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Activation of NF-E2-related factor-2 reverses biochemical dysfunction of endothelial cells induced by hyperglycemia linked to vascular disease

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ABSTRACT

OBJECTIVE – Sulforaphane (SFN) is an activator of transcription factor NF-E2-related factor-2 (nrf2) that regulates gene expression through the promoter antioxidant response element (ARE). Nrf2 regulates the transcription of a battery of protective and metabolic enzymes. The aim of this study was to assess if activation of nrf2 by SFN in human microvascular endothelial cells prevents metabolic dysfunction in hyperglycemia.

RESEARCH DESIGN AND METHODS – Human microvascular HMEC-1 endothelial cells were incubated in low and high glucose concentrations (5 and 30 mM) and activation of nrf2 assessed by nuclear translocation. The effect of SFN on multiple pathways of biochemical dysfunction, increased reactive oxygen species (ROS) formation, hexosamine pathway, protein kinase C pathway and increased formation of methylglyoxal, was assessed.

RESULTS – Activation of nrf2 by SFN induced nuclear translocation of nrf2 and increased ARE-linked gene expression. For example, 3 - 5 fold increased expression of transketolase and glutathione reductase. Hyperglycemia increased the formation of ROS – an effect linked to mitochondrial dysfunction and prevented by SFN. ROS formation was increased further by knockdown of nrf2 and transketolase expression. This also abolished the counteracting effect of SFN, suggesting mediation by nrf2 and related increase of transketolase expression. SFN also prevented hyperglycemia-induced activation of the hexosamine and protein kinase C pathways, and prevented increased cellular accumulation and excretion of the glycating agent, methylglyoxal.

CONCLUSION – We conclude that activation of nrf2 may prevent biochemical dysfunction and related functional responses of endothelial cells induced by hyperglycemia in which increased expression of transketolase has a pivotal role.

Abbreviations: AGE, advanced glycation endproduct; AKRd, aldo-keto reductase; AR, aldose reductase; ARE, antioxidant response element; DAG, diacylglycerol; GCL, γ-glutamylcysteine ligase; CK2, casein kinase 2; GSHRd, glutathione reductase; Fyn, 59-kDa src family-related protein tyrosine kinase; GFP, green fluorescent protein; GST, glutathione transferase; H_2DCFDA, 2',7'-dichlorodihydro-fluorescein diacetate; IGT, impaired glucose tolerance; Keap-1, Kelch-like ECH-associated protein 1; NQO1, quinone reductase-1; nrf2, NF-E2-related factor-2; NQO1, quinone reductase; PARP, poly(ADP-ribose)polymerase; PKC, protein kinase C; ROS, reactive oxygen species; SFN, sulforaphane; TA, transaldolase; TK, transketolase.

There is an increased risk of vascular disease in diabetes that is a major cause of patient morbidity and mortality. This gives rise to a characteristic spectrum of diabetic microvascular disease - retinopathy, nephropathy and neuropathy, and macrovascular disease – heart disease and stroke (1-4). Vascular disease in diabetes is associated with dysfunction of endothelial cells in hyperglycemia. Activation of multiple pathways of biochemical dysfunction induced in vascular endothelial cells by high glucose concentration is thought to underlie the link of hyperglycemia in diabetes to the development of vascular disease (5;6). A common feature of endothelial cell dysfunction in hyperglycemia is increased formation of reactive oxygen species (ROS) by mitochondria, oxidative stress with inactivation of glyceraldehyde-3-phosphate dehydrogenase, and accumulation of triosephosphates and fructose-6-phosphate (7-9). There is an associated activation of protein kinase C (PKC), hexosamine pathway O-linked protein glycosylation and increased glycation by methylglyoxal and other dicarbonyls forming advanced glycation endproducts (AGEs) (10-12) . This appears to be driven mainly by the accumulation of glycolytic intermediates. Recent research has indicated that activation of the reduced pentosephosphate pathway by high dose thiamine and related prodrug Benfotiamine may counter this metabolic dysfunction (9;13;14) but little is known of the endogenous coordinated stress response to decrease triosephosphate accumulation and its link to increased ROS formation and oxidative stress in hyperglycemia.

NF-E2-related factor-2 (nrf2) is a member of the cap 'n' collar subfamily of bZIP transcription factors. It is an essential transactivator of genes containing an antioxidant response element (ARE) in their promoter – reviewed in (15;16). ARE-linked genes include a battery of protective and metabolic enzymes: γ-glutamylcysteine ligase (GCL), glutathione reductase (GSHRd), aldo-keto reductase (AKRd), glutathione transferases (GSTs), quinone reductase (NQO1), nrf2 itself (17), and others (18). Nrf2-linked gene expression has a key role in the protection of cells against oxidative stress, carbonyl compounds and electrophilic agents. Interestingly, the thiamine-dependent enzyme transketolase (TK) and transaldolase are also ARE-linked genes (18). TK is considered to be the rate-controlling enzyme in the pentosephosphate pathway.

Under basal conditions, nrf2 is complexed with Kelch-like ECH-associated protein 1 (Keap1) - a BTB-Kelch protein. Keap1 is a substrate adaptor protein for a Cul 3-dependent E3 ubiquitin ligase complex, directing nrf2 for proteasomal degradation (19). Oxidative stress, electrophiles and SFN-like inducers disrupt the Keap1-nrf2 complex: nrf2 translocates to the nucleus and, combining with small maf protein (20), induces ARE-linked gene expression (21). SFN releases nrf2 from Keap1 by modification of critical cysteine thiol residues (22). Keap1 has concurrent increased susceptibility to degradation but also has ARE-linked gene expression and may be induced by nrf2 activation providing an auto-regulatory feedback loop (23). Nrf2 also undergoes nuclear export, establishing cytoplasmic/nuclear dynamic shuttling (24). Recent research has suggested the serine/threonine kinase CK2 has a role in nuclear import of nrf2 and both CK2 and tyrosine kinase Fyn influence nuclear export and degradation of nrf2 (25;26) – Fig. 1a. There is an active nrf2-Keap-1 system in vascular endothelial cells (27).

