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SIMULATING SUPER EARTH ATMOSPHERES IN THE LABORATORY

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Abstract

Several space missions, like JWST, TESS and the very recently proposed ARIEL, or ground based experiments, as SPHERE and GPI, have been proposed to measure the atmospheric transmission, reflection and emission spectra of extrasolar planets. The planet atmosphere characteristics and possible biosignatures will be inferred by studying planetary spectra in order to identify the emission/absorption lines/bands from atmospheric molecules such as water (H_2O), carbon monoxide (CO), methane (CH_4), ammonia (NH_3) etc. In particular, it is important to know in detail the optical characteristics of gases in the typical physical conditions of the planetary atmospheres and how these characteristics could be affected by radiation driven photochemical and bio-chemical reaction. The main aim of the project "Atmosphere in a Test Tube" is to provide insights on exoplanet atmosphere modification due to biological intervention. This can be achieved simulating planetary atmosphere at different pressure and temperature conditions under the effects of radiation sources, used as proxies of different bands of the stellar emission. We are tackling the characterization of extrasolar planet atmospheres by mean of innovative laboratory experiments described in this paper. The experiments are intended to reproduce the conditions on warm Earths and super Earths hosted by low mass M dwarfs primaries with the aim to understand if a cyanobacteria population hosted on a Earth-like planet orbiting an M0 star is able to maintain its photosynthetic activity and produce traceable signatures.

1 Introduction

The bonanza of extrasolar planets discovered so far (over 1786 planets, Schneider et al. 2011) unveils that most of these new planets are very different from those of our Solar System. New worlds are hot gaseous giants (hot jupiters, hot neptunes), giant planets (jupiters and saturns) and smaller rocky worlds as super earths and earths. Earths and Super earths are rocky exoplanets with mass ranging between 1 and 10 M⊕ (Valencia et al., 2007). While the lower mass limit is obvious for historical reasons the upper limit is somewhat arbitrary. It is due to the physical argument that above ~10 M⊕, planets can retain Hydrogen and Helium in their atmospheres (Ida & Lin 2004). The Earths have radius $R_P \leq 1.25$ R⊕ while super earths have radii ranging between the interval [1.25, 2.0] R⊕ (Borucki et al., 2011; Batalha et al., 2013).

Since the discovery of GJ876d in the 2005 (Rivera et al. 2005) the existence of a set of super earths has been confirmed up to now (The Extra Solar Planets Encyclopaedia[1], Schneider et al. 2011). On the other hand there are 2,013 KOI (Kepler Objects of Interest) with earth and super earth size in the NASA Exoplanets Archive[2]. The warm super earth orbiting the M star GJ 1214b (Charbonneau et al., 2009) has been the first super earth to be observed spectroscopically (Bean et al., 2010). For the time being only two techniques can be used in order to probe the atmospheres of a planet: direct imaging and transits. The direct detection of a planet is still very challenging due to its high contrast ratio with respect to the host star, which can reach values of 10^{-10} for an earth-like planet (see e. g. Lafreniére et al., 2007, Chauvin et al., 2010, Biller et al., 2013). Transits give important diagnostic on the probable modification of the atmosphere due to the history of the planet allowing transmission and emission spectroscopy. Transmission spectroscopy, possible only when the planet transits its host star along the line of sight, allows to infer the main opacity sources present in the high atmosphere of the planet (Brown et al. 2001, Tinetti et al. 2007). Complementary, emission spectroscopy (Charbonneau et al. 2005), observing the day hemisphere of the planet and exploiting its occultation during the secondary transit, gives evidence on the thermal structure of the planetary atmosphere and the emission/reflection properties of the planetary surface. In order to maximize the finding of habitable planets with transit search, a lot of surveys have

[1] http://exoplanet.eu
[2] http://exoplanetarchive.ipac.caltech.edu/index.html

been dedicated to search for earth and super earths size planets around M stars (e.g. Nutzman & Charbonneau, 2008, Sozzetti et al., 2008). Due to a more favorable ratio between the radii, some small rocky companions have been discovered in the habitable zone (HZ) of these red and cold stars. The study of super earths in the HZ of stars cooler than the Sun will challenge the paradigm of the Earth-twin orbiting a Sun-twin as the only possible cradle of life (Segura et al., 2005). In the past, starting by Huang (1959, 1960) and Dole (1964), astronomer have considered stars very different by the Sun not suitable for biology. The stars with larger masses because evolve too fast, and those with smaller masses because their luminosity is too faint. In the latter case (the M star case) the HZs are so close to the star that the planet results tidally locked to the host star (e.g. Kasting et al 1993) keeping always the same hemisphere facing the star. Haberle et al. (1996) with a one dimensional simple model and successively Joshi et al (1997) with a more complex 3-D model showed that the atmospheric heat transport could prevent the freeze out of the night hemisphere of the planet. All these considerations coupled with the realization that 75% or more of the stars are M type dwarfs (Henry, 2004) make them appealing stars to search for life.

On Earth, due to the more efficient use of energy, the evolutionary selection favorited the photochemistry instead of chemolithotrophic reaction as energy source for growth (Wolstencroft and Raven, 2003, Kiang et al., 2007a). On Earth-like planets orbiting other stars could happen the same at the correspondent evolutionary state also if the means of energy transformation although redox reactions are essential need not necessarily involve the detailed mechanisms used on Earth..

The detection of photosynthesis on a super Earth orbiting a star different by a solar twin is an attractive possibility for the remote sensing of life. In fact photosynthesis will lead to an accumulation of free O_2 in the planet atmosphere detectable via spectroscopic detection of O_3 produced by O_2 in photochemical reactions. The presence of ozone in the NIR (band at 10 μm) could be used as biosignature (Angel et al., 1986; Leger et al., 1993) also if cautions should be taken into account for false positive created by photochemical reaction in high atmosphere (e.g. Selsis et al., 2002; Kiang et al, 2007a; Kiang et al., 2007b). A detectable concentration of O_2 and/or O_3 in combination with reduced gases like CH_4 is a more robust signature of biologic activity (Lammer et al., 2009). Signatures of the presence of photosynthesis on the surface of a planet are mainly the spectral reflectance characteristics of

organisms due to the wavelength range in which they absorb the light. On Earth there are two spectral signatures. The former is the "green bump" due to the absorption of light by green plants in the range between 400 and 700 nm. The later, more significant, is the "red edge" due to the high reflectance at 700 -850 nm. The red edge causes a high contrast between the visible and the NIR in the spectra (Kiang et al., 2007a; Kiang et al., 2007b and references therein). Theoretical studies (Wolstencroft and Raven, 2002) show that the photosynthetic production of O_2 should be greater on cloud-free planets orbiting in the inner limit of the HZ of warmer solar-like stars than on those with cooler parent stars. The lower O_2 production in habitable planets orbiting colder stars is due to the poor match between the spectral energy distribution of the parent star and the properties of terrestrial pigments, but M star planets could be more productive if useful photons extend to 1.1μm for anoxygenic photosynthesis (Tinetti et al., 2006; Kiang et al., 2007b).

This introduction could not be exhaustive of the multitude of theoretical works done in these years on this topic. A lot of hypotheses have been done on role of photosynthesis in the habitability of extrasolar planets and on its efficacy under the radiation of stars of different spectral types with particular efforts on M stars.

