

## Phylogeny of Halictidae with an emphasis on endemic African Halictinae\*

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**Abstract** – We review the literature on phylogeny, fossil record, biogeography, and social evolution in Halictidae. We then present a phylogenetic analysis of tribal, generic, and subgeneric relationships within the subfamily Halictinae using a combined data set of three nuclear genes: long-wavelength (LW) opsin, *wingless*, and *EF-1 $\alpha$* . The data set includes 89 species in 34 genera representing all four halictid subfamilies, and all tribes of the subfamily Halictinae. Our study provides several new insights into the phylogeny of the African Halictinae. First, our results support a close relationship between *Mexalictus* (a small genus of bees occurring at high elevations in the mountains of western North and Central America) and the African/Asian genus *Patellapis*. Second, our results support placement of the parasitic genus *Parathrincochostoma* well within its host genus *Thrincochostoma*, suggesting that *Parathrincochostoma* should be treated as a subgenus of *Thrincochostoma*. Finally, our data set provides strong support for the monophyly of *Patellapis* (*sensu* Michener, 2000) and establishes monophyletic groups within the African subgenera that could be the basis for future taxonomic studies.

evolution / biogeography / bees / Apoidea

### 1. INTRODUCTION

Halictid bees are widely viewed as the “trash” bees of the world. Wheeler (1928, p. 90) commented that they “differ by such insignificant and elusive characters that they are the despair of taxonomists” and Michener (2000, p. 339) maligned them as “morpho-

logically monotonous”. Indeed, species in some widespread subgenera (e.g., *Lasioglossum* [*Dialictus*], *Halictus* [*Seladonia*]) are difficult to identify to species and often comprise the bulk of individuals encountered when collecting bees. However, halictids encompass much more biological, behavioral, and morphological diversity than commonly perceived. Halictidae is the second largest family of bees with over 3500 described species, and many more species remain to be discovered (Packer and Taylor, 1997). They are

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distributed on all continents except Antarctica and comprise between 20 and 60% of the bee species in faunal surveys in North America, Eurasia, and South America (MacKay and Knerer, 1979; Grixti and Packer, 2006). Some halictids are oligolectic (pollen-specialists), with behavioral and morphological adaptations for collecting pollen and/or nectar from a narrow range of closely related host-plants (e.g. *Systropha* and *Rophitinae* in the Rophitinae, *Dieunomia* and *Lipotriches* in the Nomiinae, and *Lasioglossum* [*Sphecodogastra*] in the Halictinae). Halictidae includes numerous lineages of cleptoparasitic and socially parasitic bees, and cleptoparasitism is estimated to have arisen at least eight times within Halictinae (Rozen, 2000). Halictids also show remarkable diversity in male morphologies associated with mating and courtship behavior (e.g., in Nomiinae [Wcislo and Buchmann, 1995]), but are perhaps best known for their diverse social behaviors, which include solitary, communal, semi-social and eusocial associations among adult females (see below).

Among the most interesting of the halictid bees are those from sub-saharan Africa. African halictine bees are poorly known both in terms of their phylogenetic affinities and their social behavior. In 2001 we initiated studies of the African halictine bees as part of a National Geographic project (NGS grant No. 6946-01 to BND, CE, LP, and KW). Between 2001 and 2005 we conducted field trips to South Africa (three trips), Kenya (one trip), and Madagascar (one trip) in an effort to sample and study halictid genera endemic to Africa. These studies provided important material for understanding the relationships of the African genera and for placing them in the context of a global halictid phylogeny. We report below the results of our phylogenetic analysis based on an expanded data set for these newly obtained African groups.

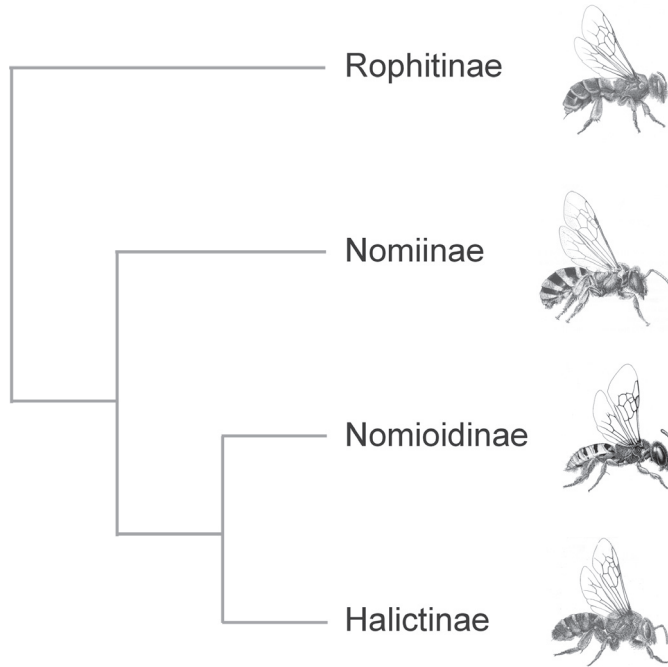
### 1.1. Diversity in social behavior

Halictid social behavior has been reviewed by Michener (1974, 1990), Seger (1991), Packer (1993a, 1997), Crespi (1996), Wcislo and Danforth (1997), and Schwarz et al.