The role, if any, of nrf2-linked gene expression in countering endothelial

dysfunction in hyperglycemia has not been disclosed. Disposal of glyceraldehyde-3-phosphate (GA3P) and fructose-6-phosphate (F-6-P) by the reductive pentosephosphate pathway induced by activation of nrf2 and increased expression of TK suggested a possible mechanism of intervention. In this report, we show that activation of nrf2 by the dietary activator SFN – limited to concentration ranges found in plasma after consumption of broccoli (28) for relevance to future clinical dietary intervention - increased the expression of protective enzymes under ARE-linked transcriptional control and prevented metabolic dysfunction in endothelial cells induced by hyperglycemia where increased expression of TK has a critical role.

RESEARCH DESIGN AND METHODS

Reagents. MCDB-131 medium, fetal calf serum, Alexa Fluor 488 rabbit anti–mouse IgG and 2',7'-dichlorodihydrofluorescein diacetate (H_2DCFDA) were purchased from Invitrogen (Paisley, U.K.). Monoclonal (mouse) anti–O-linked N-acetylglucosamine (Clone RL2) was purchased from Abcam Plc (Cambridge, U.K.). All other chemicals were obtained from Sigma-Aldrich (Poole, Dorset, UK), unless otherwise stated.

Cell culture. Human microvascular HMEC-1 endothelial cells were cultured as described (29). HMEC-1 cells (4 - 6 x 10^6) were incubated in MCDB-131 medium with 10% serum, 10 mM L-glutamate, 10 ng/ml epidermal growth factor and 1 μg/ml hydrocortisone in 92 mm diameter Petri dishes with low glucose (5 mM) and high glucose (30 mM) in the absence and presence of 4 μM SFN for 6 - 48 h, trypsinised and analysed as described below.

siRNA and vector transfection. HMEC-1 endothelial cells were incubated in MCDB-131 medium with 10% serum and then transfected with antisense ODN for nrf2 (SI00657030; Qiagen) or TK (SI02653791) using the HiPerfect transfection reagent, according to the manufacturers instructions. The incubation was then continued with normal medium for 48 h. Cultures were then continued with low and high glucose concentrations in the absence and presence of 4 μM SFN for 24 h and/or other additives. Knockdown was confirmed by RT-PCR and real time RT-PCR for nrf2 and TK. RNA was extracted using the RNeasy minikit (Qiagen), re-transcribed by SuperScript III First Strand Synthesis system (Invitrogen) and quantified by real time RT-PCR using the Taqman MGB probes, design and supplied by API. Knockdown was 60% for TK and >90% for nrf2.

Nuclear translocation of nrf2 by immunoblotting Nuclear and cytosolic proteins were isolated from HMEC-1 cells incubated with and without SFN using CelLytic NuCLEAR extraction kit (Sigma). Trypsinised cells were washed in phosphate-buffered saline at 4 ^oC and the cell pellet lysed with hypotonic lysate buffer containing dithiothreitol, a protease inhibitor cocktail and IGEPAL CA-630 on ice for 15 min. The lysate was centrifuged (11,000g, 30 s, 4 ^oC) and the supernatant used for the cytosolic extract. The pellet (crude nuclear fraction) was treated with extraction buffer containing dithiothreitol and protease inhibitor cocktail at 4^oC for 30 min, vortex mixed and centrifuged (21,000g, 4^oC, 5 min). The supernatant was collected for the nuclear extract. All protein extracts were frozen at -80 ^oC immediately until further analysis. The protein concentration was determined with EZQ Protein Quantitation Kit (Invitrogen).

Proteins in the cytosolic and nuclear fractions were separated by 10% SDS-polyacrylamide gel electrophoresis, transferred electrophoretically onto polyvinylidene difluoride membrane and the membrane blocked with 5% non-fat milk in

tris-buffered saline-Tween buffer (10 mM Tris-HCl, pH 7.5, 150 mM NaCl, 0.05% Tween-20). The membrane was probed with anti-nrf2 antibody (H-300; 1:1500 dilution) overnight at 4^{o}C. After washing, the membrane was incubated with horseradish peroxidase-conjugate second antibody (Sigma, 1:3000 diluted) for 1 h at room temperature and immunoconjugate detected with enhanced chemiluminescence. The membrane was then incubated with stripping buffer (100 mM β-mercaptoethanol, 2% SDS, 62.5 mM Tris/HCl pH 6.8), blocked with 5% non-fat milk in tris-buffered saline-Tween buffer, and re-probed with antibodies to reference proteins, β-actin and lamin A. Protein band intensities were quantified using ImageQuant TL software (Amersham Biosciences).

Characterization of biochemical dysfunction. The intracellular formation of ROS was detected using the fluorogenic probe H_2DCFDA. Cells (2 x 10^6) were incubated with and without SFN and mitochondrial inhibitors for 24 h, washed with phosphate-buffered saline and then incubated further with H_2DCFDA (20 μM) for 45 min, washed again and analysed by flow cytometry. Shorter incubations of 1 h pre-incubation in hyperglycemia with and without mitochondrial inhibitors were also performed by microplate fluorescence measurements, normalizing the fluorescence intensity to cell number. The effect of treatments on cell viability was assessed by Trypan blue exclusion.