In this framework it is interesting to explore in the laboratory how the irradiation of a M star modifies (if it does) the oxygen production of photosynthetic bacteria.

This could be done putting photosynthetic bacteria inside steel reaction cells where it is possible to maintain a mixture of gases simulating a planetary atmosphere as done in our experiment described in this paper. The reaction cells are positioned inside an environmental simulator that can control the temperature and the pressure.. In particular in section 2 we explain with what criteria we selected the environmental conditions and describe the material used in the lab experiment. The experiment, its status and the preliminary results on commissioning of the environmental simulator are described in section 3 and 4 while conclusion are given in section 5.

2 Criteria and material

The main target of the experiment is to study the modification to the secondary atmosphere of an habitable rocky planet orbiting a M star due to the presence of photosynthetic bacteria on its surface. This scenario constraints the environmental

parameters like pressure, temperature, planetary atmosphere contents and irradiation pattern that we want simulate in laboratory.

2.1 M stars and habitable planets

In the case of a planet in the HZ of an M star the situation is complicate because it is tidally locked with the star. The day side of the planet, always irradiated by the star, is hotter than the night side. If there is not any heat exchange between the two hemispheres, the night side could be so cold that all the atmosphere constituents will condense out on the surface following what is called the atmospheric collapse. Actually, the atmospheric motions reduce the day/night temperature gradient by transporting heat. Horizontal advection is strong enough to prevent the atmospheric collapse of the night hemisphere (Joshi et al., 1997, Joshi, 2003). Haberle et al. (1996) used a simple energy balance model to show that a pure CO_2 atmosphere having a value of p_0 of about 15 kPa is sufficient dense to warm up the night hemisphere of such a planet over the freezing point of CO_2. Later, with a more refined and complete 3-D model Joshi et al. (1997) show that an Earth – like planet in the HZ of an M star can support an atmosphere and if the atmosphere is as thick as 100–200 kPa it may also support liquid water on a large fraction of the surface. In fact, later models allowed completely ocean- covered water worlds (Joshi, 2003).

The atmospheric level of CO_2 and its variations due to changes on plate tectonic and volcanism are important factors for the planetary climate and temperature variation (Kasting and Catling, 2003).

Moreover the definition of the temperature range in which the green house effect is effective to sustain the superficial temperature of the planet as high as it can maintain the water in liquid state, depends by the host star, the orbital distance of the planet and the planet itself (Kasting et al., 1993 and Selsis et al., 2007). Using both the "Mars" and "Venus" criterion (Selsis et al. 2007) this range is between 216 K (first condensation of CO_2) and 373 K (water loss limit). In any case, in order to avoid difficulties in this first phase of the experiment and expensive laboratory infrastructure, we consider Earths and super Earths well inside the HZ of the host star. This make the temperature ranges between 273 K and 288 K (the current temperature value on Earth).

M dwarfs are faint hydrogen burning stars with low photospheric temperature which spectra are dominated by molecular bands like, in the visible, TiO and VO. Even the

most luminous M star (spectral type M0) is over 10 times fainter than the Sun. The steep dependence of stellar luminosity on mass results in M stars spanning a factor of about 100 in bolometric luminosity. Another characteristic of these stars is that their effective temperatures range from 3,900 K to 2,400 K, passing from M0 to M9 (see Table 1). In the visible these stars are something like 60,000 times fainter than the Sun (absolute visible magnitude change from 9 to 20). These characteristics are also reflected in the UV zone of the spectrum. The UV emission has two components that are both dependent by the age of star. Photometric component, depending by stellar surface temperature has a faint dependence by the stellar age (Gough, 1981). This component is weak for stars of spectral type later than G. Instead, the second component, due to the active chromosphere that drains energy by the dynamo mechanism in the stellar interiors, is the main cause for UV radiation emission of M stars. This component depends by the stellar rotation and this is though to decline with age (e.g. Ayers et al., 1997). When they are young they are more magnetically active and exhibit large flares and strong X-ray and UV emission. But due to their typical masses once they are burning hydrogen the HZ can be stable and in place for 100 Gyr (Tarter et al., 2007) and in part of this time the star could evolve in a stiller state. In order to maximize the output of the experiment we decide to simulate the irradiation of an M0 V star.

Table 1: M star characteristics compared with those of the Sun

Spectral Type	M_V	V-K	Teff (K)	$M/M_\odot$	$L/L_\odot$
M0V	9.0	3.5	3,900	0.50	0.0600
M3V	11.7	4.9	3,600	0.29	0.0300
M6V	16.6	7.2	3,000	0.10	0.0050
M9V	19.4	8.9	2,400	0.08	0.0002
G2V (Sun)	4.8	1.5	5,800	1.00	1.0000

2.2 Photosynthetic bacteria

Photosynthetic bacteria are the key to understand how a hot, inhospitable planet evolves in a beloved and safe green planet. In Kiang et al. (2007a) it is possible to find a review of the basic processes at work with photosynthetic organisms and a

quite complete directory of photosynthetic bacteria, algae and plants on Earth together with their habitats and main characteristics.

In phototrophs Clorophylls (Chls) both harvest the light and transduce it into chemical energy; there are four chemically different chlorophylls termed Chls a, b, c, and d in the order of their discovery. All four pigments are present in light harvesting complexes, though until recently only Chl a was thought to be indispensable for energy transduction in the photosystem reaction centers. This paradigm was challenged with the discover of the red-shifted Chl d, in the cyanobacterium *Acaryochloris marina*, where it constitute sup to 99% of all Chl. Other photosynthetic pigments are the phycobiliproteins present in the light- harvesting complex of cyanobacteria with absorption spectra ranging from 480 nm to 650 nm.

The photosynthetically active radiation (PAR) represents the active radiation boundaries between which photosynthesis can be able to operate. For plants it is enclosed between 0.40-0.70 μm. Other living organisms, such as green bacteria, purple bacteria and heliobacteria, can exploit solar light in slightly extended spectral regions or in ecological niches, such as the near-infrared. These bacteria live in environments such as the bottom of stagnant ponds, sediment and ocean depths. Because of their pigments, they form colorful mats of green, red and purple. Purple bacteria have absorbance peak in the 1.013-1.025 μm range which use bacteriochlorophyll b like *Blastochloris viridi* or *Rhodopseudomonas viridis* (that absorbs at 0.96 μm) and other bacteriochlorophylls in the range 0.7-0.9 μm (Scheer, 2003). They don't use water as H donor, and then don't release oxygen as byproduct. Other bacteria that can photosynthetize in the IR are the Chlorobium genus, with an absorbance peak in their pigments at 0.84 μm or *Rhodospirillum rubrum* and *Rhodospirillum capsulata* both absorbing at 0.87 μm (Heath et al., 1999). The bounds of the shorter PAR band could be set at 0.40–0.73 μm to acknowledge the ability of *Acaryochloris marina* to utilize the longer wavelengths (Kiang et al., 2007b; Chen et al., 2005).

