

28. J. Savarino, M. Legrand, *J. Geophys. Res.* **103**, 8267 (1998).
29. P. D. Jones, M. E. Mann, *Rev. Geophys.* **42**, 2003RG000143 (2004).
30. A. Moberg, D. M. Sonechkin, K. Holmgren, N. M. Datsenko, W. Karlen, *Nature* **433**, 613 (2005).
31. M. O. Andreae, P. Merlet, *Global Biogeochem. Cycles* **15**, 955 (2001).
32. C. McEvedy, R. Jones, *Atlas of World Population History* (Penguin, London, 1978), pp. 368.
33. W. Denevan, *The Native Population of the Americas in 1492* (Univ. of Wisconsin Press, Madison, WI, 1992), pp. 386.
34. B. Glaser, L. Haumaier, G. Guggenberger, W. Zech, *Naturwissenschaften* **88**, 37 (2001).
35. C. MacFarling Meure, thesis, University of Melbourne (2004).
36. We thank the staff of the Australian Antarctic Program, especially Casey Station, for field support; A. Smith for firm-air sampling assistance; the Bureau of Meteorology (Australia) for Cape Grim archive-air collection assistance; R. Francey, P. Steele, C. Allison, and S. Coram at CSIRO for logistical and technical help; and especially B. Ruddiman and B. Allan for valuable discussions. Supported by NSF (grant no. OPP0087357); NOAA/Climate Modeling and Diagnostics Laboratory; NIWA, New Zea-

land (Foundation for Research Science and Technology grant no. C01X0204); and the Australian Government's Antarctic Climate and Ecosystems Cooperative Research Centre and CSIRO Atmospheric Research.

#### Supporting Online Material

[www.sciencemag.org/cgi/content/full/309/5741/1714/DC1](http://www.sciencemag.org/cgi/content/full/309/5741/1714/DC1)

Materials and Methods  
References and Notes

23 May 2005; accepted 28 July 2005  
10.1126/science.1115193

## Microcephalin, a Gene Regulating Brain Size, Continues to Evolve Adaptively in Humans

Patrick D. Evans,<sup>1,2</sup> Sandra L. Gilbert,<sup>1</sup> Nitzan Mekel-Bobrov,<sup>1,2</sup> Eric J. Vallender,<sup>1,2</sup> Jeffrey R. Anderson,<sup>1</sup> Leila M. Vaez-Azizi,<sup>1</sup> Sarah A. Tishkoff,<sup>4</sup> Richard R. Hudson,<sup>3</sup> Bruce T. Lahn<sup>1\*</sup>

The gene *Microcephalin* (*MCPH1*) regulates brain size and has evolved under strong positive selection in the human evolutionary lineage. We show that one genetic variant of *Microcephalin* in modern humans, which arose ~37,000 years ago, increased in frequency too rapidly to be compatible with neutral drift. This indicates that it has spread under strong positive selection, although the exact nature of the selection is unknown. The finding that an important brain gene has continued to evolve adaptively in anatomically modern humans suggests the ongoing evolutionary plasticity of the human brain. It also makes *Microcephalin* an attractive candidate locus for studying the genetics of human variation in brain-related phenotypes.

The most distinct trait of *Homo sapiens* is the exceptional size and complexity of the brain (1, 2). Several recent studies have linked specific genes to the evolution of the human brain (3–12). One of these is *Microcephalin* (7, 8); mutations in this gene cause primary microcephaly [MCPH; Online Mendelian Inheritance in Man (OMIM) accession 251200] (13, 14). MCPH is defined clinically as severe reductions in brain size coupled with mental retardation, but remarkably, an overall retention of normal brain structure and a lack of overt abnormalities outside of the nervous system (15–17). This led to the notion that the brains of MCPH patients function normally for their size and that genes underlying MCPH are specific developmental regulators of brain size (15–17).

*Microcephalin* is one of six known loci, named *MCPH1* through *MCPH6*, for which recessive mutations lead to MCPH (14, 18–23). For four of these, the underlying genes have been identified as *Microcephalin* (*MCPH1*), *CDK5RAP2* (*MCPH3*), *ASPM* (*MCPH5*), and

*CENPJ* (*MCPH6*) (14, 21, 23). Patients with loss-of-function mutations in *Microcephalin* have cranial capacities about 4 SD below the mean at birth. As adults, their typical brain size is around 400 cm<sup>3</sup> (whereas the normal range is 1200 to 1600 cm<sup>3</sup>), and the cerebral cortex is especially small (13, 14). *Microcephalin* is suggested to control the proliferation and/or differentiation of neuroblasts during neurogenesis. This postulate was consistent with several observations. First, mouse *Microcephalin* is expressed prominently in the proliferative zones of the embryonic brain (14). Second, the *Microcephalin* protein contains several copies of the BRCT domain that is found in cell cycle regulators, such as *BRCAl* (14, 24). Finally, cell culture studies indeed suggested a role of *Microcephalin* in regulating cell cycle (25–27).

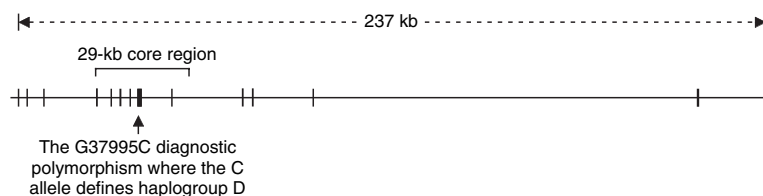
The finding that *Microcephalin* is a critical regulator of brain size spurred the hypothesis

that it might have played a role in brain evolution (16, 28). Consistent with this hypothesis, phylogenetic analysis of *Microcephalin* revealed signatures of strong positive selection in the lineage leading to humans (7, 8). Here, we examine the possibility that positive selection has continued to operate on this gene after the emergence of anatomically modern humans.

The human *Microcephalin* locus has 14 exons spanning about 236 kb on chromosome 8p23 (14) (Fig. 1). We previously sequenced all the exons in 27 humans (8). When re-analyzing the data, we noticed that one haplotype had a much higher frequency than the other haplotypes. Additionally, this haplotype differed consistently from the others at position 37995 of the genomic sequence (counting from the start codon) or position 940 of the open reading frame. This polymorphism falls in exon 8 and changes amino acid residue 314 from an ancestral aspartate to a histidine. (This polymorphism is described as G37995C with G denoting the ancestral allele.)

To investigate whether positive selection has acted on the high-frequency haplotype, we resequenced 23.4 kb of a 29-kb region centered around the G37995C polymorphism (Fig. 1). Sequencing was performed on a panel of 89 individuals from the Coriell Institute, which broadly represents human diversity (see SOM). To assign the ancestral state of polymorphisms, we also sequenced the common chimpanzee. Several GC-rich segments were not sequenced because of technical difficulties. The resulting sequence data contained 220 polymorphic sites, including 213 single-nucleotide polymorphisms (SNPs) and 7 insertion/deletion polymorphisms (indels) (table S1).

Haplotypes were inferred using the PHASE 2.1 program (29, 30). A total of 86 haplotypes



**Fig. 1.** Genomic structure of the human *Microcephalin* gene. The region sequenced in the 89-individual Coriell panel is bracketed.

