## Factors regulating productivity in chemoautotrophic symbioses;

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#### Abstract

Symbioses involving sulfide-oxidizing bacteria and various metazoan phyla dominate megafaunal assemblages at cold seeps and hydrothermal v worldwide. The predominant species found living at cold seeps in Montrey Bay are the vesicomyid clams*Calyptogena kilmeriand C pacifica*. The growth and survival of these clams depend directly upon the productivity of their chemoautorphic endosymbionts, which is fueled by the oxidation of sulfide. For this reason, sulfide (energy) availability and sulfide physiology are thought to constrain symbiont and host production. Additional factors, however, are potentially equal in importance. Here we describe research concerning the productivity of two common clam species in relation to environmentally or physiologically limited processes. Although both species inhabit sulfide-rich sediments and depend nutritionally on their symbionts, many aspects of their life styles differ considerably. Our results indicate that C pacifica is physiologically oised for the uptake of sulfide, as measured by increased sulfide consumption ates, sulfide binding ability, and internal sulfide levels, as well as energy urnover, as measured by sulfide oxidation potential, sulfur metabolism enzymes and bacterial densities. In addition, C pacifica demonstrates higher rates of oxygen consumption and aerobic metabolism, than *C. kilmeri*. Growth rates of *C pacifica* (3% y<sup>1</sup>), however, are considerably slower than *C. kilmeri* (15% y<sup>1</sup>). This is surprising given that C pacifica possesses a seemingly greater potential for processing sulfide. These results contradict the idea that sulfide limits the productivity of these two systems and, for this reason, we believe they are onstrained by factors other than energy limitation.

#### Introduction

In the late 1970's scientists discovered novel deen-sea ecosystems based on chemosynthetic processes, fueled primarily by hydrogen sulfide, rather than photosynthetic carbon input (Corliss et al. 1979). Despite the poisonous nature of sulfide, numerous metazoans are known to inhabit areas of high sulfide. One such sulfide-rich ecosystem, teeming with chemosynthetically supported life, is Monterey Bay, California, USA.



Invertebrate communities living in association with cold seeps in Monterey Canvon were discovered in the 1980's. High faunal biomass in these areas is supported almost entirely by bacterialchemosynthesis, made possible by high concentrations of hydrogen sulfide, available for bacterial conversion into energy. In fact, many seep animals exist in symbiotic relationships with carbon-fixing, sulfide-oxidizing bacteria, relying upon these symbionts for nutrition (review Childress and Fisher 1992) The two predominant clams living in such symbioses at the Monterey seeps are *Calyptogena kilmeri* and *C. pacifica* (Barry *et al.* 1996).

#### Anatomy

Invertebrate-bacterial symbioses are extremely common among deep-sea environments of high sulfide. The host presumably benefits from nutritional integration with thesymbionts which overcomes the problem of food limitation in the deep sea, whereas the symbionts benefit from an environment protected from fluctuations of necessary metabolites. These arrangements have proven extremely successful as estimates of biomass and growth rates of some phyla have surpassed those of most animal life on the planet



# with emphasis on Calyptogena kilmeri and Calyptogena pacifica.

### Background

Calyptogena kilmeri and C pacifica are two widely distributed vesicomyidelams found at cold seeps in Monterey Bay. They serve as excellent model systems to explore factors limiting productivity in symbiotic

Both of these species inhabit sulfide-rich seep sediments and depend nutritionally on their symbionts, however, many aspects of their life styles differ considerably, suggesting differing strategies to thrive in seep envir



Invertebrate hosts and microbes form complex symbiotic associations involving tightly coupled physiological and biochemical processes. Figure 1 illustrates a few of the intermediate steps involving symbiont and host metabolism: metabolite uptake (1-3), bacterial assimilati transfer to host (4-7). The organic carbon eventually incorporated by the host, via host metabolism (8-9), is allocated to maintenance (10), growth (11), and reproduction.



Ve recently examined physiological processes comparing C, pacifica to C, kilmeri Our results indicate that there are notable differences between these two species.

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Para	meter	C. pacific	a		C. kilmer			sig
Uptal	ke of metabolites							
1	O 2 Uptake (mmoles / g / h)	14.0	-	5	3.0		5	*
	Hemoglobin (mM)	$1.3_{\pm} 0.1$		14	$1.8 \pm 0.1$		21	*
2	H S Uptake and Transport	See Table 3			See Table 3			
3	CO2 Uptake ( moles / g / h)	10.0		5	6.0		5	*
	CO <sub>2</sub> Internally (mM)	4.7± 2.3		16	3.3 ± 0.8		43	^
Bacte	rial metabolism							
4	Bacterial density (x 10 <sup>4</sup> g / g animal)	6.6± 0.7		8	3.9 ± 0.3		5	*
5	Sulfide consumption	See Table 3			See Table 3			
6	H S Oxidation (turnover by symbionts)	See Table 3			See Table 3			
7	Elemental sulfur (S) level (% of gill)	1.3± 2.1		7	4.1 ± 2.4		11	*
	Polysulfide (S <sub>a</sub> <sup>2</sup> ) level (% of gill)	$0.3_{\pm} 0.1$		6	0.6 ± 0.1		4	*
Host	metabolism							
8	Aerobic (Citrate synthase [LU.] g1 indiv)	1.0± 0.5		8	0.3 ± 0.2		7	
9	Anaerobic (Dehydrogenases [LU.] g1 indiv)	4.9 <sub>±</sub> 1.0		6	$7.1 \pm 0.8$		5	^
	% alanopine dehydrogenase	68.8 ± 1.6			29.6 ± 4.2			*
	% octopine dehydrogenase	10.6 ± 2.3			55.7 ± 5.3	-		*
Main	tenance							
10	pH regulation (H <sup>+</sup> -ATPases [%])	66.2 ± 0.6	-	2	$29.3 \pm 1.1$		3	^
Host	production							
11	Growth rate	See Table 4			See Table 4			

 C. pacifica demonstrates higher CO - and O - uptake rates, higher internal CO- levels, more symbiontspe gram of biomsk, higher levels elemental sulfur, a higher aerobic potential, as measured by CS activity in both gill and adductor muscle, and a larger percentage of ATPase activity to the elimination of protons.