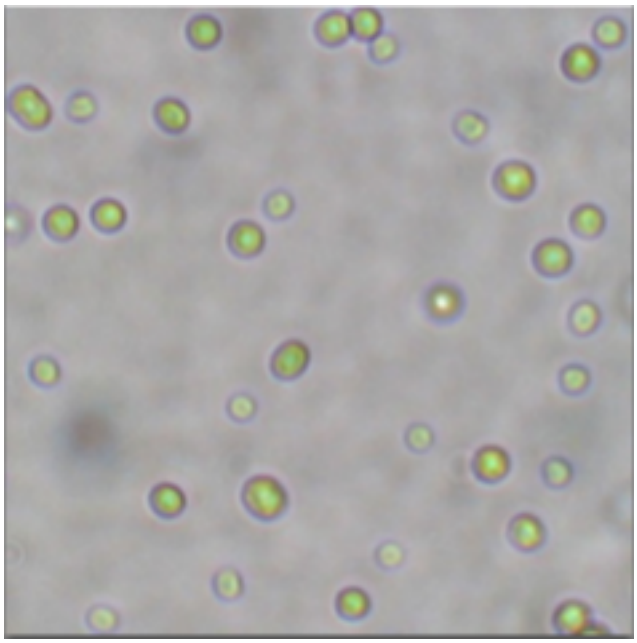

Figure 1: Image of *Acaryochloris marina*

The cyanobacterium *Acharyochloris marina* (see Figure 1) produces Chlorophyll d and it utilizes far-red light, at 710 nm wavelength (Figure 2) and in condition of faint irradiation at shorter wavelength it may be adapted to receive light at longer wavelength (Kiang et al., 2007a).

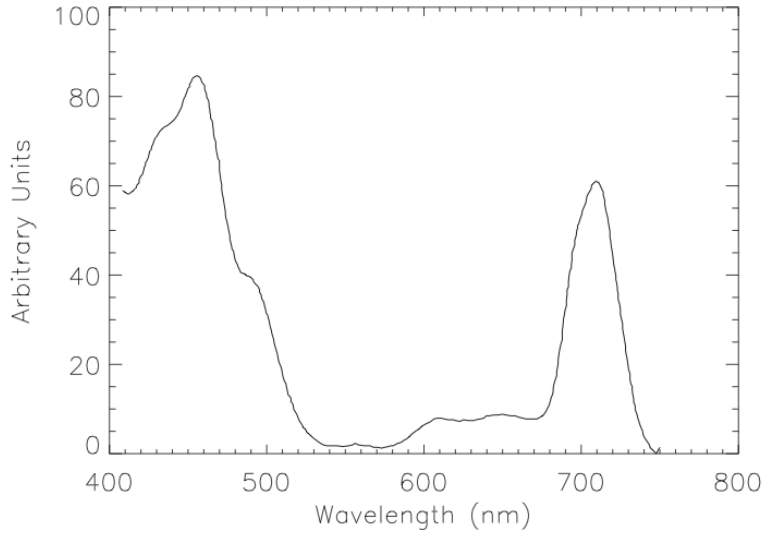

Figure 2: Absorption spectra of Acaryochloris Marina. The spectrum show the features that are typical for Chl d-dominate organisms (adapted by Mohr et al., 2010).

It is worth to mention other two kinds of photosynthetic bacteria: *Chroococcidiopsis spp.* and *Halomicronema hongdechloris.*

The former is a desiccation-tolerant lithic cyanobacterium (see Figure 3).

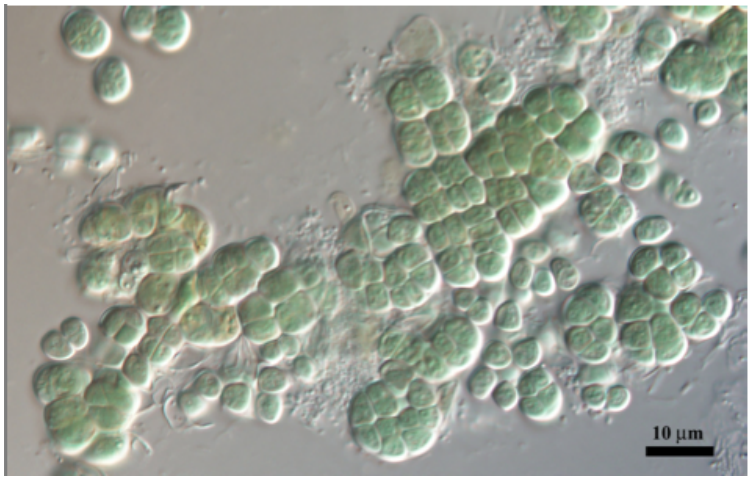

Figure 3: Image of *Chroococcidiopsis*

According to Billi et al. (2001), O_2 production (evolution–uptake) was recorded to be around 40 fmol/cell/h in normal growth condition (20 µmol of photons m^{-2} s^{-1}). Taking as approximation a typical bacterial cell mass around 1pg this would lead to a production of about 40 mmol/g/h. *Chroococcidiopsis* normal growth condition are 20 µmol of photons m^{-2} s^{-1}. The theoretical photosynthetic production limit is 0.1×10^{-6} mol of photons m^{-2} s^{-1}.

The latter is a filamentous cyanobacterium that contains chlorophyll f (shown in Figure 4).

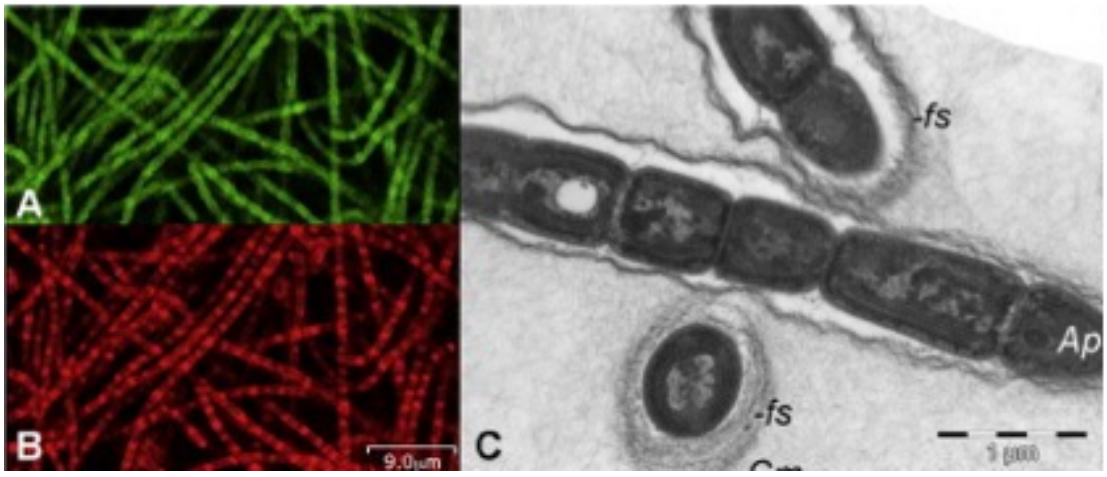

Figure 4: Image of *Halomicronema hongdechloris*

Photosynthetic organisms can produce gases, like O_2 (or O_3 from its photolysis), and nitric oxides like N_2O, NO_X, CH_3Cl or COS from the breakdown of organic matter. All these molecules can modify exoplanet's atmospheres and can be detected remotely from Earth. O_2 can be produced even abiotically through photolysis and the effects of carbon burial and hydrogen escape (Selsis et al. 2002; Kiang et al 2007a). Though, its simultaneous presence with other reduced gases can be explained only with biotic processes that maintain chemical disequilibrium (Kiang et al., 2007b).