<sup>1</sup>Howard Hughes Medical Institute, Department of Human Genetics, <sup>2</sup>Committee on Genetics, <sup>3</sup>Department of Ecology and Evolution, University of Chicago, Chicago, IL 60637, USA. <sup>4</sup>Department of Biology, University of Maryland, College Park, MD 20742, USA.

\*To whom correspondence should be addressed. E-mail: [blahn@bsd.uchicago.edu](mailto:blahn@bsd.uchicago.edu)

were identified along with their frequencies (Fig. 2 and table S2). One haplotype, denoted 49, had a much higher frequency than the other haplotypes. It had the derived C allele at the G37995C SNP site and corresponded to the high-frequency haplotype in the aforementioned exon-only polymorphism survey (8). In the Coriell panel, haplotype 49 had a frequency of 33% (59 out of 178 chromosomes) and is found in all the populations sampled in the panel. The remaining 85 haplotypes varied in frequency from 0.6 to 6.2% (1 to 11 chromosomes).

Positive selection on an allele can increase the frequency of the haplotype bearing the allele while maintaining extended linkage disequilibrium (LD) around that allele (31–36). Our data on haplotype 49 are consistent with these signatures of selection. We formally tested the statistical significance of positive selection using the previously established coalescent model (37, 38). Given the slight uncertainty in haplotype inference, we considered only the 18 individuals in the Coriell panel who are homozygous for haplotype 49 (table S1).

By simulation, we calculated the probability of obtaining 18 or more individuals (out of 89) who are homozygous for a single haplotype across a region of 220 segregating sites under neutral evolution. Here, recombination and gene conversion rates were set to values previously established for the *Microcephalin* locus (39), and a demographic model with a severe bottleneck followed by exponential growth was assumed (see SOM). Prior studies have shown that the bottleneck specified here is likely to be much more stringent than that associated with the real demographic history of human populations (40, 41); thus, the test is conservative (38). Under these parameters, the probability of obtaining 18 homozygotes out of 89 is highly significant ( $P = 0$  based on 5,000,000 replicates).

We then tested several additional demographic models, including (i) constant size, (ii) very ancient expansion, (iii) very recent expansion, (iv) repeated severe bottlenecks with subsequent expansion, and (v) population structure with between two and five subpopulations (see SOM). All produced exceedingly significant results. Even though the exact demographic history of humans is yet to be defined, our tests are highly significant under a broad range of demographic scenarios, which furthers the argument that the statistical significance is unlikely to be altered by reasonable variations in the supposed human demography. We also tested the significance of the inferred haplotype data (i.e., the significance of having 59 copies of haplotype 49 among 178 chromosomes), which similarly produced highly significant results. These data strongly suggest that haplotype 49 was driven to high frequency by positive selection. However, our data do not address whether the positive selection is

frequency-dependent selection, heterozygote advantage, or simple additive positive selection.

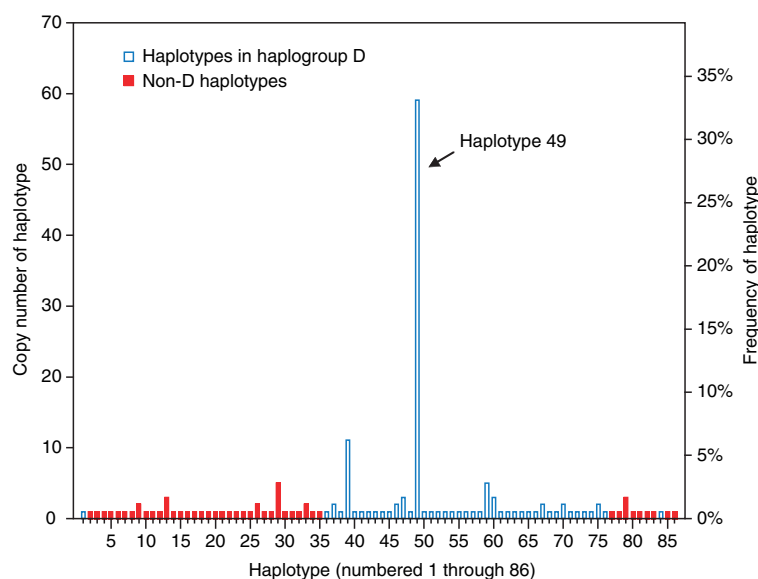
Using the G37995C polymorphism as a diagnostic site, we divided all the haplotypes into two groups: those that carry the derived C allele and those that carry the ancestral G allele. We designated the former group as haplogroup D (where D stands for “derived”). It includes 43 haplotypes that together have a 70% frequency in the Coriell panel, and haplotype 49 is the predominant member (table S2). Although the derived C allele at the G37995C site only provides an operational definition for haplogroup D, several observations make evident that haplogroup D is systematically different from the non-D haplotypes. First, this haplogroup consists exclusively of haplotype 49 or its minor variants, whereas non-D haplotypes show much greater sequence divergence from haplogroup D chromosomes. This greater divergence is because haplogroup D and non-D haplotypes have multiple fixed differences relative to each other in addition to G37995C (table S2). The only exceptions are a few recombinant haplotypes between D and non-D chromosomes (discussed below). Second, for sites that are polymorphic within haplogroup D chromosomes (excluding recombinants between D and non-D chromosomes), the non-D chromosomes are invariably monomorphic for the ancestral alleles. These data indicate that haplogroup D constitutes a genealogical clade of closely related haplotypes that is altogether separate from the more distantly related non-D haplotypes (again, excluding recombinants between D and non-D chromosomes, which represent mixed genealogies).

Collectively, the above observations support an evolutionary scenario with two aspects.

First, haplotype 49 swept from a single copy to high frequency in a short period of time. Second, during the sweep, minor variants of haplotype 49 emerged through rare mutations and recombinations. These variants, together with haplotype 49, make up haplogroup D. Haplotype 49 evidently represents the most recent common ancestor (MRCA) of haplogroup D, because it consistently has the ancestral allele for the sites polymorphic within haplogroup D.

We next estimated the coalescence age (i.e., time to MRCA) of haplogroup D chromosomes in the Coriell panel. We used the average number of mutations from the MRCA of a haplogroup clade to its descendant lineages as a molecular clock for estimating the age of the clade (42, 43). This approach is known to be unbiased by demographic history (42). The age of haplogroup D was found to be ~37,000 years, with a 95% confidence interval of 14,000 to 60,000 years. In comparison, the coalescence age of all the chromosomes in the Coriell panel is about 1,700,000 years. The emergence of anatomically modern humans has been estimated to be 200,000 years before present (44). Haplogroup D is obviously much younger, which indicates that positive selection was at work in a period considerably postdating the emergence of anatomically modern humans in Africa. We note that the age of haplogroup D coincides with the introduction of anatomically modern humans into Europe about 40,000 years ago, as well as the dramatic shift in the archeological record indicative of modern human behavior, such as art and the use of symbolism (i.e., the “Upper Paleolithic revolution”) (45).