Hexosamine pathway activity was assessed by a quantitative dot Western blot assay for O-linked N-acetylglucosamine modified protein. Cytosolic protein extracts (0.7 μg in 5 μl phosphate buffered saline) were immunoblotted with RL2 antibody (1:800 dilution in blocking buffer) for 3 h at room temperature, blocked with 10% BSA and washed (30). The immunocomplexes were detected with Alexa Fluor 488 rabbit anti–mouse IgG (1:700 dilution) and quantified by microplate fluorimetry. The concentrations of methylglyoxal in HMEC-1 cells and medium of cultures with and without SFN were determined by derivatisation with 1,2-diaminobenzene and quantitation by liquid chromatography with tandem mass spectrometric detection and stable isotopic dilution analysis (12). PKC activity was assayed in membrane and particulate fractions of HMEC-1 cells with exogenous diacylglycerol (1,2-dioleoyl-sn-glycerol) activator and epidermal growth factor receptor peptide fragment VRKRTLRRL as substrate (9).

Statistical analyses. All statistical analyses were performed using paired Student's t test, and results are expressed as mean ± SD. A P value < 0.05 was considered to be significant.

RESULTS

Activation of nrf2 and ARE-linked gene expression in endothelial cells by the dietary activator sulforaphane. We investigated the activation status of nrf2 in human microvascular endothelial cells by assessing nuclear translocation of human nrf2 by immunoblotting in cytosolic and nuclear fractions and confocal microscopy of nrf2-GFP fusion protein**.** HMEC-1 endothelial cells incubated in model hyperglycemia (30 mM glucose) showed no significant nuclear translocation of nrf2 with respect to normoglycemic control (5 mM glucose) after incubation for 6 h. Addition of SFN (4 μM) gave a 2-fold increase in nuclear nrf2 in both normoglycemic and hyperglycemic cultures. In the normoglycemic culture, the concentration of nrf2 in the cytosol was decreased concomitantly; whereas in the hyperglycemic culture, the concentration of nrf2 in the cytosol was increased. This suggests the double insult of hyperglycemia and SFN increased the cellular content of nrf2

protein (Fig. 2, a. and b.). This concentration of SFN did not induce significant cytotoxicity in HMEC-1 cells in incubations for up to 48 h - as assessed in previous studies (31).

Real time RT-PCR analysis of target ARE-linked gene expression revealed a marked 5-fold induction of TK mRNA in cells stimulated with SFN (Fig. 3a) and a lower, 3 - 4 fold increase in glutathione reductase (GSHRd) mRNA (Fig. 3 b). The cytosolic activity of TK was increased 40 - 60% by exposure to SFN (Fig. 3c). The increase in expression of TK induced by SFN in normoglycemic and hyperglycemic cultures was prevented by knockdown of nrf2 expression (Fig. 3a). To confirm that there is an ARE-linked induction of gene expression by SFN in HMEC-1 cells, we studied the mRNA levels of the typical nrf2-linked, ARE-mediated gene, quinone reductase-1 (NQO1). NQO1 expression, normalized to culture in normoglycemia, was increased approximately 2-fold in HMEC-1 cells incubated with 4 μM SFN in normoglycemic and hyperglycemic conditions. Basal and SFN-induced expression of NQO1 in hyperglycemic culture was decreased 56% and 31%, respectively, by knockdown of nrf2 with antisense ODN – Fig. 3d

Increased formation of reactive oxygen species by endothelial cells in hyperglycemia, reversal by sulforaphane and critical role of transketolase. To examine the effect of ARE-linked gene expression on biochemical dysfunction in hyperglycemia, the cellular production of ROS was quantified. Hyperglycemic culture of endothelial cells produced a 3-fold increased formation of ROS - Fig. 4a. This was not induced by addition of 25 mM L-glucose (which does not permeate into endothelial cells) to the normoglycemic control. Incubation of endothelial cells with SFN reversed the increase in ROS by 73% - suggesting activation of ARE-linked gene expression prevented increased ROS formation. Increased ROS formation by HMEC-1 cells in hyperglycemic cultures was prevented by incubation for 60 min with mitochondrial inhibitors prevented increased ROS formation. ROS formation (percentage of normoglycemic control): p-trifluoromethoxycarbonylcyanide phenylhydrazone (10 μM), 94 ± 3%; rotenone (5 μM), 104 ± 3%; and myxothiazole (2 μM), 88 ± 3%. This suggests that dysfunction of mitochondria was a primary source of the increased ROS and electron flux through complex I and III contributed to this effect. Incubation of HMEC-1 cells with these inhibitors for 24 h decreased cell viability by 40 – 50% and masked the effect of rotenone.

We next sought evidence if the prevention of increased formation of ROS was dependent on nrf2. We transfected endothelial cells with antisense ODN to knockdown expression of nrf2 - confirmed by PCR and real time RT-PCR (Fig. 4b). Decreasing the expression of nrf2 exacerbated ROS production in both normoglycemic and hyperglycemic cultures: ROS was increased 145% with 5 mM glucose and 190% with 30 mM glucose, with respect to cells with 5 and 30 mM glucose transfected with scrambled ODN. The increased formation of ROS with nrf2 knockdown was maintained and not reversed by SFN, consistent with the prevention of increased ROS by SFN being mediated by activation of nrf2 rather than a direct antioxidant effect of SFN. Knockdown of TK by transfection of endothelial cells with antisense ODN also exacerbated ROS production in both normoglycemic and hyperglycemic cultures: ROS was increased 247% and 131% respectively, with respect to cultures with 5 and 30 mM glucose transfected with scrambled ODN. The increased formation of ROS with antisense TK ODN knockdown was maintained and not reversed by SFN, consistent with the prevention of increased

ROS by SFN being mediated by increased expression of TK (Fig. 4b).

