson 1974), and a few of these genes were cloned recently, such as D8 (Ikeda et al. 2001) and D3 (Winkler and Helentjaris 1995). In a previous study (Beavis et al. 1991), QTL detected for plant height were compared in four populations, and nine QTL were identified in the chromosomal regions known to have qualitative genetic loci ( $D 8, D 3$, na2, gll7, $y d 2$, etc.) affecting plant height. In this study, four QTL detected for average internode length reside in the same chromosomal regions as five qualitative loci for plant height, such as $q A I L 1$ versus $D 8, q A I L 9$ versus $D 3, q A I L 5 b$ versus na2 and gl17, qAIL3b versus $y d 2$. The
plant height QTL, $q P H 1 b, q P H 3$, and $q P H 5 b$, share the same molecular markers as $D 8, y d 2$, $n a 2$, and gll7. Robertson (1985) pointed out that minor allelic variants at qualitative loci could be responsible for quantitative effects. In support of this suggestion, the results of this study imply that the QTL for average internode length may be related to qualitative genes controlling plant height. Qualitative genes may influence plant height by controlling average internode length. Hence, qualitative genes, which are located in the same chromosomal regions as QTL for average internode length, should be considered candidate genes. By the approach of associated analysis, quantitative trait genes for plant height may be isolated in the future.

Acknowledgments This work was supported by the State Key Basic Research and Development Plan of China and High Technology Project of China. The authors thank two anonymous reviewers for their valuable comments and suggestions.

## References

Abler BSB, Edwards MD, Stuber CW (1991) Isoenzymatic identification of quantitative trait loci in crosses of elite maize inbreds. Crop Sci 31:267-274
Ajmone-Marsan P, Monfredini G, Ludwig WF, Melchinger AE, Franceschini P, Pagnotto G, Motto M (1994) Identification of genomic affecting plant height and their relationship with grain yield and elite maize cross. Maydica 39:133-139
Austin DF, Lee M, Veldboom LR (2001) Genetic mapping in maize with hybrid progeny across testers and generations: plant height and flowering. Theor Appl Genet 102:163-176
Beavis WD, Grant D, Albertsen M, Fincher R (1991) Quantitative trait loci for plant height in four maize populations and their associations with qualitative genetic loci. Theor Appl Genet 83:141-145
Berke T, Rocheford T (1995) Quantitative trait loci for flowering, plant and ear height, and kernel traits in maize. Crop Sci 35:1542-1549
Coe E Jr (1980) Dominant dwarf D8 is between bz2 and gs on chromosome 1. MNL 54(26):27
Helentjaris T, Torres-Jerez I, McCreery T (1993) The use of bulk segregant analysis to map dwarf mutants. MNL 67(174):109
Ikeda A, Ueguchi-Tanaka M, Sonoda Y, Kitano H, Koshioka M, Futsuhara Y, Matsuoka M, Yamaguchi J (2001) Slender rice, a constitutive gibberellin response mutant, is caused by a null mutation of the