If haplogroup D indeed experienced a recent selective sweep, it should show low poly-



**Fig. 2.** Frequencies of 86 inferred *Microcephalin* haplotypes in the 89-individual Coriell panel. Haplotypes in haplogroup D are indicated by blue-edged bars; non-D haplotypes are indicated by solid red bars.

morphism and an excess of rare alleles (46). To confirm this, we calculated nucleotide diversity ( $\pi$ ) and Tajima's  $D$  for the 47 individuals who are homozygous for haplogroup D chromosomes, and we compared these values to those of the non-D chromosomes. The  $\pi$  value of the D chromosomes is lower, by a factor of 12, than that of the non-D chromosomes (0.000077 and 0.00092, respectively), even though the D chromosomes represent about 70% of the chromosomes in the panel. Tajima's  $D$ , which is a summary statistic for the frequency spectrum of alleles, is  $-2.3$  for haplogroup D (whereas it is  $-1.2$  for the non-D chromosomes). This strongly negative Tajima's  $D$  indicates a starlike genealogy for haplogroup D chromosomes (47). Thus, both summary statistics contrast sharply between D and non-D chromosomes and are consistent with the recent age and rapid expansion of haplogroup D. We note that these calculations do not provide a statistically stringent test of positive selection, because they are done on subsets of the genealogy. Nevertheless, they do

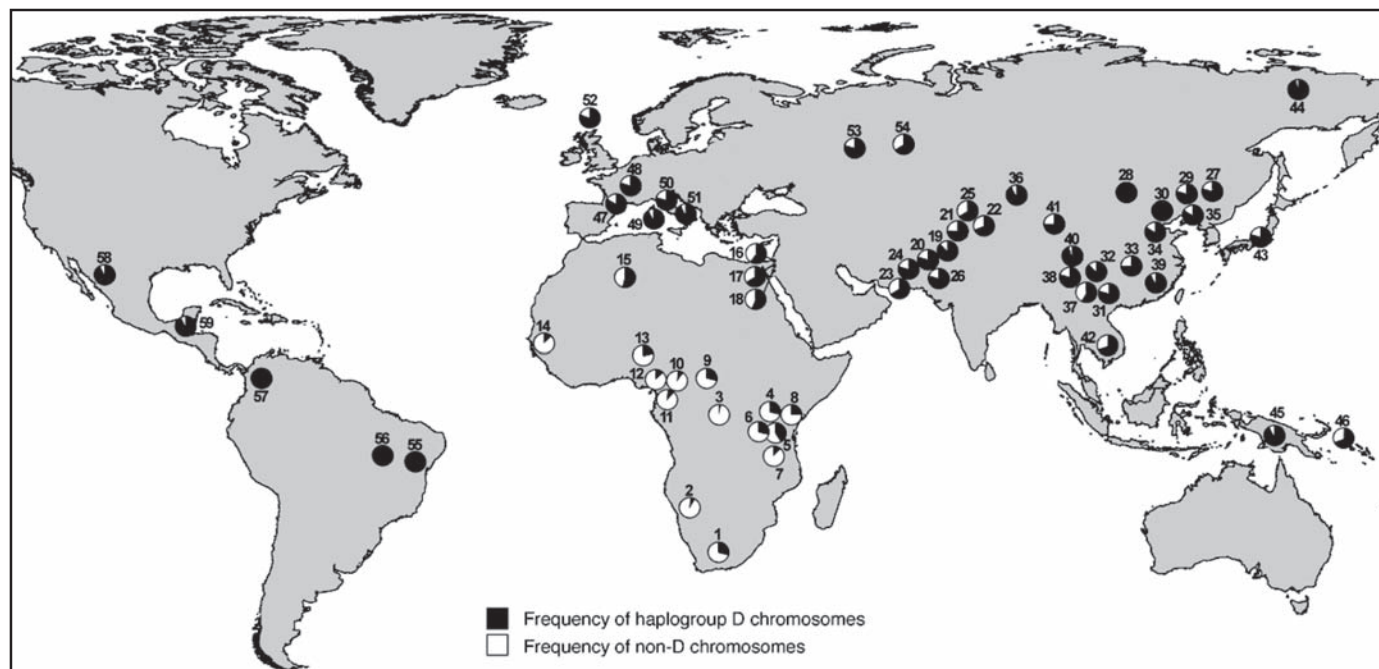
reveal qualitative signatures of positive selection that further corroborate the more stringent statistical tests described earlier.

Another sign of a positive selective sweep is extended LD around the selected allele. This is apparent in the region of *Microcephalin* investigated here, where haplogroup D chromosomes show near-complete LD across the entire region. The only exceptions are haplotypes 1, 68, and 84 (each found in a single copy in the Coriell panel), which are recombinants between D and non-D chromosomes as evidenced by recombination tracts (table S2). The remaining 121 copies of haplogroup D chromosomes show no evidence of recombination. By comparison, the non-D chromosomes do not display any significant LD across the region.

To probe the extent of LD beyond the 29-kb core region, we sequenced the Coriell panel for two segments of about 3 kb each, situated at the beginning and end of the gene separated from each other by about 235 kb. In these flanking regions, there is clear evidence

of LD decay from the core region, which supports the idea that selection has most likely operated on a site (or sites) around the core region. Our present data cannot resolve the exact site(s) of selection, and the G37995C nonsynonymous SNP used to define haplogroup D is just a candidate.

To obtain a more detailed frequency distribution of haplogroup D across the globe, we analyzed a much larger human population panel containing 1184 globally diverse individuals. We genotyped the diagnostic G37995C SNP in this panel to infer the frequency of haplogroup D chromosomes (Fig. 3). Geographic variation was observed, with sub-Saharan populations generally having lower frequencies than others. The statistic for genetic differentiation,  $F_{ST}$ , is 0.48 between sub-Saharan and others, which indicates strong differentiation (48) and is significantly higher than the genome average of 0.12 ( $P < 0.03$  based on previously established genomewide  $F_{ST}$  distribution) (49). Such population differentiation may reflect a Eurasian origin of haplogroup D, local adaptation, and/or



**Fig. 3.** Global frequencies of *Microcephalin* haplogroup D chromosomes (defined as having the derived C allele at the G37995C diagnostic SNP) in a panel of 1184 individuals. For each population, the country of origin, number of individuals sampled, and frequency of haplogroup D chromosomes are given (in parentheses) as follows: 1, Southeastern and Southwestern Bantu (South Africa, 8, 31.3%); 2, San (Namibia, 7, 7.1%); 3, Mbuti Pygmy (Democratic Republic of Congo, 15, 3.3%); 4, Masai (Tanzania, 27, 29.6%); 5, Sandawe (Tanzania, 32, 39.1%); 6, Burunge (Tanzania, 28, 30.4%); 7, Turu (Tanzania, 23, 15.2%); 8, Northeastern Bantu (Kenya, 12, 25%); 9, Biaka Pygmy (Central African Republic, 32, 26.6%); 10, Zime (Cameroon, 23, 8.7%); 11, Bakola Pygmy (Cameroon, 24, 10.4%); 12, Bamoun (Cameroon, 28, 17.9%); 13, Yoruba (Nigeria, 25, 24%); 14, Mandenka (Senegal, 24, 16.7%); 15, Mozabite [Algeria (Mzab region), 29, 53.5%]; 16, Druze [Israel (Carmel region), 44, 60.2%]; 17, Palestinian [Israel (Central), 40, 63.8%]; 18, Bedouin [Israel (Negev region), 44, 54.6%]; 19, Hazara (Pakistan, 20, 85%); 20, Balochi (Pakistan, 23, 78.3%); 21, Pathan (Pakistan, 23, 76.1%); 22, Burusho (Pakistan, 25, 66%); 23, Makrani (Pakistan, 24,