Multiple pathways of biochemical dysfunction in endothelial cells in hyperglycemia and reversal by sulforaphane. Reversal of increased mitochondrial ROS formation is expected to counter biochemical dysfunction in multiple pathways. Hyperglycemia increased the activity of PKC by 180% in the cytosolic fraction and 347% in the membrane fraction of HMEC-1 endothelial cells. These increases were reversed partially in both cytosolic and membrane fractions by SFN: the hyperglycemia-induced increase in PKC activity was reversed 40% in the cytosolic fraction and 67% in the membrane fraction (Fig. 5, a. and b.). Similarly, hexosamine pathway activated enzymatic O-linked glycosylation increased 2-fold in hyperglycemia and was reversed by SFN (Fig. 5c.). Finally, the concentrations of methylglyoxal in HMEC-1 cells and medium was increased 2-fold in hyperglycemia and reversed by SFN (Fig. 5d.).

DISCUSSION

The reversal of biochemical dysfunction of endothelial cells in hyperglycemia by SFN suggests that activation of nrf2 and related ARE-linked gene expression is a novel strategy to suppress endothelial cell dysfunction and possibly also the development of vascular disease in diabetes. For example, decreased cellular and extracellular secretion of methylglyoxal is expected to prevent dicarbonyl glycation of cellular and extracellular matrix proteins and prevent hyperglycemia-induced endothelial cell detachment and anoikis (12). Similar increased ARE-linked gene expression in human aortal endothelial cells engineered by adenovirus-mediated expression of Nrf2 protected human aortal endothelial cells from cytotoxicity induced by hydrogen peroxide, and tumour necrosis factor-α-induced increased expression of monocyte chemoattractant protein-1 and vascular adhesion molecule-1. This suggests that increased activation of nrf2 may also confer anti-atherogenic activity (32).

Activation of nrf2 by SFN produced nuclear accumulation of the 98 kDa band of nrf2 protein. The 98 kDa band has ARE binding activity and is produced by kinase CK2-catalysed phosphorylation. Nrf2 levels in cells are regulated by further phosphorylation, nuclear export and degradation – which may be enhanced by ARE-linked expression of Keap1 (23;26;33). Nrf2 may also exhibit ARE-linked expression. Quantitative Western blotting for nrf2 revealed increased nrf2 in both the nucleus and cytosol in hyperglycemia with SFN treatment, suggesting total cellular nrf2 protein had increased but only with co-stimulation from both SFN and hyperglycemia. The expression of nrf2 is ARE-regulated with nrf2 binding to its own promoter (17). Hyperglycemia may synergise with SFN to increase nrf2 via ARE-induced expression by increasing endogenous activators of nrf2 – such as 4-hydroxynonenal (34), increasing the activity of CK2 via Ca^{2+}/calmodulin dependent mechanisms (25;35), or other activatory mechanism. This may reflect an ARE-linked, anti-stress response to hyperglycemia in endothelial cells – albeit too weak to mount an adequate defence against the metabolic insult.

Although nrf2 activation leads to increased expression of enzymes linked to antioxidant functions countering oxidative stress, in this study increased expression of TK made a critical and pivotal intervention. The increased reductive pentosephosphate pathway activity thereby achieved counters the accumulation of triosephosphates and fructose-6-phosphate driving metabolic dysfunction (9). The protective role of the

reductive pentosephosphate pathway in ARE-linked gene expression has hitherto not been identified, although increased TK expression is associated with the dietary restriction model of healthy ageing (36). Enzymatic activity of TK in the pentosephosphate pathway produces indirectly the NADPH co-factor for other ARE-linked gene products GSHRd, AKRd, NQO1, and thioredoxin reductase that contribute to countering the effects of oxidative stress (37). Therefore, contributions of other ARE-linked gene expression in synergism with TK in reversal metabolic dysfunction in hyperglycemia cannot be excluded.

Multiple pathways of biochemical dysfunction in HMEC-1 cells induced by hyperglycemia were reversed by SFN except reversal of PKC activation was resistant to the response. This may be due to ARE-linked induced expression of 1-acylglycerol-3-phosphate O-acyltransferase 3 by SFN (38). This enzyme converts lysophosphatidic acid to phosphatidic acid – a precursor of the PKC activator diacylglycerol (DAG) in *de novo* synthesis stimulated in hyperglycemia. The increased expression of this ARE-linked gene may maintain increased levels of DAG although the increased concentrations of glycolytic intermediates upstream driving *de novo* synthesis of DAG are expect to have been corrected. Prolonged exposure to increased levels of DAG has been suggested as a stimulus of increased PKC expression in hyperglycemia associated with diabetes that may underlie the increased specific activity of PKC in both membrane and cytosolic cell fractions found herein. Alternatively, when we studied activation of transketolase by high dose thiamine supplements (7), we also found an incomplete reversal of hyperglycemia-induced increase in PKC activity. This may suggest that factors other than those influenced by transketolase contribute to PKC activation in hyperglycemia.

The prevention of nrf2 activation by SFN by addition of GSH may be interpreted as an effect of GSH acting as an antioxidant. However, SFN and other isothiocyanates bind reversibly to non-protein thiols such as GSH, decreasing the concentration of free SFN available to modify Keap1. The residual low concentration of free SFN is inadequate to produce a pharmacological response and undergoes slow inactivation by spontaneous hydrolysis (39). The inhibition of nrf2 activation by GSH may, therefore, also reflect interception of SFN in the extracellular medium and prevention of it reaching the cellular receptors such as Keap1.

SFN and related isothiocyanates are cytotoxic to endothelial cells and other cells at higher concentrations than used herein (typically 20 – 40 μM) (31;40). The cytotoxicity is mediated through interactions with death receptors and apoptotic signalling (41), also involving inhibition of p38 MAP kinase (42), MEKK1 (43) and protein phosphatase M3/6 (44), and activation of ERK1/2 and JNK (45). This is independent of the disruption of the Keap1-nrf2 complex. Such concentrations of SFN are higher than achieved by *Brassica* vegetable consumption (28) and related cell signalling is expected to be of limited relevance to dietary exposures of SFN.