SLR1 gene, an ortholog of the height-regulating gene GAI/RGA/RHT/D8. Plant Cell 13:999-1010
Knapp SJ, Stroup WW, Ross WM (1985) Exact confidence intervals for heritability on a progeny mean basis. Crop Sci 25:192-194
Koester RP, Sisco PH, Stuber CW (1993) Identification of quantitative trait loci controlling days to flowering and plant height in two near isogenic lines of maize. Crop Sci 33:1209-1216
Kozumplik V, Pejic I, Senior L, Pavlina R, Gra ham G, Stuber CW (1996) Use of molecular markers for QTL detection in segregating maize populations derived from exotic germplasm. Maydica 41:211-217
Lance R, Lee M (1996) Genetic mapping of quantitative trait loci in maize in stress and nonstress environments:II. Plant height and flowering. Crop Sci 36:1320-1327
Lander ES, Green P, Abrahamson J, Barlow A, Daly MJ, Lincoln SE, Etoh T (1987) MAPMAKER: an interactive computer package for constructing primary genetic linkage maps of experimental and natural populations. Genomics 1:174-181
Lübberstedt T, Melchinger AE, Schǒn CC, Utz HF, Klein D (1997) QTL mapping in testcrosses of European flint lines of maize I. Comparison of different testers for forage yield traits. Crop Sci 37:921-931
Melchinger AE, Utz HF, Schǒn CC (1998) Quantitative trait locus (QTL) mapping using different testers and independent population samples in maize reveals low power of QTL detection and large bias in estimates of QTL effects. Genetics 149:383-403
Rhoades MM, Dempsey E (1954) Further studies on crossing over in inversion 3a. MNL 28(55):56-58
Robertson DS (1974) Linkage data for yellow dwarf 2 on chromosome 3. MNL 48(41):70-72
Robertson DS (1985) A possible technique for isolating genomic DNA for quantitative traits in plants. J Thero Bio 117:1-10
Schǒn CC, Lee M, Melchinger AE, Guthrie WD, Woodman WL (1993) Mapping and characterization of quantitative trait loci affecting resistance against second-generation European corn borer in maize with the aid of RFLPs. Heredity 70:648-659
Schǒn CC, Melchinger AE, Boppenmaizer J, BrunklausJung E, Herrmann RG, Seitzer JF (1994) RFLP mapping in maize: quantitative trait loci affecting testcross ferformance of elite European flint lines. Crop Sci 34:378-389
Touzet P, Winkler RG, Helentjaris T (1995) Combined genetic and physiological analysis of a locus contributing to quantitative variation. Theor Appl Genet 91(2):200-205
Veldboom LR, Lee M (1994) Molecular-marker-facilitated studies of morphological traits in maize. II: Determination of QTLs for grain yield and yield components. Theor Appl Genet 89:451-458
Veldboom LR, Lee M (1996) Genetic mapping of quantitative trait loci in maize in stress and nonstress environments: II. Plant height and flowering. Crop Sci 36:1320-1327

Wang S, Basten CJ, Zeng ZB (1994) Windows QTL Cartographer 2.0. Department of Statistics, North Carolina State University, Raleigh, NC
Winkler RG, Helentjaris T (1995) The maize dwarf3 gene encodes a cytochrome p 450-mediated early step in gibberellin biosynthesis. Plant Cell 7:1307-1317

Yan JB, Tang H, Huang YQ, Zheng YL, Li JS (2003) Dynamic QTL analysis for plant height in different developing stages in maize. Chin Sci Bull 48:1959-1964
Zeng ZB (1994) Precision mapping of quantitative trait loci. Genetics 136:1457-1468


[^0]:    T. Ji-hua $\cdot$ T. Wen-tao $\cdot$ Y. Jian-bing
    M. Xi-qing • M. Yi-jiang • D. Jin-rui -
    L. Jian-Sheng ( $\boxtimes$ )

    National Maize Improvement Center of China, Key Lab of Crop Genomics and Genetic Improvement, China Agricultural University, Yuanmingyuan West Road, Haidian, Beijing 100094, China
    e-mail: lijiansheng@cau.edu.cn
    T. Ji-hua

    College of Agronomy, Henan Agricultural University, Zhengzhou 450002, China

[^1]:    $L N$ leaf numbers, $I N$ internode numbers, $A I L$ average internode length, $P H$ plant height
    ${ }^{\text {a }} \mathrm{P}_{1}$ represents one parent, Z 3 , and $\mathrm{P}_{2}$ represents another parent, 871
    ${ }^{\mathrm{b}} \sigma_{\mathrm{g}}^{2}$, the genotypic variance; $\sigma_{\mathrm{g} \times 1}^{2}$, the genotype and location interaction variance; $\sigma_{\mathrm{e}}^{2}$, the environments variance
    ${ }^{c}$ The broad-sense heritability of the four measured traits
    ${ }^{\text {d }}$ The confidence intervals of broad-sense heritability between 5 and $95 \%$ significant levels

[^2]:    ${ }^{\text {a }}$ QTL for plant height and their chromosomal locations detected in this study
    ${ }^{\mathrm{b}}$ QTL for plant height and their flanking markers identified in previous reports, which has the same chromosomal regions with detected QTL in this paper