62.5%); 24, Brahui (Pakistan, 25, 78%); 25, Kalash (Pakistan, 24, 62.5%); 26, Sindhi (Pakistan, 25, 78%); 27, Hezhen (China, 9, 77.8%); 28, Mongola (China, 10, 100%); 29, Daur (China, 10, 85%); 30, Orogen (China, 10, 100%); 31, Miaozi (China, 9, 77.8%); 32, Yizu (China, 10, 85%); 33, Tujia (China, 10, 75%); 34, Han (China, 41, 82.9%); 35, Xibo (China, 9, 83.3%); 36, Uygur (China, 10, 90%); 37, Dai (China, 9, 55.6%); 38, Lahu (China, 10, 85%); 39, She (China, 9, 88.9%); 40, Naxi (China, 10, 95%); 41, Tu (China, 10, 75%); 42, Cambodian (Cambodia, 11, 72.7%); 43, Japanese (Japan, 27, 77.8%); 44, Yakut [Russia (Siberia region), 25, 98%]; 45, Papuan (New Guinea, 17, 91.2%); 46, NAN Melanesian (Bougainville, 18, 72.2%); 47, French Basque (France, 24, 83.3%); 48, French (France, 28, 78.6%); 49, Sardinian (Italy, 26, 90.4%); 50, North Italian [Italy (Bergamo region), 13, 76.9%]; 51, Tuscan (Italy, 8, 87.5%); 52, Orcadian (Orkney Islands, 16, 81.3%); 53, Russian (Russia, 24, 79.2%); 54, Adygei [Russia (Caucasus region), 15, 63.3%]; 55, Karitiana (Brazil, 21, 100%); 56, Surui (Brazil, 20, 100%); 57, Colombian (Colombia, 11, 100%); 58, Pima (Mexico, 25, 92%); 59, Maya (Mexico, 25, 92%).

demographic factors such a bottleneck associated with human migration out of Africa 50,000 to 100,000 years ago.

Previous studies have shown that *Microcephalin* is a specific regulator of brain size (13, 14) and that this gene has evolved under strong positive selection in the primate lineage leading to *Homo sapiens* (7, 8). Here, we present compelling evidence that *Microcephalin* has continued its trend of adaptive evolution beyond the emergence of anatomically modern humans. The specific function of *Microcephalin* in brain development makes it likely that selection has operated on the brain. Yet, it remains formally possible that an unrecognized function of *Microcephalin* outside of the brain is actually the substrate of selection. If selection indeed acted on a brain-related phenotype, there could be several possibilities, including brain size, cognition, personality, motor control, or susceptibility to neurological and/or psychiatric diseases. We hypothesize that D and non-D haplotypes have different effects on the proliferation of neural progenitor cells, which in turn leads to different phenotypic outcomes of the brain visible to selection.

References and Notes

1. J. N. Spuhler, *The Evolution of Man's Capacity for Culture* (Wayne State Univ. Press, Detroit, MI, 1959).
2. J. H. Jerison, *Evolution of the Brain and Intelligence* (Academic Press, New York, 1973).
3. W. Enard et al., *Nature* **418**, 869 (2002).

4. J. Zhang, *Genetics* **165**, 2063 (2003).
5. P. D. Evans et al., *Hum. Mol. Genet.* **13**, 489 (2004).
6. N. Kouprina et al., *PLoS Biol.* **2**, E126 (2004).
7. Y. Q. Wang, B. Su, *Hum. Mol. Genet.* **13**, 1131 (2004).
8. P. D. Evans, J. R. Anderson, E. J. Vallender, S. S. Choi, B. T. Lahn, *Hum. Mol. Genet.* **13**, 1139 (2004).
9. R. J. Ferland et al., *Nat. Genet.* **36**, 1008 (2004).
10. H. H. Stedman et al., *Nature* **428**, 415 (2004).
11. F. Burki, H. Kaessmann, *Nat. Genet.* **36**, 1061 (2004).
12. S. Dorus et al., *Cell* **119**, 1027 (2004).
13. A. P. Jackson et al., *Am. J. Hum. Genet.* **63**, 541 (1998).
14. A. P. Jackson et al., *Am. J. Hum. Genet.* **71**, 136 (2002).
15. W. B. Dobyns, *Am. J. Hum. Genet.* **112**, 315 (2002).
16. G. H. Mochida, C. A. Walsh, *Curr. Opin. Neurol.* **14**, 151 (2001).
17. C. G. Woods, J. Bond, W. Enard, *Am. J. Hum. Genet.* **76**, 717 (2005).
18. E. Roberts et al., *Eur. J. Hum. Genet.* **7**, 815 (1999).
19. L. Moynihan et al., *Am. J. Hum. Genet.* **66**, 724 (2000).
20. C. R. Jamieson, C. Govaerts, M. J. Abramowicz, *Am. J. Hum. Genet.* **65**, 1465 (1999).
21. J. Bond et al., *Nat. Genet.* **32**, 316 (2002).
22. G. F. Leal et al., *J. Med. Genet.* **40**, 540 (2003).
23. J. Bond et al., *Nat. Genet.* **37**, 353 (2005).
24. T. Huyton, P. A. Bates, X. Zhang, M. J. Sternberg, P. S. Freemont, *Mutat. Res.* **460**, 319 (2000).
25. S. Y. Lin, S. J. Eledge, *Cell* **113**, 881 (2003).
26. X. Xu, J. Lee, D. F. Stern, *J. Biol. Chem.* **279**, 34091 (2004).
27. M. Trimborn et al., *Am. J. Hum. Genet.* **75**, 261 (2004).
28. S. L. Gilbert, W. B. Dobyns, B. T. Lahn, *Nat. Rev. Genet.* **6**, 581 (2005).
29. M. Stephens, N. J. Smith, P. Donnelly, *Am. J. Hum. Genet.* **68**, 978 (2001).
30. M. Stephens, P. Donnelly, *Am. J. Hum. Genet.* **73**, 1162 (2003).
31. S. A. Tishkoff et al., *Science* **293**, 455 (2001).
32. P. C. Sabeti et al., *Nature* **419**, 832 (2002).
33. E. Wang et al., *Am. J. Hum. Genet.* **74**, 931 (2004).
34. T. Bersaglieri et al., *Am. J. Hum. Genet.* **74**, 1111 (2004).
35. E. E. Thompson et al., *Am. J. Hum. Genet.* **75**, 1059 (2004).
36. H. Stefansson et al., *Nat. Genet.* **37**, 129 (2005).