There was increased formation of ROS by HMEC-1 cells in model hyperglycemia of 30 mM D-glucose. The lack of similar effect induced by L-glucose indicated glucose entry and metabolism into cells was required for increased ROS formation. Increased ROS formation was linked to mitochondrial dysfunction – consistent with previous reports (10), where complex I and complex III were involved – the former probably by reverse electron flow from complex II (46). Incubation of HMEC-1 cells with the mitochondrial inhibitors for 24 h produced significant cytotoxicity and

masked the role of complex I in hyperglycemia-induced ROS formation. Incubation with inhibitors for 1 h did not induce cytotoxicity. Similar effects may have compromised the outcome of previous studies of this type (47). Other sources of increased ROS formation in microvascular endothelial cells in hyperglycemia have been identified: activation of vascular NADPH oxidase, inactivation and uncoupling of endothelial nitric oxide synthase (48;49), related upstream signalling linked to poly(ADP-ribose)polymerase (PARP) (7), aldose reductase (AR) (50), and the xanthine/xanthine oxidase system (51). Mitochondrial metabolism appears to be a major site of ROS formation in the HMEC-1 cell culture model. There are inter-related mechanisms, however, that may explain how SFN-induced ARE-linked gene expression could prevent activation of other pathways. Activation of PARP may be prevented by SFN-induced increased expression of antioxidant enzymes, preventing oxidative damage to DNA. The activation of the polyol pathway may be prevented by SFN decreasing the cellular concentration of methylglyoxal and thereby preventing methylglyoxal-induced expression of AR (52).

These findings provide the biochemical basis for the link of a vegetable-rich diet with decreased endothelial dysfunction (53) – including that part of the Mediterranean diet (54), suggesting dietary exposure to nrf2 activators derived from cruciferous vegetables may have be involved. The functional importance of this finding requires further evaluation. It is expected that the protective effect of nrf2 activators may be stratified by severity of exposure to hyperglycemia, with increasing effects in the non-diabetic, impaired glucose tolerance and diabetic populations. Hyperglycemia alone did not induced ARE-linked gene expression in HMEC-1 endothelial cells - even when confronted with damaging insults of oxidative stress and accumulation of dicarbonyls. Physiological activators of nrf2 are thought to be the lipid peroxidation-derived aldehyde, 4-hydroxynonenal and J_3-isoprostanes (34;55). The metabolic insult of hyperglycemia is of non-lipidic origin. This may be why the nrf2 system does not respond strongly to it. There is now evidence that diabetes does induce a weak nrf2-mediated protective response – in the high fat diet mouse model of type 2 diabetes, fore example (56). The weak response suggests an effective protection to prevent vascular disease has not been mounted and, therefore, provides a clear future opportunity for pharmacological intervention. Cruciferous vegetable consumption and synthetic activators of nrf2 are expected to decrease the risk of vascular disease in diabetes.

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FIG. 1. Nrf2 activation and transketolase expression in human HMEC-1 endothelial cells *in vitro*. a. Schematic diagram showing activation of nrf2 and dynamic nuclear-cytoplasmic shuttling of nrf2 for expression of ARE-linked. b. Multiple pathways of biochemical dysfunction induced by hyperglycemia in microvascular endothelial cells and effect of nrf2 activated, ARE-mediated induction of transketolase expression. Other mechanisms of biochemical dysfunction may be involved.

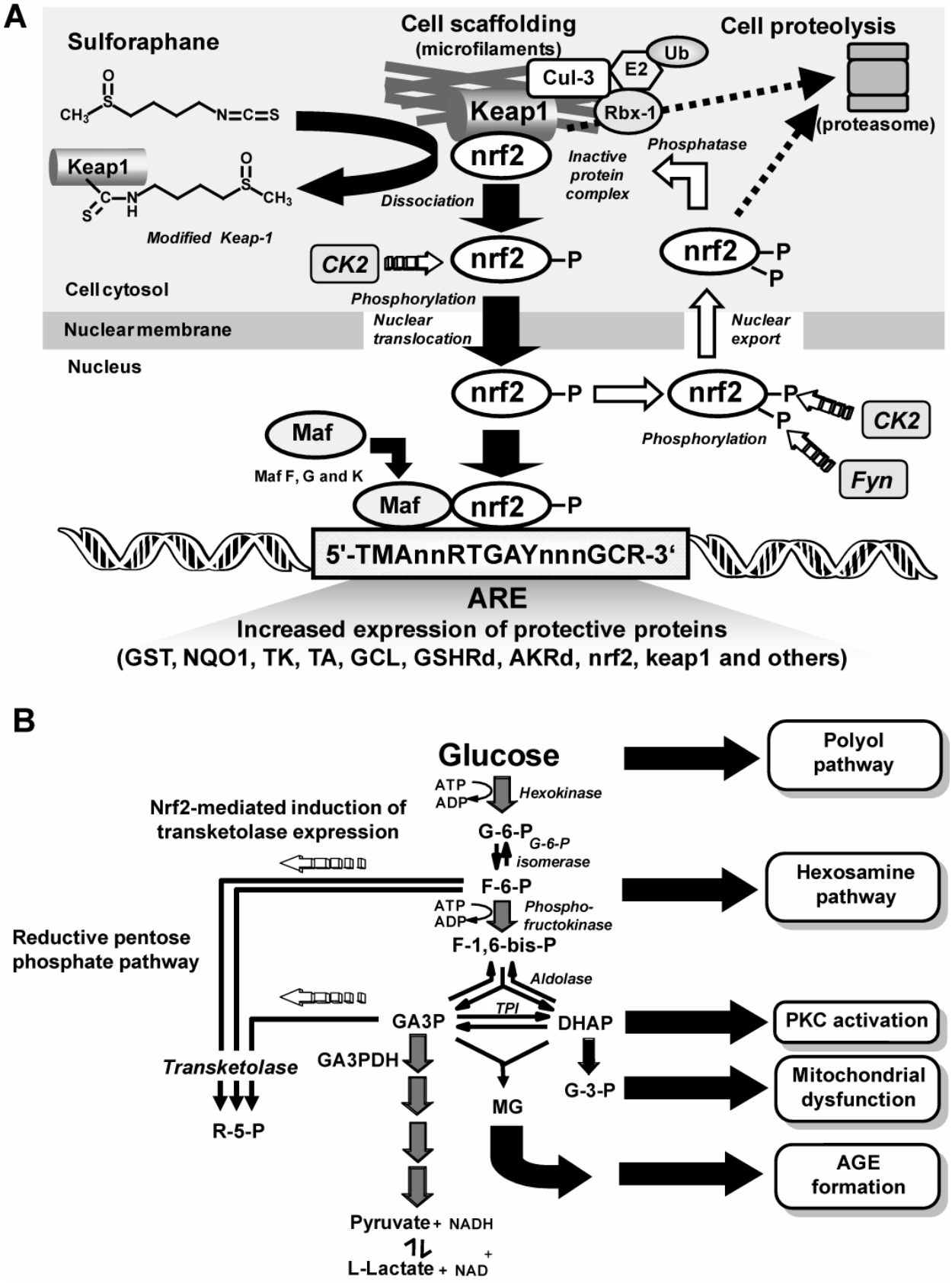
A
Sulforaphane
Cell scaffolding
(microfilaments)
Cell proteolysis
Cul-3
E2
Ub
Rbx-1
Keap1
(proteasome)
nrf2
Phosphatase
Inactive protein complex
Dissociation
Modified Keap-1
CK2
nrf2 P
nrf2 P P
Cell cytosol
Phosphorylation
Nuclear membrane
Nuclear translocation
Nuclear export
Nucleus
nrf2 P
nrf2 P P
CK2
Phosphorylation
Fyn
Maf
Maf F, G and K
Maf nrf2 P
5'-TMAnnRTGAYnnnGCR-3'
ARE
Increased expression of protective proteins
(GST, NQO1, TK, TA, GCL, GSHRd, AKRd, nrf2, keap1 and others)
B
Glucose
ATP
ADP
Hexokinase
Polyol pathway
Nrf2-mediated induction of transketolase expression
G-6-P
G-6-P isomerase
F-6-P
Hexosamine pathway
ATP
ADP
Phospho-fructokinase
Reductive pentose phosphate pathway
F-1,6-bis-P
Aldolase
TPI
GA3P
DHAP
PKC activation
Transketolase
GA3PDH
G-3-P
Mitochondrial dysfunction
R-5-P
MG
AGE formation
Pyruvate + NADH
L-Lactate + NAD+