(2006). Halictids are important bees for studies of social evolution because they show enormous intra- (Sakagami and Munakata, 1972; Yanega, 1988; Eickwort et al., 1996; Richards, 2000; Soucy, 2002) and inter-specific (Wcislo et al., 1993) variation in social behavior. Variation in social behavior within species is often linked with altitude and latitude such that populations at high elevation and latitude tend to be solitary, while populations at lower elevation and latitude tend to be eusocial (Sakagami and Munakata, 1972; Packer et al., 1989; Eickwort et al., 1996; Soucy, 2002; Soucy and Danforth, 2002). Diversity in social behavior among species was often thought to be due to frequent independent origins of eusociality in halictids (Eickwort, 1986) but recent molecular studies within and among genera of Halictinae have suggested that *reversals* from eusociality to solitary nesting may be common (Packer, 1997; Danforth, 1999; Danforth et al., 1999, 2003; Brady et al., 2006). Based on these studies, eusociality is estimated to have arisen just three to four times within Halictidae (Danforth, 1999; Brady et al., 2006; Coelho, 2002). Dating analyses based on relaxed clock methods (Brady et al., 2006) indicate that eusociality evolved recently in halictid bees (20–22 myBP). The impressive intraspecific variation in sociality within halictid species, their tendency to revert from eusociality to solitary nesting, and the recency of eusocial origins are all consistent with the view that halictids are in an early stage of social evolution relative to more advanced social taxa such as corbiculate bees, vespid wasps and ants (Brady et al., 2006).

### 1.2. Fossil record and antiquity

The halictid fossil record was reviewed by Engel (2001) as part of a complete review of Baltic amber bees. Halictid bees are present in the Baltic (Engel, 2001) and Dominican amber deposits (Engel, 1995, 1996, 2000; Engel and Rightmyer, 2000). All known halictid fossils are members of the tribes Augochlorini (Engel, 1995, 1996, 2000; Engel and Rightmyer, 2000), Halictini (Engel, 2001), and Caenohalictini (Michener and Poinar, 1996).



**Figure 1.** Subfamily-level phylogeny for the Halictidae based on morphology (Pesenko, 1999) and DNA sequence data (Danforth et al., 2004). Habitus drawings are as follows: Rophitinae (*Rophites algirus*), Nomiinae (*Lipotriches* [*Austronomia*] *australiana*), Nomioidinae (*Nomioides facilis*), Halictinae (*Agapostemon angelicus*). All drawings were done by Frances Fawcett except the *Lipotriches*, which is reproduced with permission from *Insects of Australia*, 2nd edition, Vol. II, p. 997, Fig. 42.45 (permission to use figures granted by CSIRO Australia and Cornell University Press).

Given the presumed phylogenetic affinities of these tribes (see below), this suggests that much of halictid evolution occurred prior to the oldest fossil (*Electrolictus antiquus*; Engel, 2001), which is estimated to be 42 million years old. Estimates based on fossil-calibrated chronograms (Danforth et al., 2004) suggest that all four subfamilies diverged well before the Cretaceous/Tertiary (K/T) boundary (65 my BP). A compression fossil from the Early Eocene (52–54.5 myBP) Quilchena site in British Columbia (Engel and Archibald, 2003) is seemingly a halictine, but assignment to tribe or genus is problematic. Nel and Petrulevičius (2003) recently described a *Lasiglossum* from Oligocene deposits in France. Despite the extensive fossil record of halictid bees (as compared to most other bee families), the minimum age estimate obtained from the fossils alone is considerably inferior to estimates based on combining molecular and

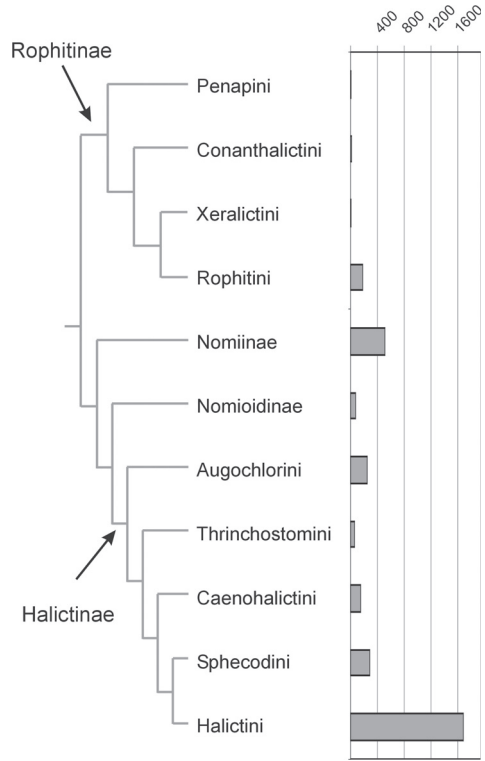
fossil data (Danforth et al., 2004; Brady et al., 2006).