37. R. R. Hudson, *Oxf. Surv. Evol. Biol.* **7**, 1 (1990).
38. R. R. Hudson, *Bioinformatics* **18**, 337 (2002).
39. A. Kong et al., *Nat. Genet.* **31**, 241 (2002).
40. E. Zietkiewicz et al., *J. Mol. Evol.* **47**, 146 (1998).
41. H. Harpending, A. Rogers, *Annu. Rev. Genomics Hum. Genet.* **1**, 361 (2000).
42. H. Tang, D. O. Siegmund, P. Shen, P. J. Oefner, M. W. Feldman, *Genetics* **161**, 447 (2002).
43. R. Thomson, J. K. Pritchard, P. Shen, P. J. Oefner, M. W. Feldman, *Proc. Natl. Acad. Sci. U.S.A.* **97**, 7360 (2000).
44. I. McDougall, F. H. Brown, J. G. Fleagle, *Nature* **433**, 733 (2005).
45. R. G. Klein, *The Human Career: Human Biological and Cultural Origins* (Univ. of Chicago Press, Chicago, 1999).
46. M. Bamshad, S. P. Wooding, *Nat. Rev. Genet.* **4**, 99 (2003).
47. F. Tajima, *Genetics* **123**, 585 (1989).
48. S. Wright, *Evolution and the Genetics of Populations* (Univ. of Chicago Press, Chicago, 1978).
49. J. M. Akey, G. Zhang, K. Zhang, L. Jin, M. D. Shriver, *Genome Res.* **12**, 1805 (2002).
50. We thank the Coriell Institute for Medical Research, the Centre d'Etude du Polymorphisme Humain (CEPH), and A. Froment for human DNA samples. We thank H. M. Cann, S. Dorus, E. E. Eichler, N. M. Pearson, A. Di Rienzo, M. Kreitman, and J. K. Pritchard for technical support and/or helpful discussions. Supported in part by the Searle Scholarship and the Burroughs Wellcome Career Award (to B.T.L.), and David and Lucile Packard Career Award, the Burroughs Wellcome Career Award, and NSF grant BCS-0196183 (to S.A.T.).

Supporting Online Material

www.sciencemag.org/cgi/content/full/309/5741/1717/DC1  
 Materials and Methods  
 Tables S1 and S2  
 References and Notes

18 April 2005; accepted 14 June 2005  
 10.1126/science.1113722

# Ongoing Adaptive Evolution of *ASPM*, a Brain Size Determinant in *Homo sapiens*

Nitzan Mekel-Bobrov,<sup>1,2</sup> Sandra L. Gilbert,<sup>1</sup> Patrick D. Evans,<sup>1,2</sup> Eric J. Vallender,<sup>1,2</sup> Jeffrey R. Anderson,<sup>1</sup> Richard R. Hudson,<sup>3</sup> Sarah A. Tishkoff,<sup>4</sup> Bruce T. Lahn<sup>1\*</sup>

The gene *ASPM* (*abnormal spindle-like microcephaly associated*) is a specific regulator of brain size, and its evolution in the lineage leading to *Homo sapiens* was driven by strong positive selection. Here, we show that one genetic variant of *ASPM* in humans arose merely about 5800 years ago and has since swept to high frequency under strong positive selection. These findings, especially the remarkably young age of the positively selected variant, suggest that the human brain is still undergoing rapid adaptive evolution.

Homozygous null mutations of *ASPM* cause primary microcephaly, a condition characterized by severely reduced brain size with otherwise normal neuroarchitecture (1). Studies

have suggested that *ASPM* may regulate neural stem cell proliferation and/or differentiation during brain development, possibly by mediating spindle assembly during cell division (1, 2). Phylogenetic analysis of *ASPM* has revealed strong positive selection in the primate lineage leading to *Homo sapiens* (3–5), especially in the past 6 million years of hominid evolution in which *ASPM* acquired about one advantageous amino acid change every 350,000 years (4). These data argue that *ASPM*

may have contributed to human brain evolution (3–6). Here, we investigate whether positive selection has continued to operate on *ASPM* since the emergence of anatomically modern humans.

Human *ASPM* has 28 exons with a 10,434–base pair open reading frame (1) (fig. S1). We resequenced the entire 62.1-kb genomic region of *ASPM* in samples from 90 ethnically diverse individuals obtained through the Coriell Institute and from a common chimpanzee (7). This revealed 166 polymorphic sites (table S1). Using established methodology (7), we identified 106 haplotypes. One haplotype, numbered 63, had an unusually high frequency of 21%, whereas the other haplotypes ranged from 0.56% to 3.3% (fig. S2). Moreover, this haplotype differed consistently from the others at multiple polymorphic sites (save for a few rare haplotypes that are minor mutational or recombinational variants of haplotype 63, as discussed later) (table S2). Two of these polymorphic sites are nonsynonymous, both in exon 18, and are denoted A44871G and C45126A (numbers indicate genomic positions from the start codon, and letters at the beginning and end indicate ancestral and derived alleles, respectively). These two sites reside in a region of the open reading frame that was shown previously to have experienced par-

<sup>1</sup>Howard Hughes Medical Institute, Department of Human Genetics, <sup>2</sup>Committee on Genetics, <sup>3</sup>Department of Ecology and Evolution, University of Chicago, Chicago, IL 60637, USA. <sup>4</sup>Department of Biology, University of Maryland, College Park, MD 20742, USA.

\*To whom correspondence should be addressed. E-mail: blahn@bsd.uchicago.edu

**Science Supporting Online Material*****Microcephalin*, a Gene Regulating Brain Size, Continues to Evolve Adaptively in Humans**

Patrick D. Evans, Sandra L. Gilbert, Nitzan Mekel-Bobrov, Eric J. Vallender, Jeffrey R. Anderson, Leila M. Vaez-Azizi, Sarah A. Tishkoff, Richard R. Hudson, Bruce T. Lahn