FIG. 2. Nuclear translocation of nrf2 in human HMEC-1 endothelial cells *in vitro* activated by sulforaphane. Immunoblotting for nrf2 (98 kDa band): a. nuclear fraction, and b. cytosolic fraction. Densitometric intensity ratios are mean ± SD (n = 3). Key to incubations: N, 5 mM glucose; H, 30 mM glucose; N+SFN, 5 mM glucose + 4 μM SFN; and H+SFN, 30 mM glucose + 4 μM SFN. Significance: *, ** and *** indicate $P<0.05$, $P<0.01$ and $P<0.001$ with respect to N; o and oo, indicate $P<0.05$ and $P<0.01$ with respect to H.

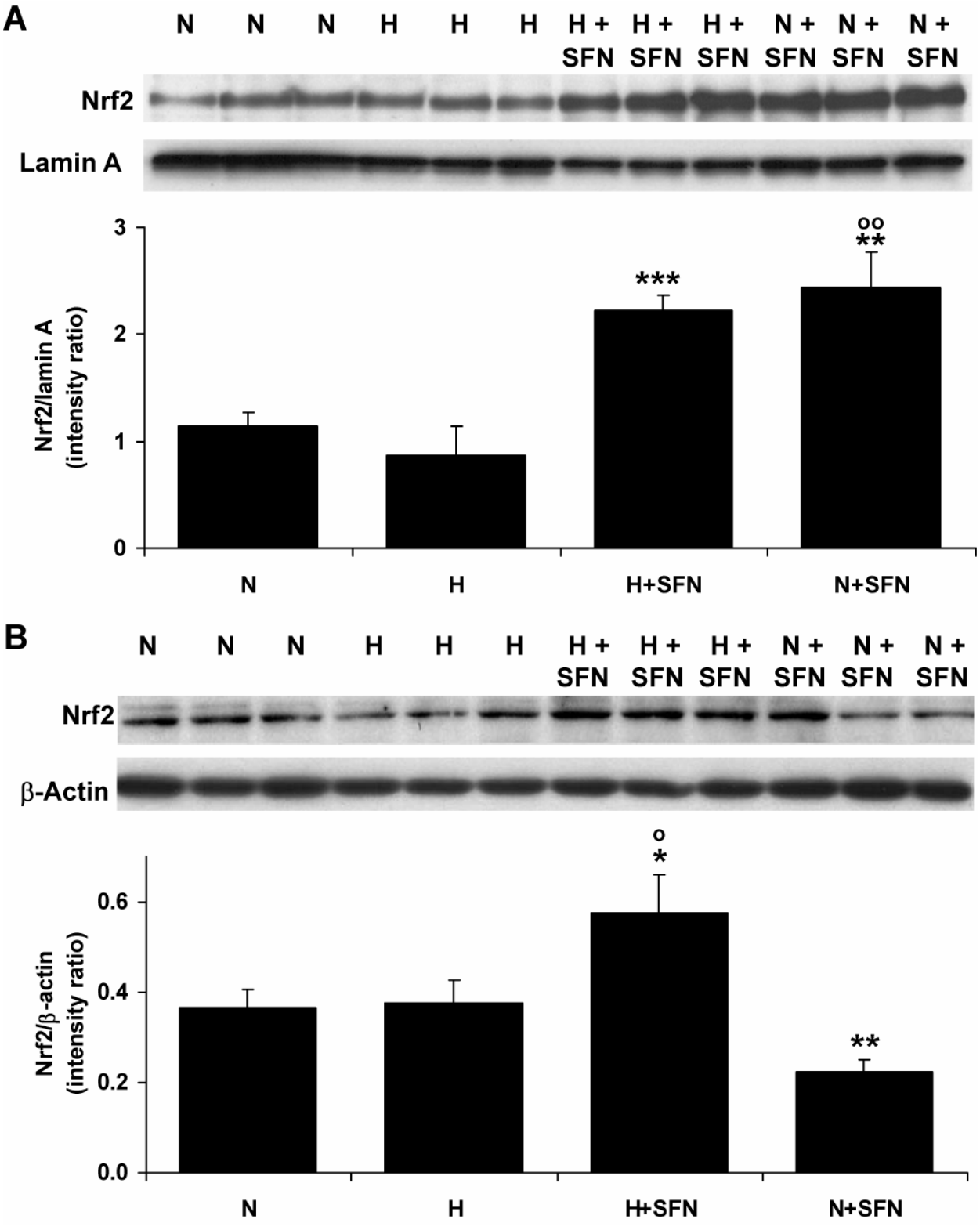
A
N N N H H H H + SFN H + SFN H + SFN N + SFN N + SFN N + SFN
Nrf2
Lamin A
Nrf2/lamin A (intensity ratio)
3
2
1
0
***
oo
**
N
H
H+SFN
N+SFN
B
N N N H H H H + SFN H + SFN H + SFN N + SFN N + SFN N + SFN
Nrf2
β-Actin
Nrf2/β-actin (intensity ratio)
0.6
0.4
0.2
0.0
o
*
**
N
H
H+SFN
N+SFN