Detectability of photosynthetic processes depends on biotic productivity, which depends on several factors, like availability of resources (water, light, minerals, electron donors, nutrients etc..). During photosynthesis, light impacts on photo-receptive organisms that split water molecules and produce proton gradients and energy useful for the cells (Kiang et al., 2007a). It is crucial to understand the ability of organisms to use photosynthesis on M star planets. PAR on M star planets can be lower than the average terrestrial value even by an order of magnitude (Heath et al., 1999). Nevertheless this could not represent a problem because several marine organisms on Earth evolved to use only 5×10^{-4} times the average flux received at the Earth's surface, like sulfur bacteria with peculiar light-harvesting antenna, known as chlorosome, that permit to use only small fractions of light intensities (McKay, 2000). In these regions radiation is dominated by red or IR radiation.

For our purpose we need a bacterium that is resistant to harsh conditions (e.g. an extremophile) and capable to photosynthetize in the NIR as explained in the previous paragraph. The choice has been fallen on the cyanobacterium *Acharyochloris marina.* The use of this kind of bacteria defines the working wavelength range of the experiment to be in between 400 nm and 900 nm (see Figure 2).

Studies on O_2 productivity of these bacteria show that it is directly dependent on bacteria growth. The increase in number of bacteria often shows a phase in which the specific growth rate starts at a value of zero and then accelerates to a maximal value in a certain period of time, resulting in a lag time. In addition, growth curves contain a final phase in which the rate decreases and finally reaches zero, so that an asymptote is reached. When the growth curve is defined as the logarithm of the number of organisms plotted against time, these growth rate changes result in a sigmoidal curve, with a lag phase just after t=0 followed by an exponential phase and then by a

stationary phase. In order to evaluate the production of O_2 by these bacteria we use as growth model, the Gompertz model fully described in Zwietering et al. (1990), that take into account the bacteria rate of growth, the lag time and the maximum value that could be reached in growth. These three parameters depend by the bacterial population and are derived by measuring them. Behrendt et al. (2012) characterized in biochemical and biophysical way the *Acharyochloris marina* obtaining also the required parameters. The O_2 production is evaluated multiplying the rate of growth for the maximum gross photosyntesys found by Behrendt et al, (2012) that is 1,272 µmol O_2/mg Chld/h in the NIR (700≤ λ≤730 nm) and 1,128 µmol O_2/mg Chld/h in the VIR range (300≤λ<700 nm). The result is shown in .

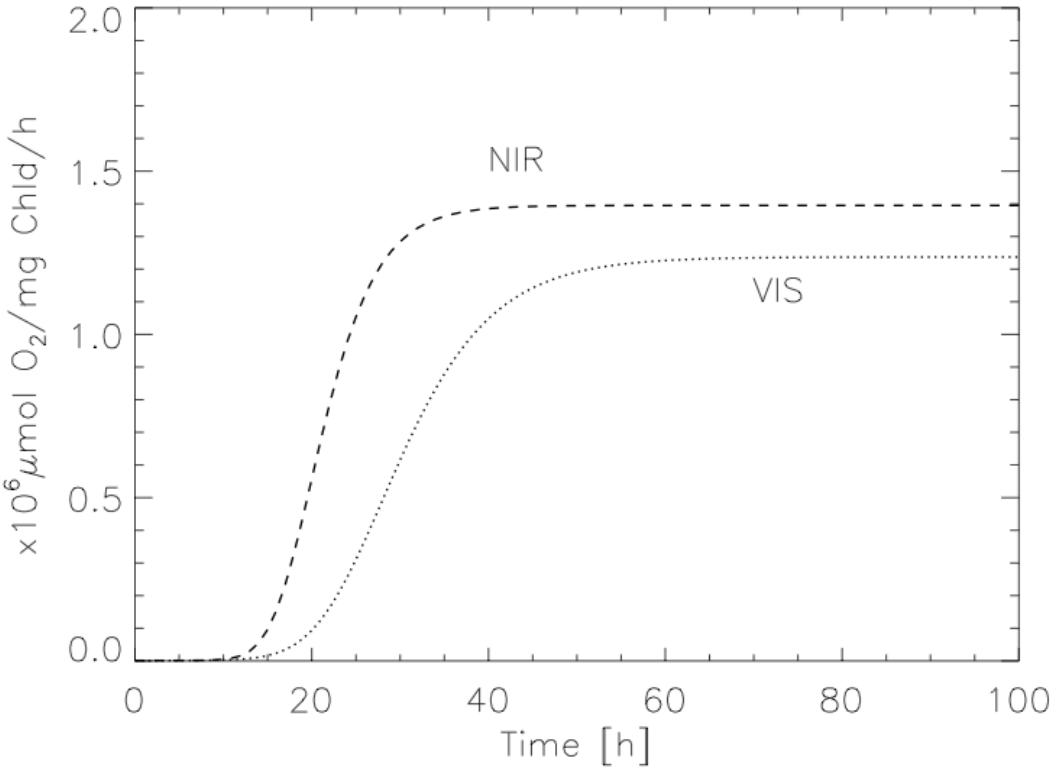

Figure 5: The gross O_2 production of *Acharyochloris marina.* In the Gompertz model the maximum is represented by the plateau at the maximum rate.

2.3 Summary

To summarize (Table 2) in our experiment we measure the oxygen production of the *Acharyochloris Marina* considering a habitat very similar to that on Earth, but illuminated by a M0V star.

Table 2: Summary of values of quantities to be simulated into the laboratory

Planet	Planetary Mass (M⊕)	$1 \leq M_P \leq 10$
	Planetary Radius (R⊕)	$1 \leq R_P \leq 2$
	Surface Gravity	$0.99 \leq \log g \leq 1.99$
	Surface Temperature (K)	$273 \leq T \leq 288$
	Surface Pressure (kPa)	$10^2 \leq P \leq 2 \times 10^2$
	Atmospheric Chemical Composition	CO_2, N_2, CO, H_2O, O_2
Star	Sp STAR	M0V
	T_{eff}	3900
Sample	Wavelength range (nm)	$400 \leq \lambda \leq 900$
	Bacterium	*Acharyochloris marina*

Once obtained this result we will be able to test the oxygen production by this cyanobacterium under other, more challenging, environmental conditions using our simulator.

3 The experiment plan and set up

A lot of theoretical works have been done on this topic and all of them suggest theory to be verified with laboratory experiments. The amount of work necessary to verify most of the suggested hypothesis is a huge and challenging one. But, because also a long walk begins with a single step, we start with this first approach to the experimental verification on the efficiency of production of O_2 by photosynthetic organisms illuminated by a cold black body. The main pipeline of the experiment consists of three phases. The first phase is a fiduciary experiment in which we will measure the photosynthetic bacteria O_2 production in terrestrial conditions using a solar simulator. This first phase is necessary to understand if the set up of the experiment allows us to perform all the measurements we need. On the contrary it let us to comprehend what and why there is something wrong. Once performed this first phase it will be possible to move to the next step and change the irradiation source. For this second phase we will use an illuminator that simulate the M0 star irradiation.

Finally, we will change the gas mixture (the atmosphere) inside the cells (the third phase), keeping M0 star simulator source as in the previous step, to perform the experiment in different environmental condition. At this phase the composition of the mixture of gas to be used will come by theoretical simulation of super earth atmospheres.