C. kilmeri demonstrates a larger amount of polysulfides in the gill tissues, a higher anaerobic potential, and a higher percentage of dehydrogenase activity comprised of ODH activity.

We investigated sulfide-related processes in C. kilmeri and C. pacifica (noted in pink above) in order to determine if sulfide-limitation, at any step during sulfide movement, can act to constrain productivity in symbiotic systems. References Acknowledgements Corlisset al. 1979 Science203, 1073-1083. Patrick Whaling Bethany Schaarschmidt Kurt Buck nnette Goar Annette Goan Dr. Ed DeLong Victoria Orphan Dr. Christina Pres Peter Girguis



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In order for chemoautotrophic symbioses to thrive, symbionts must have access to an electron donor used as an energy source (sulfide), an electron acceptor (oxygen or nitrate), and inorganic carbon (CO<sub>2</sub> or HCO<sub>2</sub>). These associations depend upon metabolism of the endosymbionts, which is fueled by the oxidation of sulfide, and, therefore, dependent upon sufficient sulfide flux from sea floor seepage or venting.

For this reason, the supply of sulfide and sulfide physiology are usually considered the most influential parameters limiting production by symbiont and host.



ne productivity of a symbiotic system is uenced primarily by the ability to proce sulfide (energy), than growth rates of the host (a component of host production) should correlate component of host production) snown contents positively with sulfide uptake rates, the ability to transport sulfide, and ability of the symbio to extract energy from sulfide (Table 2).

To support bacterial production of organic carbon using sulfide as an energy source:

the host must achieve sufficient rates of sulfide uptake and internal sulfide levels via extremely effective sulfide binding mechanisms (Table 3).

the endosymbiont populations present in these hosts must harness energy from the oxidation of sulfur compounds, presumably via effective enzymatic pathways (Table 3).

the host uses organic carbon transferred from the symbiont to fuel host processes, such as growth and reproduction (Table 4)

Table 3 describes our measurements of parameters involved in acquiring sulfide energy, including sulfide uptake and binding, sulfide availability to symbionts, and other sulfide-related physiology.

#### Table 3: Sulfide-related parameters observed in C. pacifica and C. kilmeri-

Rate		C. pacifica		C. kilmeri		\$						
H2SUptake (whole animal -mmoles / g animal / h)		50 ·	5	2-3 -	5							
H2SConsumption (gill - mnoles / g animal / h)		11 -	4	2 -	5							
H2S levels internally ( mM)		2.9 ± 2.0 -	16	1.1 ± 0.8 <sup>-</sup>	43							
H2S Binding (above ambient levels)		10-60x -	10	5-10x -	10							
Zinc (mM)	$Zn + H_2S = ZnS$	6.3±0.8 -	14	4.7 ± 0.3 -	36							
Blood volume (% of total biomass)		36.8 ± 6.0 -	10	32.5 ± 4.6 -	10							
H2S Oxidation (SOxidase [I.U.])	$H_2S(orHS) + O_2 = SO_3^2$	6.7 ± 2.1 ·	6	4.2 ± 1.0 -	8							
APS reductase ([LU.])	$SO_3^2 + AMP = APS + 2e$	41.0 ± 4.6 -	5	$28.2 \pm 1.0$ <sup>-</sup>	6							
Up arrows = higher rates, Down arrows = lower rates (* indicates significant difference >95%, * indicates significant difference >90%)												
<ul> <li>C. pacifica demonstrated higher rates of who under physiologically relevant conditions that</li> </ul>	ole animal H <sub>2</sub> S uptake as we in <i>C. kilmeri</i>	ell as sulfide co	nsun	aption by intact	gill	ŝ						
•C. pacifica demonstrated increased H <sub>2</sub> S levels internally, resulting from increased sulfide binding ability.												

increased sulfide binding ability in C. pacifica is presumably due to higher concentrations of zinc (which binds ulfide 1:1) and a larger % blood (as related to total body mass ), than C kilmeri

trated significantly higher sulfid oxidase and APS reductase activity than C. kilmeri. Sulfide . pacifica de xidase is involved in the oxidation of sulfide to sulfite (pathway providing the most reducing power), and APS eductase catalyzes the reactions between AMP and sulfite.

Table 4 describes our measurements of productivity-related parameters.

Production	C. pa	cifica		C. ki	lmeri		sig
Reproduction (gonad – as % of total weight)	6.2 ± 0.4		12	$5.5 \pm 0.2$		13	ns
Growth rate (% yr <sup>4</sup> )	3	-	50	15		50	
differences in gonad biomass (both = 5-69	ing one	of totalb	ioma	(e)	SIVE EVIC	ience	101
differences in gonad biomass (both ~ 5-69	6 as a %	of totalb	ioma	is).	SIVE EVIC	ience	101
<ul> <li>Mark and recenture studies have provides</li> </ul>	6 as a %	of totalb	ioma:	is).	5% v <sup>1</sup> fc	rC k	ilmer

These two symbiotic systems do not appear to be sulfide limited. C. kilmeri lives in higher sulfide levels and C. pacifica has a greater ability to take up sulfide. Data show that internal sulfide levels are sufficient for bacterial productivity in both species and that growth rates of the hosts do not correlate with sulfide-related physiology.

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regulation constrains productivity in C. pacifica.