DOI: 10.1126/science.1113722

**Materials and Methods***Sequence acquisition and preliminary analysis*

A panel of 89 human samples from the Coriell Institute that broadly represent worldwide populations was used for resequencing. It includes 9 sub-Saharan Africans (Coriell numbers: 17341–17349), 7 North Africans (17378–17384), 9 Iberians (17091–17097, 17099, 17100), 7 Basques (15883–15887, 16185, 16188), 9 Russians (13820a, 13838, 13852, 13876, 13877, 13911–13914), 9 Middle Easterners (17331–17340), 9 South Asians (17021–17024, 17026–17030), 8 Chinese (16654, 16688, 16689, 17014, 17015, 17017–17019), 1 Japanese (11587), 8 Southeast Asians (17081, 17083, 17085–17090), 6 Pacific Islander (17385–17388, 17390, 17391), and 7 Andeans (17301, 17302, 17306–17310). A common chimpanzee (*Pan troglodytes*) was also included in the sequencing. Double-stranded sequences in regions of interest were obtained by PCR amplification followed by sequencing of PCR products. Sequenced regions include 24750–26292, 26988–29992, 30561–32132, 32841–42938, 43006–44351, 45808–49406, 50123–50908, and 52305–53776 (the first base of the initiation codon of *Microcephalin* is defined as position 1). The core region used for haplotype analysis spans 29027 bases (24750–53776), of which 23416 bases were sequenced. Sequence chromatograms were aligned by the Sequencher software (Gene Codes Corporation, Ann Arbor, MI). Polymorphisms were detected by direct visual inspection of sequence chromatograms. The ancestral alleles of polymorphisms were called using the chimpanzee sequence as outgroup. Inference of haplotypes from the diploid sequence data was performed using the PHASE 2.1 software as described (S1, S2), which is available online at <http://www.stats.ox.ac.uk/mathgen/home.html>. Nucleotide diversity ( $\pi$ ) and Tajima's *D* were calculated using the program DnaSP 3.51, as described previously (S3). To avoid uncertainties of haplotype inference, the 47 individuals who are homozygous for haplogroup D chromosomes were used for the calculation of  $\pi$  and Tajima's *D* of this haplogroup. Inferred haplotypes were used to calculate  $\pi$  and Tajima's *D* for the non-D chromosomes. Recombinants between D and non-D chromosomes were excluded from the calculation.

*Genotyping*

Genotyping of the G37995C nonsynonymous polymorphism in *Microcephalin* was performed on a panel of 1184 human samples. This panel does not overlap with the Coriell panel described above. It consists of the HGDP CEPH diversity panel as described previously (S4), minus the following two sets of samples. One is a set of duplicated samples that needed to be removed, including HGDP00472, HGDP00452, HGDP00457, HGDP00980, HGDP00650, HGDP00583, HGDP00111, HGDP00220, HGDP00813, HGDP01233, HGDP00762, HGDP00770, HGDP00657, HGDP00658, HGDP00660, and HGDP01149. The other is a set of samples that failed to be

genotyped due to technical reasons (e.g., poor DNA quality), including HGDP01263, HGDP00633, HGDP00635, HGDP00636, HGDP00644, HGDP00579, HGDP00581, HGDP00584, HGDP00698, HGDP00700, HGDP00722, HGDP00723, HGDP00724, HGDP00725, HGDP00730, HGDP00731, HGDP00732, HGDP00734, HGDP00746, HGDP00076, HGDP00090, HGDP00109, HGDP00115, HGDP00122, HGDP00125, HGDP00141, HGDP00254, HGDP00281, HGDP00782, HGDP00783, HGDP01023, HGDP01193, HGDP01311, HGDP01334, HGDP00766, HGDP00768, HGDP00662, HGDP00520, HGDP00666, HGDP01077, HGDP01386, HGDP01402, HGDP00890, HGDP00707, HGDP00708, HGDP00995, HGDP00998, HGDP01010, and HGDP00841. The CEPH panel originally contained 1064 individuals, and had 999 individuals remaining after removing the above two sets of samples. Demographic information for the HGDP CEPH diversity panel is available online at <http://www.cephb.fr>. In addition, the panel contained 185 sub-Saharan African samples collected by S. A. Tishkoff and A. Froment (sample collection was approved by the Institutional Review Board at the University of Maryland). The samples included 23 Turu, 32 Sandawe, 28 Burunge, and 27 Masai individuals from Tanzania; they also included 24 Bakola Pygmy, 28 Bamoun, and 23 Zime individuals from Cameroon. To perform genotyping, a small region encompassing the G37995C polymorphism was amplified by PCR, followed by sequencing of the PCR product. Genotype was scored by visual inspection of the sequence chromatograms.  $F_{ST}$  was calculated as described previously (S5). The exact formulas are available on pages 143–155 of (S6).

### *Statistical analysis*

To test the statistical significance that the frequency of haplotype 49 departs from neutral expectation, we used a previously described simulation method based on the coalescent process as implemented in the ms software (S7, S8). First, the following parameters were specified: the number of chromosomes, the number of segregating sites, recombination rate, gene conversion rate, and demographic model. Recombination rate of the *Microcephalin* region was set at the locus-specific value of 1.9 cM/Mb as obtained in a previous genomewide survey (S9), and gene conversion rate was set to be the same as recombination rate with an average tract length of 100 bp. The gene conversion model was as previously described (S10), which assumes that the tract length is geometrically distributed. Nine demographic models were tested:

- 1) constant population with an effective size of  $10^4$ ,
- 2) an ancient population expansion from  $10^4$  at 5,000 generations ago exponentially to  $10^7$  today,
- 3) a recent population expansion from  $10^4$  at 1,000 generations ago exponentially to  $10^7$  today,
- 4) a severe bottle neck starting 5,000 generations ago that reduced the population from  $10^4$  instantly to  $10^3$  and lasted until 2,500 generations ago at which point the population started to expand exponentially to  $10^7$  today,
- 5) repeated bottlenecks for five successive rounds starting 7000 generations ago, each from  $10^4$  instantly to  $10^3$  for 500 generations followed by exponential recovery back to  $10^4$  over another 500 generations, except at the end of the fifth bottleneck 2500 generations ago which was followed by exponential growth to  $10^7$  today,
- 6) population structure where the initial 178 chromosomes were split equally into 2 different subpopulations under constant population size with 1 migration per generation, and
- 7 to 9) population structure where the initial 178 chromosomes were split equally into 3 to 5 different subpopulations with 1 migration per generation. Command lines in the ms program to input the above demographic models were as follows:

- 1) Constant population size:

```
./ms 178 100000 -s 220 -r 11.6104 29027 -c 1 100 |./samh 18| wc
```

2) Ancient population expansion:

```
./ms 178 100000 -s 220 -r 11610.4 29027 -c 1 100 -G 55262.04223 -eG 0.000125 0 |./samh 18| wc
```

3) Recent population expansion:

```
./ms 178 100000 -s 220 -r 11610.4 29027 -c 1 100 -G 276310.2112 -eG 0.000025 0 |./samh 18| wc
```

4) Several bottleneck:

```
./ms 178 100000 -s 220 -r 11610.4 29027 -c 1 100 -G 147365.446 -eG 0.0000625 0 -eN 0.000125 0.001 |./samh 18| wc
```

5) Repeated bottlenecks with subsequent expansion:

```
./ms 178 100000 -s 220 -r 11610.4 29027 -c 1 100 -G 147365.446 -eG 0.0000625 0 -eN 0.000075 0.001 -eG 0.000075 184206.8074 -eG 0.0000875 0 -eN 0.0001 0.001 -eG 0.0001 184206.8074 -eG 0.0001125 0 -eN 0.000125 0.001 -eG 0.000125 184206.8074 -eG 0.0001375 0 -eN 0.00015 0.001 -eG 0.00015 184206.8074 -eG 0.0001625 0 -eN 0.000175 0.001 |./samh 18| wc
```

6) Population structure with 2 subpopulations:

```
./ms 178 100000 -s 220 -r 11.6104 29027 -c 1 100 -es 0.0 1 .5 -eM 0.0 1.0 |./samh 18| wc
```