The set up of the experiment consists in an environmental simulator that can host up to six reaction cells in which the biological sample will be placed and irradiated. In the following of this paragraph the set up of the experiment is outlined.

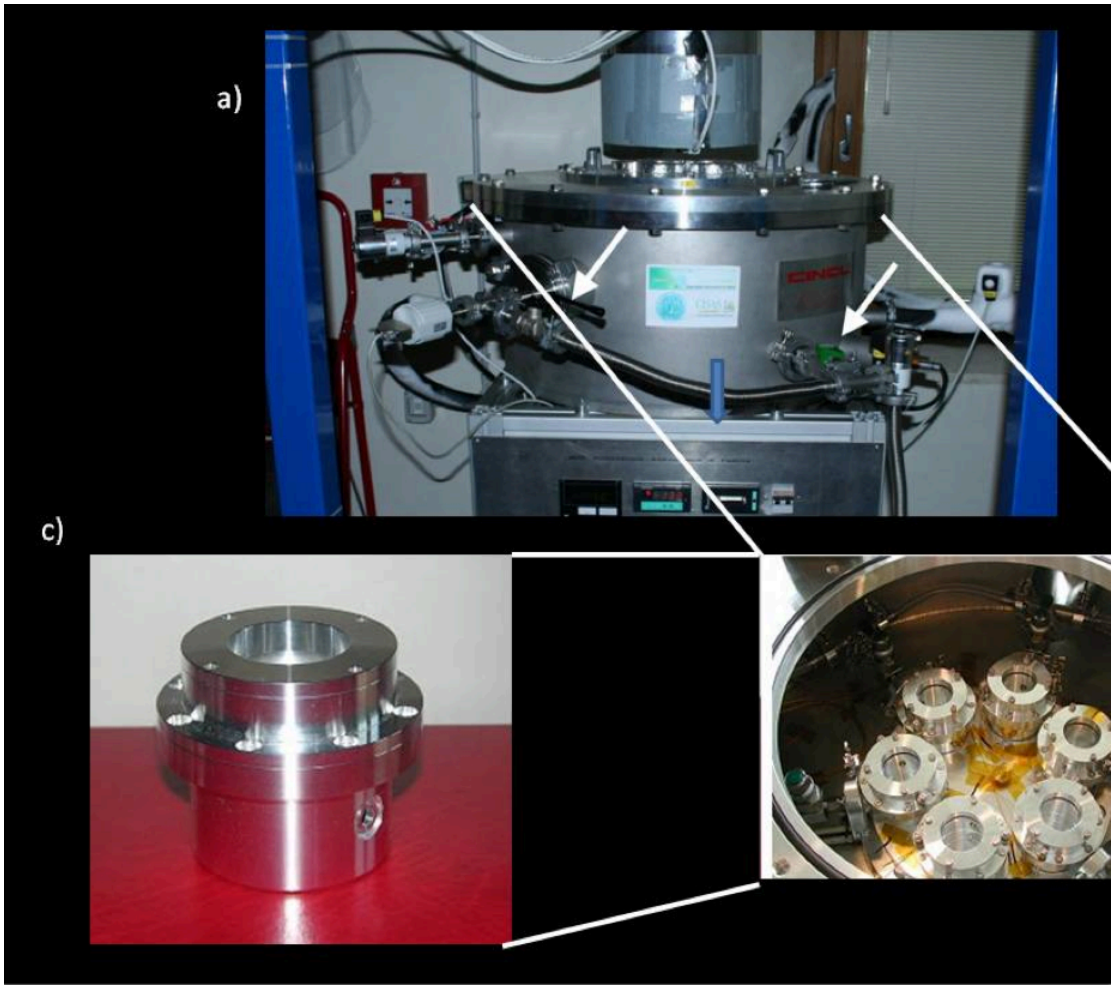

Figure 6: In the picture can be seen the instrumental complex in toto (a) and the particular of one of the six cells located inside it (b, c). The LISA SAM has been built by the Astronomy Department of University of Padua).

The environmental simulator LISA (Laboratorio Italiano Simulazione Ambienti) (Galletta et al., 2006), shown in Figure 6, was designed and built with the technical support of the Center of Study and Space Activity of Padua (CISAS) in order to perform survival experiments of terrestrial bacteria strains. It has been exploited in the

past to study the limit of bacterial life on the planet Mars (Galletta et al., 2010). It allows up to six simultaneous experiments in corresponding steel reaction cells of 250 cc, each containing a bacterial or soil sample. The cover of the simulator has a glass window (that could be obscurated) to allow the illumination of the inner part by an external light source. LISA environmental chambers can reproduce the conditions of many rocky planet locations near the surface. Temperature inside the simulator could be modified during the experiment in the range between +100 ◦C and -100 ◦C whenever we need to simulate a variation between day and night or summer and winter. A pump system modifies the pressure inside the chamber in order to keep void during cooling. The temperature in the chamber could be raised acting on resistances or lowered by means a closed circuit with liquid nitrogen or glycol depending by the lower limit we like to reach. The reaction cells containing the biological samples are cooled by contact with a large aluminum dish that is at the top of a small reservoir of about 2000 cm^3 containing the refrigerator liquid fed by an external receptacle. A PT100 temperature sensor connected with an electric valve opens or close the refrigerator liquid flux. If needed, an electric resistance between the dish and the reservoir is open, in a feedback circuit that keeps the vessels temperature fixed within ±0.5◦C.

Atmosphere with different compositions could be pumped inside the reaction cells.

Cells are connected with the outer part by pipes with mechanical filters to let the gas to course and avoid biological material to go inside the cryostat chamber.

The reaction cells have been slightly modified from the original design mainly because we plan to measure the oxygen content and its variation by means the diode laser spectroscopy based on fluorescence. This technique has had a large development in both monitoring of gases in food packaging and in the medical field. The instrument tunes the wavelength of the laser across a wavelength interval where the probed gas has an absorption line and the light is sent along a path through the gas. A detector records the intensity of the light that passed through the gas as a function of time. The measurement is based on the Beer - Lambert law (Beer, 1852):

$$I(\lambda) = I_0(\lambda) \cdot e^{-\sigma(\lambda) \cdot c \cdot L}$$

Where c is the concentration of the gas, L the length of the light path and σ is the molar absorptivity. If the path length is given in centimeters, and the concentration in

molecules per cm^3, σ must have the unit cm^2 for the exponent to be dimensionless. To extract the concentration from the measured intensity decrease, both σ and L must obviously be known. L is fixed by the reaction cell dimension while σ could, in principle, be known from databases such as HITRAN[3] (Rothman et al., 2010), and indeed sometimes this is the way to proceed. However, it is probably more common to perform calibration measurements with known gas concentrations. The main advantages of this approach is that it is then not necessary to know exactly which absorption line of the gas is being monitored, and that the whole signal processing chain is automatically considered in the calibration (Werle, 1998). For this the reaction cell have been redrawn in order to allow laser light to pass through the gas.

Once the reaction cells will be filled with the gas at the required pression (it depends by the phase of the experiment) and the sample will be accommodated inside them, the sample will be lightened. During the first phase we will use a solar simulator, while in the second phase we will use an M0 star light simulator.