FIG. 3. Effect of sulforaphane on ARE-linked gene expression in HMEC-1 endothelial cells in hyperglycemic culture *in vitro*. a. Effect of hyperglycemia and SFN on expression of transketolase, with and without knockdown of nrf2. b. Effect of hyperglycemia and SFN on expression of GSHRd. c. Transketolase activity of HMEC-1 cells: effect of hyperglycemia and SFN. d. Effect of hyperglycemia and SFN on expression of NQO1. TK, GSHRd and NQO1 mRNA were quantified by real time RT-PCR; AN(nrf2) – transfection for nrf2 knockdown. Data are mean ± SD (n = 3 except n = 6 for TK activity). Significance: *, ** and *** indicate P<0.05, P<0.01 and P<0.001 with respect to N; oo indicates P<0.01 with respect to H.

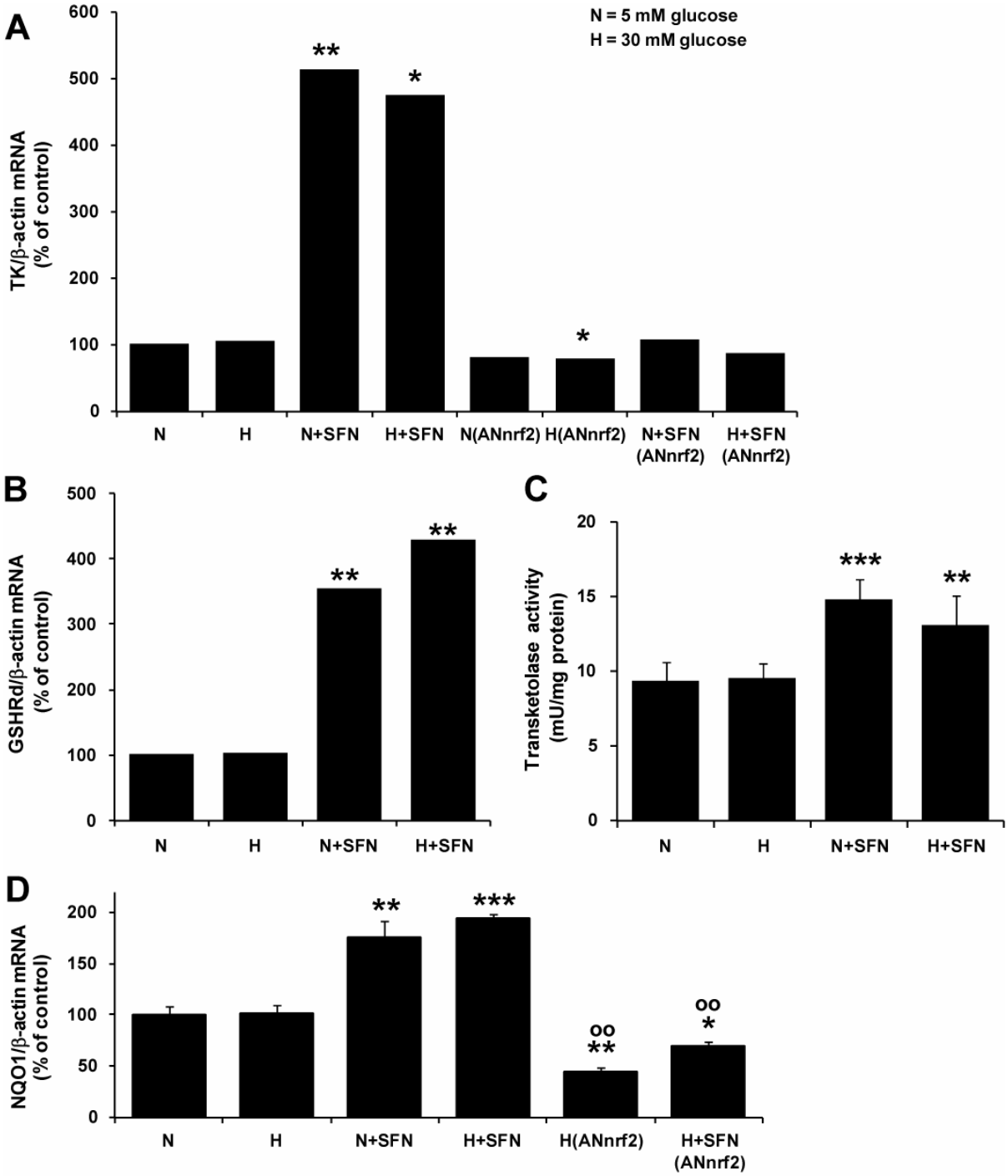

FIG. 4. Biochemical dysfunction in HMEC-1 endothelial cells in hyperglycemic culture and reversal by sulforaphane *in vitro*: reactive oxygen species formation. a. and b. Assessment of cellular ROS formation. Key: H(L-), 5 mM D(+)-glucose with 25 mM