7) Population structure with 3 subpopulations:

```
./ms 178 100000 -s 220 -r 11.6104 29027 -c 1 100 -es 0.0 1 0.3333 -es 0.0 1 0.5 -eM 0.0 1.0 |./samh 18| wc
```

8) Population structure with 4 subpopulations:

```
./ms 178 100000 -s 220 -r 11.6104 29027 -c 1 100 -es 0.0 1 .25 -es 0.0 1 .333 -es 0.0 1 .5 -eM 0.0 1.0 |./samh 18| wc
```

9) Population structure with 5 subpopulations:

```
./ms 178 100000 -s 220 -r 11.6104 29027 -c 1 100 -es 0.0 1 0.2 -es 0.0 1 0.25 -es 0.0 1 0.333 -es 0.0 1 0.5 -eM 0.0 1.0 |./samh 18| wc
```

### Age estimation

We estimated the age of haplogroup D using a mutation-based method as previously described (S11). This method simply relies on averaging the number of mutations along each lineage from the most recent common ancestor (MRCA) to the sampled chromosome. This averaging produces an estimate of the time to MRCA that is unbiased by demographic history (S11). Let  $t$  denote the time to MRCA for haplogroup D in units of mutations. The value of  $t$  could be estimated as follows: To start, we decided to focus only on the 47 individuals who are homozygous for haplogroup D chromosomes (rather than using all the inferred copies of haplogroup D). This avoided uncertainties in haplotype inference. We also note that there are no evident recombinants between D and non-D types among these 47 individuals, which is important because the absence of such recombinants is a necessary condition for our methodology (S11, S12). Using chimpanzee sequence as an outgroup, we deduced the MRCA sequence of haplogroup D, which happens to be the same as the sequence of haplotype 49. We next added up the total number of mutations separating the MRCA and the 94 chromosomes sampled in the 47 individuals. This number was 93, which was divided by 94 to yield  $\hat{t}$ , the estimate of  $t$ , at 0.989. This value was then divided by 23416 (the total length of DNA sequenced) to yield an estimate for the number of mutations per base ( $\hat{T}$ ) of  $4.2 \times 10^{-5}$ . By comparing human and chimpanzee sequences in this region, the rate of human-chimpanzee nucleotide divergence ( $D$ ) in this region was estimated at 0.0136 mutations per base. Finally, human-chimpanzee divergence time ( $L$ ) was set at  $6 \times 10^6$  years. Most estimates of this time is between  $5 \times 10^6$

and  $6 \times 10^6$  years. We chose the upper one to be conservative. The estimated time to MRCA in years was then obtained, using the simple formula  $(2\hat{T}/D)*L$  as described previously (S11), at 37,281 years before present. The coalescence age of the entire Coriell panel was calculated in a similar manner. There are a total of 8136 mutations between the 178 chromosomes in the Coriell panel and the deduced MRCA sequence, which leads to an age estimate of 1,722,347 years. We note that owing to recombination, this estimated age is actually the average of multiple coalescence ages corresponding to multiple recombination blocks that coalesce independently.

The 95% confidence interval (CI) for the age of haplogroup D was estimated by an analytical approach that is an extension of a previously described method (S12). Let  $y_i$  denote the number of differences between the

MRCA and the  $i^{\text{th}}$  chromosome. The value of  $\hat{t}$  would be  $(\sum_{i=1}^n y_i)/n$ , where  $n$  is the number of chromosomes

sampled. The variance of  $\hat{t}$  is  $[\sum_{i=1}^n \text{var}(y_i) + 2\sum_{i<j} \text{cov}(y_i, y_j)]/n^2$ . If we assume an infinite-sites model, each  $y_i$  is

Poisson distributed with mean  $t$ . The  $\text{var}(y_i)$  is simply  $t$ , and the  $\text{cov}(y_i, y_j)$  is simply  $t - t_{ij}$ , where  $t_{ij}$  is the time of the most recent common ancestor of chromosome  $i$  and chromosome  $j$  (S11, S12). Therefore the

variance of our estimate is  $t/n + 2[\sum_{i<j} (t - t_{ij})]/n^2$ . There are  $n(n-1)/2$  terms in this sum, so this can be

written as  $t - 2[\sum_{i<j} (t_{ij})]/n^2$  or  $t - [(n-1)/n]\bar{t}_{ij}$ , where  $\bar{t}_{ij}$  is the average time to the most recent common

ancestor of a pair of chromosomes.  $\bar{t}_{ij}$  can be estimated as one-half the average pairwise differences between

the 94 chromosomes, calculated as  $(1/2)\sum_{k=1}^m \{2f_k(m - f_k)/[m(m-1)]\}$  or  $\sum_{k=1}^m \{f_k(m - f_k)/[m(m-1)]\}$ , where

$f_k$  is the count of the derived allele at the  $k^{\text{th}}$  polymorphic site and  $m$  is the total number of polymorphic sites. So

we can estimate the variance of  $\hat{t}$  by  $\hat{t} - [(n-1)/n]\sum_{k=1}^m \{f_k(m - f_k)/[m(m-1)]\}$ . For the 94 haplogroup D

chromosomes sampled in the 47 individuals, there are 34 SNP sites. Let  $N_x$  designate the number of sites where

the count of the derived allele is  $x$ . For our data,  $N_1 = 23$ ,  $N_2 = 2$ ,  $N_3 = 5$ ,  $N_4 = 1$ ,  $N_{15} = 2$ ,  $N_{17} = 1$ , and all

others  $N_x$  values are zero. Thus, based on our data, the estimate for the variance of  $\hat{t}$  is 0.094, and the estimate

for the standard error of  $\hat{t}$  is  $\sqrt{0.094} = 0.307$ . Assuming that the  $\hat{t}$  estimator is roughly normally distributed, the

95% CI of  $\hat{t}$  would be approximately 0.376 to 1.60. This corresponds, in units of years, a CI of 14175 to 60387

years before present. We note that this CI does not consider uncertainties in mutation rate. It also does not

consider uncertainties in the estimated human-chimpanzee divergence time, which can only be inferred from

fossil records and molecular data, and cannot be directly observed.



**References and Notes**

- S1. M. Stephens, N. J. Smith, P. Donnelly, *Am. J. Hum. Genet.* **68**, 978 (2001).
- S2. M. Stephens, P. Donnelly, *Am. J. Hum. Genet.* **73**, 1162 (2003).
- S3. J. Rozas, R. Rozas, *Bioinformatics* **15**, 174 (1999).
- S4. H. M. Cann *et al.*, *Science* **296**, 261 (2002).
- S5. B. S. Weir, C. C. Cockerham, *Evolution* **38**, 1358 (1984).
- S6. B. S. Weir, *Genetic Data Analysis* (Sinauer Associates, Sunderland, 1990).
- S7. R. R. Hudson, *Oxford Surv.Evol. Biol.* **7**, 1 (1990).
- S8. R. R. Hudson, *Bioinformatics* **18**, 337 (2002).
- S9. A. Kong *et al.*, *Nat. Genet.* **31**, 241 (2002).
- S10. C. Wiuf, J. Hein, *Genetics* **155**, 451 (2000).
- S11. R. Thomson, J. K. Pritchard, P. Shen, P. J. Oefner, M. W. Feldman, *Proc. Natl. Acad. Sci. U.S.A.* **97**, 7360 (2000).
- S12. H. Tang, D. O. Siegmund, P. Shen, P. J. Oefner, M. W. Feldman, *Genetics* **161**, 447 (2002).