The power of the illuminators has been chosen in order to allow bacterial photosynthesis and taking into account the planet to star separation for which HZ is defined. Void will be kept inside the chamber around the aluminum cells to avoid frosting damages. Before and during sample irradiation will be taken measurements of gas composition, expecting a peak of gas concentration variations at the end of bacteria metabolic and photosynthetic process. M0 star spectra have strong absorbing lines and bands, so it is difficult to reproduce exactly it as they are not good black body emitters. Once this illuminator will be ready, measurements will be taken as in the fiduciary step using the new lamp instead the Sun simulator one.

4 Commissioning results and Status

The experiment is under development and it is evolving on more than one level, each of which is an essential part of it.

In the laboratoratory we tested the performance of the environmental chamber. The thermal efficiency of LISA has been evaluated cooling the dish up to -80C and evaluating the temperature inside the reaction cell obtaining the same results described by Galletta et al. 2007 and shown in Figure 7.

[3] http://www.cfa.harvard.edu/hitran/

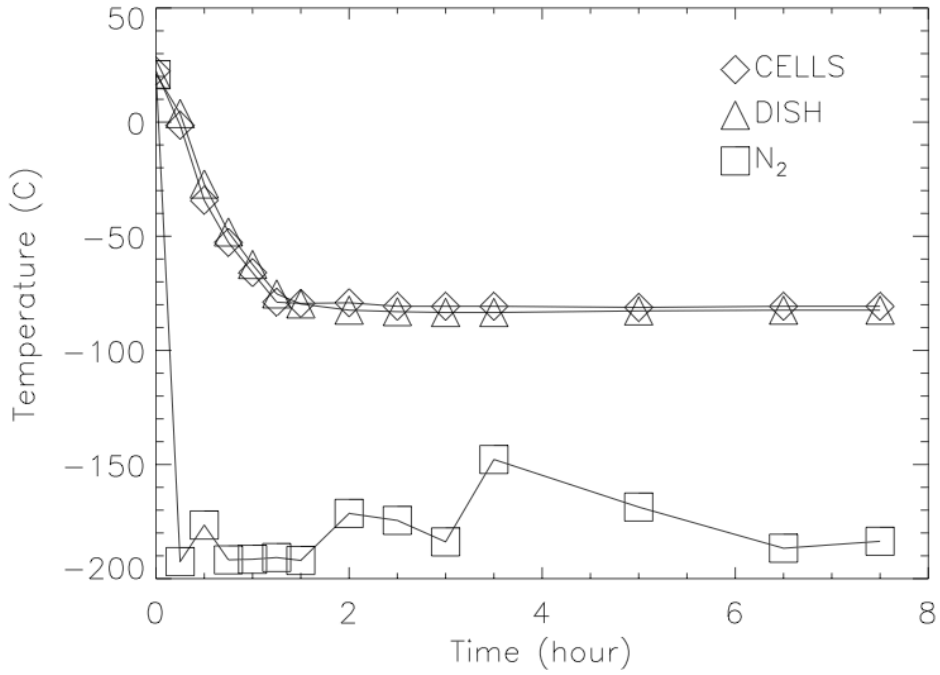

Figure 7: Decreasing of temperature inside the Environmental Simulator

Moreover we tested also the reaction cells for leakage with a Helium leak detector in order to know how they are able to confine the gasses. The main parameters to keep under control is the time constant τ calculated as $\tau = t/\ln[k/(P_0-P(t))]$ where P(t) is the pressure at time t, P_0 is the zero pressure, k is the difference between P_0 and P(t=0) that is the first measure kept close to the vacuum. The loss rate Q_A, calculated as $P_0/\tau V$ where V is the cell volume, that is 0.25 l. Each cell has an average time constant of about 2000 hr and a loss rate Q_A=13 Pa/h.

As cells, all the hardware described in the previous section was refurbished in order to up to date most of the electronics and control system. So we performed an experiment in order to test all the hardware in working condition. This "commissioning" experiment was designed to determine how short-term Martian atmospheric conditions affect vegetative cells survivability. Martian conditions were simulated using LISA facility. Cellular suspension (layered on sterile coverslip dehydrated under sterile air flux) were exposed respectively to "winter" and "summer" Mars-like conditions. The selected winter and summer Martian conditions are reported in Table 3. In both cases specimens were exposed to UV and continuum lamp (UV-samples) or shielded from any light (Dark samples). Once colonies are formed, they are counted to establish the survival percentage with respect to the original sample. Details of the procedure and on results will be object of another

paper (Galletta et al., 2015) and they will not described here. In general, as visible in Figure 8, vegetative cells at "Summer" temperature are soon killed by the UV radiation of the lamp simulating the Sun on Mars surface. In particular, *Deinococcus radiodurans*, a bacterium known to resist to an instantaneous dose of up to 5,000 Gy[4] of ionizing radiation with no loss of viability (while 10 Gy can kill a human) appears to not resist to the combined action of the desiccation due to the low pressure (0.7 kPa) and the UV light, showing no survival in our samples after 5-8 minute of exposure.

A higher resistance is shown by the *Bacillus pumilus* SAFR032 strain[5], whose survival is still 10% after 10 minutes exposure. However, no cells survive after a few minutes of irradiation. Cooling of these cells produce an additional distruption, due to the sublimation of the water present inside the cells or to its conversion to ice. This means that, as expected, no biological activity of vegetative cells, rich in water for more than 75%, is possible in Martian conditions.

Table 3: Environmental parameters in the Mars simulated chambers and on the surface of Mars for the same parameters estimated from MGC TES data (Hargitai et al 2008) at the temperature at 14h after the Solstices at the Equator (labelled Summer day) and at the temperature at 14h in the Winter Solstice at 40°N (labelled Winter Day)

Parameter		
	Winter day	Summer day
Surface temperature (°C)	-80	+20
Atmospheric pressure (mbar)	7.5	7.5
UV flux (Wm^2)	3.4	3.4
Atmospheric composition (%)		
CO_2	95.5	95.5
Ar	1.6	1.6
N_2	2.7	2.7
O_2	0.13	0.13
CO	0.07	0.07