L(-)-glucose; AN(nrf2) – transfection with antisense ODN for nrf2 knockdown; AN(TK) transfection with antisense ODN for TK knockdown; and AN(scr) - transfection with scrambled sequence antisense ODN. Data are mean ± SD (n = 3). Significance: * and o represent significance with respect to normoglycemic (N) and hyperglycemic (H) control with 1, 2 and 3 symbols representing P<0.05, P<0.01 and P<0.001, respectively.

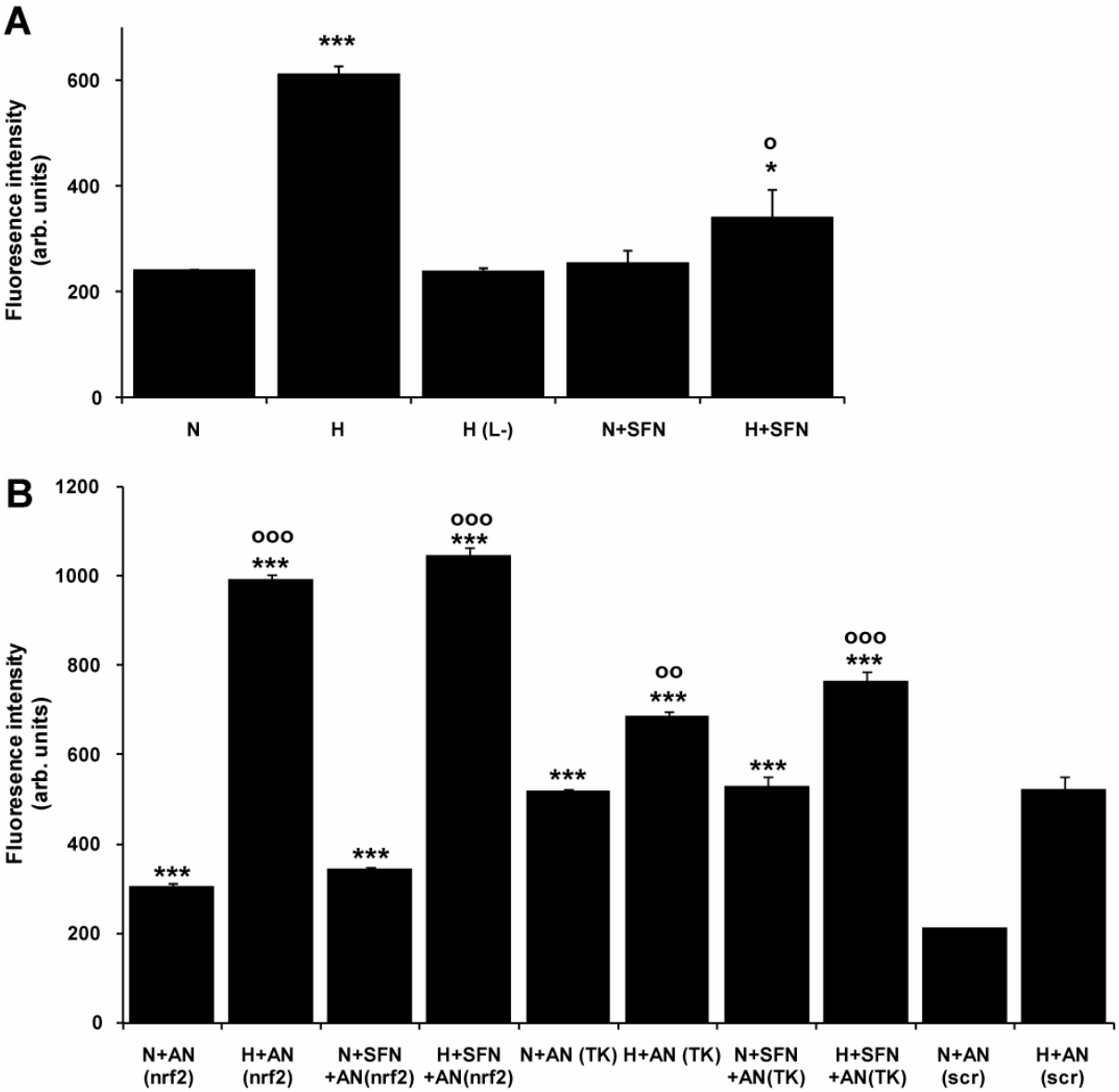

FIG. 5. Biochemical dysfunction in HMEC-1 endothelial cells in hyperglycemic culture and reversal by sulforaphane *in vitro*. a. and b. Cytosolic and membrane PKC activity, respectively. c. O-linked protein glycosylation of cell protein extracts. d. Effect on methylglyoxal concentrations: concentration in culture medium (solid bars) and cellular content (open bars). Data are mean ± SD (n = 3). Significance: *, ** and *** indicate P<0.05, P<0.01 and P<0.001 with respect to N; oo and ooo, indicate P<0.01 and P<0.001 with respect to H.