Table with 4 columns: ID, and three columns of letters (likely 'G', 'A', 'C'). IDs range from 31708-31712 down to 38538. Some IDs are highlighted in red, such as 37995 and 38066.

Main grid of 31 rows and 38 columns. Each cell contains a character (G, A, C) or a number (e.g., 77s, 81s, 87s, 91s). The grid is mostly composed of 'G', 'A', and 'C', with occasional numbers representing specific values. The layout is consistent across the rows, with the same sequence of characters and numbers appearing in each row.

43807
43806
43805
43804
43803
43802
43801
43800
43799
43798
43797
43796
43795
43794
43793
43792
43791
43790
43789
43788
43787
43786
43785
43784
43783
43782
43781
43780
43779
43778
43777
43776
43775
43774
43773
43772
43771
43770
43769
43768
43767
43766
43765
43764
43763
43762
43761
43760
43759
43758
43757
43756
43755
43754
43753
43752
43751
43750
43749
43748
43747
43746
43745
43744
43743
43742
43741
43740
43739
43738
43737
43736
43735
43734
43733
43732
43731
43730
43729
43728
43727
43726
43725
43724
43723
43722
43721
43720
43719
43718
43717
43716
43715
43714
43713
43712
43711
43710
43709
43708
43707
43706
43705
43704
43703
43702
43701
43700
43699
43698
43697
43696
43695
43694
43693
43692
43691
43690
43689
43688
43687
43686
43685
43684
43683
43682
43681
43680
43679
43678
43677
43676
43675
43674
43673
43672
43671
43670
43669
43668
43667
43666
43665
43664
43663
43662
43661
43660
43659
43658
43657
43656
43655
43654
43653
43652
43651
43650
43649
43648
43647
43646
43645
43644
43643
43642
43641
43640
43639
43638
43637
43636
43635
43634
43633
43632
43631
43630
43629
43628
43627
43626
43625
43624
43623
43622
43621
43620
43619
43618
43617
43616
43615
43614
43613
43612
43611
43610
43609
43608
43607
43606
43605
43604
43603
43602
43601
43600
43599
43598
43597
43596
43595
43594
43593
43592
43591
43590
43589
43588
43587
43586
43585
43584
43583
43582
43581
43580
43579
43578
43577
43576
43575
43574
43573
43572
43571
43570
43569
43568
43567
43566
43565
43564
43563
43562
43561
43560
43559
43558
43557
43556
43555
43554
43553
43552
43551
43550
43549
43548
43547
43546
43545
43544
43543
43542
43541
43540
43539
43538
43537
43536
43535
43534
43533
43532
43531
43530
43529
43528
43527
43526
43525
43524
43523
43522
43521
43520
43519
43518
43517
43516
43515
43514
43513
43512
43511
43510
43509
43508
43507
43506
43505
43504
43503
43502
43501
43500
43499
43498
43497
43496
43495
43494
43493
43492
43491
43490
43489
43488
43487
43486
43485
43484
43483
43482
43481
43480
43479
43478
43477
43476
43475
43474
43473
43472
43471
43470
43469
43468
43467
43466
43465
43464
43463
43462
43461
43460
43459
43458
43457
43456
43455
43454
43453
43452
43451
43450
43449
43448
43447
43446
43445
43444
43443
43442
43441
43440
43439
43438
43437
43436
43435
43434
43433
43432
43431
43430
43429
43428
43427
43426
43425
43424
43423
43422
43421
43420
43419
43418
43417
43416
43415
43414
43413
43412
43411
43410
43409
43408
43407
43406
43405
43404
43403
43402
43401
43400
43399
43398
43397
43396
43395
43394
43393
43392
43391
43390
43389
43388
43387
43386
43385
43384
43383
43382
43381
43380
43379
43378
43377
43376
43375
43374
43373
43372
43371
43370
43369
43368
43367
43366
43365
43364
43363
43362
43361
43360
43359
43358
43357
43356
43355
43354
43353
43352
43351
43350
43349
43348
43347
43346
43345
43344
43343
43342
43341
43340
43339
43338
43337
43336
43335
43334
43333
43332
43331
43330
43329
43328
43327
43326
43325
43324
43323
43322
43321
43320
43319
43318
43317
43316
43315
43314
43313
43312
43311
43310
43309
43308
43307
43306
43305
43304
43303
43302
43301
43300
43299
43298
43297
43296
43295
43294
43293
43292
43291
43290
43289
43288
43287
43286
43285
43284
43283
43282
43281
43280
43279
43278
43277
43276
43275
43274
43273
43272
43271
43270
43269
43268
43267
43266
43265
43264
43263
43262
43261
43260
43259
43258
43257
43256
43255
43254
43253
43252
43251
43250
43249
43248
43247
43246
43245
43244
43243
43242
43241
43240
43239
43238
43237
43236
43235
43234
43233
43232
43231
43230
43229
43228
43227
43226
43225
43224
43223
43222
43221
43220
43219
43218
43217
43216
43215
43214
43213
43212
43211
43210
43209
43208
43207
43206
43205
43204
43203
43202
43201
43200
43199
43198
43197
43196
43195
43194
43193
43192
43191
43190
43189
43188
43187
43186
43185
43184
43183
43182
43181
43180
43179
43178
43177
43176
43175
43174
43173
43172
43171
43170
43169
43168
43167
43166
43165
43164
43163
43162
43161
43160
43159
43158
43157
43156
43155
43154
43153
43152
43151
43150
43149
43148
43147
43146
43145
43144
43143
43142
43141
43140
43139
43138
43137
43136
43135
43134
43133
43132
43131
43130
43129
43128
43127
43126
43125
43124
43123
43122
43121
43120
43119
43118
43117
43116
43115
43114
43113
43112
43111
43110
43109
43108
43107
43106
43105
43104
43103
43102
43101
43100
43099
43098
43097
43096
43095
43094
43093
43092
43091
43090
43089
43088
43087
43086
43085
43084
43083
43082
43081
43080
43079
43078
43077
43076
43075
43074
43073
43072
43071
43070
43069
43068
43067
43066
43065
43064
43063
43062
43061
43060
43059
43058
43057
43056
43055
43054
43053
43052
43051
43050
43049
43048
43047
43046
43045
43044
43043
43042
43041
43040
43039
43038
43037
43036
43035
43034
43033
43032
43031
43030
43029
43028
43027
43026
43025
43024
43023
43022
43021
43020
43019
43018
43017
43016
43015
43014
43013
43012
43011
43010
43009
43008
43007
43006
43005
43004
43003
43002
43001
43000

G	T	C	A	A	C	A	G	A	T	A	T	T	T	C	A	C	G	T	T	C	G	T	T	C	G	G	T	T	A	A	C	C	G	T	T	C	G
---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---