[4] GY(=Grey) is the absorption of 1J of energy by 1kg of matter

[5] kindly provided by Prof. K. Venkateswaran, JPL, CA, USA

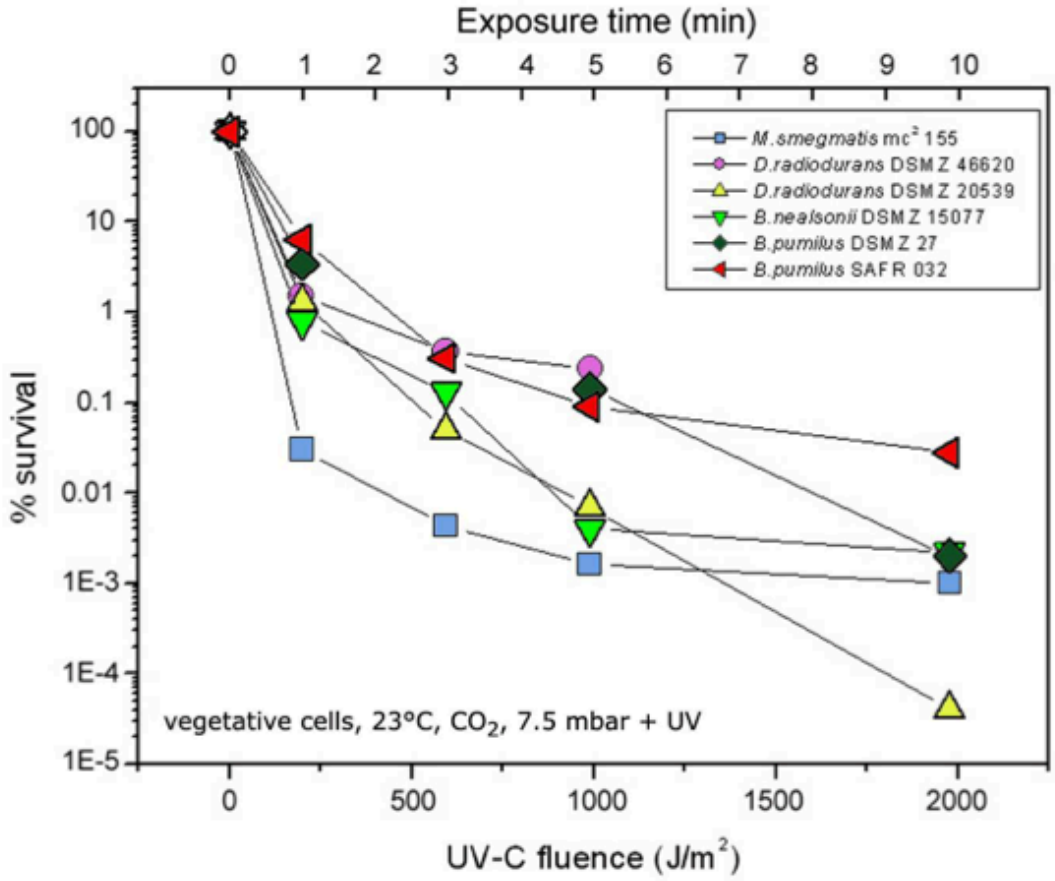

Figure 8: Survival in simulated Martian conditions for vegetative cells. All strains are killed in few minutes.

Then we have freezed the samples, bringing them to winter day temperature on Mars (typically -80 ◦C) and exposing them for longer time to UV radiation. These experiments are still ongoing, but preliminary results indicates that one of our *Bacillus* strains has a particular capability to survive in Martian conditions. More details will be found in Galletta et al. (2015).

This experiment made us aware of the functionality of the experimental set up. In the commissioning expeiment just described we used an UV lamp. In the planned experiment a new illuminator is necessary. Actually, what kind of lluminator and its characteristics are critical for the experiment. As planned the first phase foresees that the biological samples will irradiate with a solar light simulator. It is important to recall that it is not necessary to mimic the whole solar and stellar spectrum but only that portion that is inside the wavelength range defined in Table 2.

In order to build irradiation sources suitable for the experiment aims we have analyzed different light sources, mainly lamps and LEDs.

Lamps as Tungsten-Halogen, Mercury, Xenon and Metal-Halide are the most common radiation sources on the marketplace. They are broadband devices with an emission spectrum that can cover different wavelengths. The advantage of lamps is their high luminosity and broad bandwidth, but their disadvantage is the stillness of the spectrum and the impossibility to change it in real time. Though, the Tungsten-Halogen and the Xenon lamps are the only capable to emit in the NIR bandwidth (the peak of M0 type stars). One problem with this kind of lamps is the UV radiation. The emission of UV could be reduced or eliminated by means of appropriate filtering. We don't use this part of the spectrum of the lamp because we are not interested, at the moment, to test the survival of photosynthetic bacteria once exposed to UV. We want to test their productivity rate once they are in a safe location.

For the fiduciary experiment we need to reproduce the solar spectrum at the top of the atmosphere of the Earth and for this purpose Mercury, Xenon and Metal-Halide lamps could be useful like for example a Xenon Arc lamp a ORIEL 6258 300 Watt Ozone Free.

In order to represent the M0 star's planets we have to replicate the M0 star irradiation at the top of the atmosphere of an extrasolar terrestrial planet. LEDs could be a good alternative to lamps. In fact they are cheap, small and more versatile. For our purpose, one of the best features of LEDs is that it is possible to modulate their intensity and switch it on or off driving it by a PC. This is a great advantage respect to lamp illumination, that have a fix spectrum, because an array of LEDs could be used to match the desired part of the spectrum lightening them in sequence.

In collaboration with the Engineering Department of the Padova University we started a project to build a LED illuminator that is able to simulate the irradiation of an M0 star at different star-planet distances. We individuate the suitable LEDs for this aim. In Figure 9 the radiation curves of these LEDs are superimposed on the Black Body curve representative of an M0 star at the top of the atmosphere of a planet at the distance of the inner limit of the correspondent HZ. The illuminator is under development and we are tackling some hardware and optical problems. First of all we need a homogeneous illumination of the reaction cell that have an acceptance window of 40 mm in diameters. A solution is to use an integrating sphere that produce in

output a quasi-lambertian beam with a beam diameter a little bit greater than the reaction cell window.

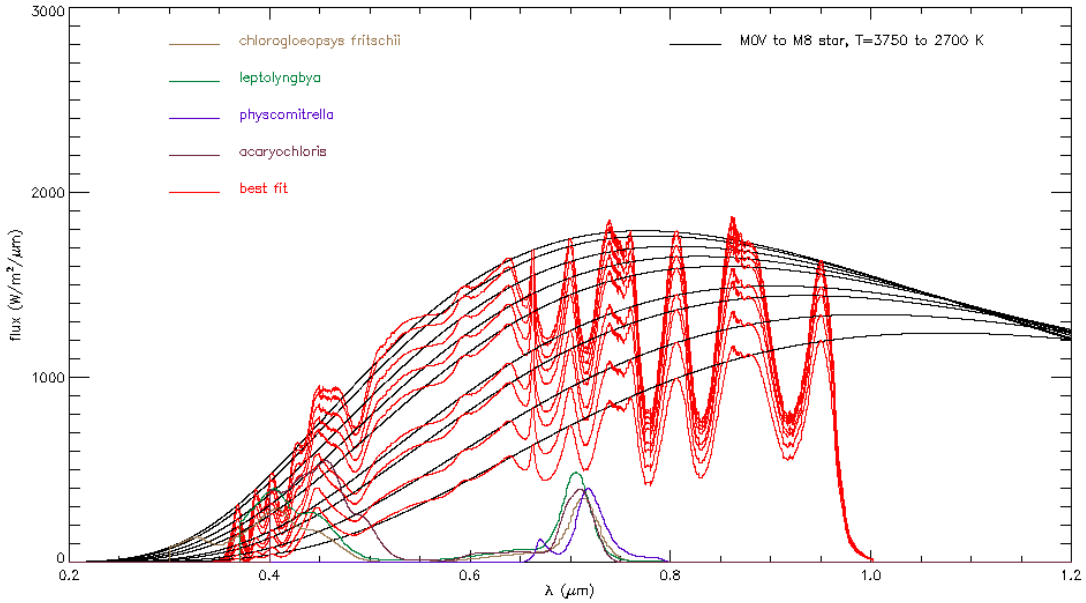

Figure 9: The radiance curves of several LEDs useful to mimic the Black Body representative of the M stars (from M0 to M8 spectral type). The absorption spectra of several photosynthetic bacteria are also shown.