### 1.3. Previous phylogenetic studies

Recent phylogenetic results based on a combination of molecular and morphological data support the sister-group relationship between Halictidae and Colletidae + Stenotritidae (Danforth et al., 2006). Both morphological (Pesenko, 1999) and molecular (Danforth et al., 2004) studies support the monophyly of the four traditionally recognized subfamilies: Rophitinae, Nomiinae, Nomioidinae, and Halictinae. Relationships among the four subfamilies are reasonably well supported (Fig. 1). Rophitinae comprises genera that are unique among halictid bees in that virtually all species are host-plant specialists (Patiny et al., 2007). Preferred host plants are restricted to the

Euasterid I clade, and the orders Gentianales, Lamiales, and Solanales appear to be the most important host plants. Patiny et al. (2007) provide a generic-level phylogeny for the subfamily based on a combination of morphological and molecular data and divided the subfamily into four monophyletic tribes (Penapini, Conanthalictini, Xeralictini and Rophitini; Fig. 2). Their tree implies frequent host-shifting in the Rophitinae with little evidence of co-cladogenesis between bees and their host plants (Patiny et al., 2007).

Nomiinae is a primarily paleotropical group with a diversity of genera in the African and Asian tropics and a small number of genera in Europe and North America. They are absent from South America and only enter the Neotropics in Cuba and southern Mexico. Nomiinae includes the only ground-nesting, solitary bee ever managed for commercial pollination: *Nomia melanderi* (Bohart, 1972). Catalogs exist for the African (Pauly, 1990), Australian (Cardale, 1993; Walker, 2006), and Western Hemisphere faunas (Moure and Hurd, 1987), but no detailed phylogenetic studies have been conducted at the generic, subgeneric, or species levels. This is unfortunate because the Nomiinae are a biologically fascinating group with bizarre and elaborate male morphologies (mostly involving hind legs [see Ribble, 1965 for illustrations] and genitalia) and courtship behaviors involving acoustic communication (Wcislo et al., 1992; Wcislo and Buchmann, 1995). Social behavior varies from species that nest solitarily to communal associations (Batra, 1966b; Wcislo, 1993; Vogel and Kukuk, 1994; Wcislo and Engel, 1997). Some species are host-plant specialists (Minckley et al., 1994) while others are clearly polylectic (Wcislo, 1993). In southern Africa, many species in the genus *Lipotriches* (subgenus *Lipotriches*) are narrowly oligolectic grass specialists that fly early in the morning before wind disperses the grass pollen (Immelman and Eardley, 2000; Tchuenguem Fohouo et al., 2002, 2004). Phylogenetic studies would help define more accurately the genera within Nomiinae, some of which are questionably monophyletic (e.g., *Nomia*, *Lipotriches*).



**Figure 2.** Relationships among subfamilies and tribes based on previous studies (Danforth et al., 2004; Patiny et al., 2007). Histogram at right shows the number of described species (data from Michener, 2000).

Nomioidinae are sometimes treated as a tribe of Halictinae (Engel, 2005), but they are morphologically very distinct and subfamily status seems warranted (Pesenko, 1999; Pesenko and Pauly, 2005). Nomioidinae include small to tiny metallic blue-green and yellow bees which occur primarily in arid regions of southern Europe, Africa and Madagascar, central Asia, and Australia (a single species, *Ceylactus (Ceylactus) perditellus*, occurs in Australia). Pesenko (2000) provided a phylogeny for Nomioidinae based on morphology and hypothesized an African origin, and Pesenko and Pauly (2005) revised the subfamily in Africa (excluding Madagascar). A diverse fauna exists in arid regions of Madagascar (Pauly et al., 2001). Some species of Nomioidinae are reported to be solitary (Radchenko, 1979) while others are reported

to form communal associations among nest-mates (Batra, 1966a).