5 Conclusions

M stars are the new frontier for the search for life and this in despite of their activity in young age and faintness. On the other hand M dwarf stars are very numerous in the Galaxy and also are simpler target in search for small planets with the transit technique. In fact, for the time being, some scores are gathered also in HZ of these stars. To handle remote sensing of the presence of life means to be able to individuate and interpret the biosignatures in the planetary spectra. In this paper we discussed the difficulties and the possibility of a life system to thrive on a planet in the HZ of a M star. As result we were able to define a set of scientific requirements for physical quantities to be reproduced in a laboratory experiment. Moreover we have outlined an experiment that is focused on the analysis of the formation of biosignatures in the atmosphere of a super Earth orbiting an M0 star well inside its habitable zone. The experimental set up was tested with a gas mixture simulating the Mars atmosphere. With this “commissioning” test we analyze the capacity of survivability of vegetative

cells under the UV irradiation in simulated different Martian condition. This experiment gave also useful information on the behavior of the environmental chamber during several duty cycles. It shows also that various kinds of experiments may be performed in the simulator, from inorganic chemistry to biological activity. Furthermore we outlined how to reproduce the irradiation coming from a host star colder then our Sun.

References

Angel, J. R. P., Cheng, A. Y. S., & Woolf, N. J. 1986, Nature, 322, 341

Ayres, T. R., 1997, J. Geophys. Res., 102, 1641

Batalha N.H., Rowe J.F., Bryson S.T., 2011, Ap. J. Supp. Ser., 204, 24

Bean J.L., Miller - Ricci E., Homeier D., 2010, Nature, 468, 669

Beer, A., 1852, Annalen der Physik 162, 78

Behrendt L, Schrameyer V, Qvortrup K, Lundin L, Sørensen SJ, Larkum AW, Kühl M. Appl Environ Microbiol. 2012 Jun; 78(11): 3896-904. doi: 10.1128/AEM.00397-12. Epub 2012 Mar 30

Borucki W. J., Koch D.G., Barsi G., et al., 2011, Ap. J., 736, 19

Brown T.M., 2001, Ap. J., 553, 1006.

Charbonneau D., Allen L.E., Megeath S.T. et al., 2005, ApJ., 626, 523.

Charbonneau D., Berta Z.K., Irwin J. et al., 2009, Nature, 462, 891

Biller, Liu, Wahhaj, et~al., 2013, ApJ, 777, 160

Billi, D., Viaggiu, E., Cockell, C. S., Rabbow, E., Horneck, G. and Onofri, S. 2011 Astrobiology 1 (0): 65–73.

Chauvin, Lagrange, Zuckerman et al. 2005, A&A, 438, L29

Chen, M., Telfer, A., Lin, S., Pascal, A., Larkum, A.W.D., Barber, J., and Blankenship, R.E. (2005) Photobiol. Sci. 4(12), 1060–1064.

Galletta, G., D'Alessandro, M., Bertoloni, G., Fanti, G., Dainese, E., Pelizzo, M., Ferri, F., Pavarin D., Bettanini C., Bianchini G., Debei, S.: 2007, Mem. S.A.It. Vol. 78, 608

Gough, D.O., 1981, Sol. Phys., 74, 21

Haberle, R.M., McKay, C.P., Tyler, D., and Reynolds, R.T., (1996), In *Circumstellar Habitable Zones: Proceedings of the First International Conference*, edited by L.R. Doyle, Travis House Publications, Menlo Park, CA, pp. 29.

Heath, M.J., Doyle, L.R., Joshi, M.M., and Haberle, R.M. (1999) Orig. Life Evol. Biosph. 29, 405–424.

Henry, T.J. (2004), In *ASP Conference Series 318: Spectroscopically and Spatially Resolving the Components of the Close Binary Stars*, edited by R.W. Hilditch, H. Hensberge, and K. Pavlovskipp, Astronomical Society of the Pacific, San Francisco, pp. 159.

Huang, S.-S. (1959), *Proc. Astron. Soc. Pacific*, 71, 421.

Huang, S-S. (1960), *Sci. Am.* 202, 55.

Ida, S., \& Lin, D. 2004, ApJ, 604, 388

Joshi, M.M., 2003, Astrobiology, 3, 415

Joshi, M.M., Haberle, R.M., and Reynolds, R.T. (1997), *Icarus* 129, 450.

Kasting J.F., Catling D., 2003, Annu. Rev. Astron. Astrophys., 2003, 41, 429

Kasting J.F., Whitmire D.P., Reynolds R.T., 1993, Icarus, 101, 108

Kiang, N. Y., J. Siefert, Govindjee, R. E. Blankenship and V. S. Meadows (2007). Astrobiology, 7(1) 222-51 (a)

Kiang, N.Y., Segura, A., Siefert, J., Tinetti, G., Govindjee, Blankenship, R.E., Cohen, M., Siefert, J., Crisp, D., and Meadows, V.S. (2007) Astrobiology 7(1), 252. (b)

Lafreniére, Doyon, Marois, et al., 2007, ApJ, 670, 1367

Lammer, H.; Bredehöft, J. H.; Coustenis, A.; Khodachenko, M. L.; Kaltenegger, L.; Grasset, O.; Prieur, D.; Raulin, F.; Ehrenfreund, P.; Yamauchi, M.; Wahlund, J.-E.; Grießmeier, J.-M.; Stangl, G.; Cockell, C. S.; Kulikov, Yu. N.; Grenfell, J. L.; Rauer, H., 2009: Astron. Astrop. Rev., 17, 181

Leger, A., Pirre, M., & Marceau, F. J. 1993, A&A, 277, 309

McKay, C.P. (2000) Geophys. Res. Lett. 27, 2153

Nutzman P., Charbonneau D., 2008, PASP, 120, 317

Raven, J.A. (1984) New Phytol. 98(4), 593

Rivera, E. J., et al. 2005, ApJ, 634, 625

Rothman L.S., Gordon J.E., Barber R.J. et al., 2010, J. Quant. Spectrosc. And Rad. Transf., 111, 2139

Scheer, H.; Green, B. R.; Parsons W. E., (2003), Advances in Photosynthesis and Respiration 13, 29

Schneider J., Dedieu C., Le Sidaner P., et al., 2011, A&A, 532, A79

Segura A., Kasting J.F., Meadows V. et al., 2005, Astrobiology, 5, 706

Selsis F., Kasting J.F., Levrard B. et al., 2007, A\&A, 476, 1373

Selsis F., Despois D., Parisot J.-P., 2002, A&A, 388, 985

Sozzetti, A., Lattanzi, M.G., Ligori, S., Smart, R.L., et al. 2008, In: Publications OATO (ed.) Pathways Towards Habitable Planets, Internal Report, vol. 110, pp. 20

Tarter J.C., Backus P.R., Mancinelli R.L. et al., 2007, Astrobiology, 7, 30

Tinetti G., Liang M.C., Vidal-Madjar A. et al., 2007, Ap. J., 654, L99.

Tinetti G., Rashby S., Yung Y.L., 2006, Ap.J., 644, L129

Valencia D., Sasselov D.D., O'Connell R.J., 2007, ApJ, 665, 1413

Werle, P., 1998, Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy 54, 197

Wolstencroft, R.D. and Raven, J.A. (2002) Icarus 157(2), 535–548.

Zwietering M.H., Jongenburger I., Rombouts F.M., van't Riet, 1990, Appl. Environ. Microbiol., 56, 1875