Halictinae is the largest subfamily of halictid bees and it includes over 2400 species (Michener, 2000) in five tribes (Danforth et al., 2004) (Fig. 2). Foundations for halictine classification were laid by Michener (1978a,b) in his studies of the “strong-veined” genera. Halictinae is an enormous group and includes all of the eusocial and cleptoparasitic lineages of Halictidae. The group is clearly monophyletic with several morphological synapomorphies (including the divided prepygidial fimbria on the female T5). Tribes recognized by Danforth et al. (2004) include Augochlorini, Thrinchostomini, Caenohalictini, Sphecodini, and Halictini *sensu stricto*. Recognition of five tribes rather than the traditional two (Augochlorini and Halictini *sensu lato*; e.g., Michener, 2000) highlights the strikingly different morphologies and behaviors of these groups. The classification presented by Engel (2005; p. 28) is not based on a phylogenetic analysis of the group and recognizes paraphyletic taxa (“Tribe Halictini” and “Subtribe Halictina” *sensu* Engel).

Much phylogenetic work has been done in the Augochlorini (Eickwort, 1969b; Danforth and Eickwort, 1997; Engel, 2000). Recent morphological studies (Engel, 2000) have supported monophyly of two major lineages: the subtribe Corynura (including *Corynura*, *Halictillus* and *Rhinocorynura*) and the subtribe Augochlorina (the remaining genera). However, relationships at the generic and subgeneric levels were not robustly supported based on morphology alone. Augochlorini is a primarily Neotropical group with some representatives in temperate parts of North and South America. Social behavior is highly variable among species and genera and recent reports of facultative eusociality in *Augochloropsis* (Coelho, 2002) and *Megalopta* (Arneson and Wcislo, 2003; Smith et al., 2003; Wcislo et al., 2004) suggest that we are far from having a clear understanding of the full range and taxonomic distribution of eusociality. There are three cleptoparasitic genera/subgenera (*Temnosoma*, *Megalopta* [*Noctoraptor*], and *Megommation* [*Cleptomation*]) which appear to represent recent and independent derivations of cleptoparasitism.

Thrinchostomini currently includes two closely related genera (*Thrinchostoma* and *Parathrinchostoma*) of large, non-metallic bees. Thrinchostomini are restricted to the African and Asian tropics and Madagascar (Pauly et al., 2001). Of the 56 species of *Thrinchostoma*, 12 are endemic to Madagascar (Pauly et al., 2001). Some species are host-plant specific, with Asian species of *Diagonozus* apparently specializing on elongate flowers of the genus *Impatiens* (Sakagami, 1991; Sakagami et al., 1991). In Madagascar *Thrinchostoma* are collected on Melastomataceae, Malvaceae, Mimosaceae, Verbenaceae, Acanthaceae, and Asteraceae (Pauly et al., 2001). Melastomataceae has poricidal anthers and female *Thrinchostoma* “buzz” the flowers when foraging (Pauly et al., 2001). We have also observed this behavior on *Solanum* in South Africa and on Melastomataceae in Madagascar (BND, pers. obs.). We know little about the nesting biology or social behavior of Thrinchostomini, although their phenology in southern Africa would suggest that they are solitary. *Parathrinchostoma* includes two species that are endemic to Madagascar. Females lack pollen collecting structures and are inferred to be cleptoparasites of *Thrinchostoma* (Michener, 1978a, 2000), although this has never been verified through nest excavations.

Caenohalictini (like Augochlorini) is restricted to the New World and includes the large, mostly metallic bees related to *Agapostemon*. Eickwort (1969a) described the numerous morphological characters that distinguish this group from the superficially similar Augochlorini and Roberts and Brooks (1987) provided a generic-level revision of the tribe. Janjic and Packer (2003) analyzed the phylogeny of *Agapostemon* based on morphology using three other genera as outgroups. Unlike Augochlorini, which includes eusocial species, species of Caenohalictini are either solitary or communal (Abrams and Eickwort, 1980; Packer, 2006). Some genera (e.g., *Rhinotula*) are nocturnal bees with enlarged ocelli and compound eyes. Females of many species in other genera appear to be particularly active in the early morning.

Sphecodini includes four genera of cleptoparasitic bees (Michener, 1978a). Females

in these genera enter the nests of other bees and lay their eggs on or near the host's pollen provisions. Like other cleptoparasitic bees, females lack pollen-collecting structures and are heavily sclerotized. *Sphecodes* is the largest genus and is widespread on all continents except Australia. Hosts of *Sphecodes* include solitary and social Halictinae, Nomioidinae (*Nomioides*), Andrenidae (*Andrena*, *Calliopsis*, *Melitturga*, and *Perdita*) and Colletidae (*Colletes* and *Lonchopria*) (Michener, 2000). Unlike many cleptoparasitic bees, female *Sphecodes* aggressively attack the host female (or females in social nests) and may kill the host female(s) before oviposition in recently provisioned cells (Legewie, 1925; Knerer, 1973; Danforth, 1989; Sick et al., 1994). Female *Sphecodes monilicornis* were observed to close the host (*Lasioglossum malachurum*) cells following oviposition and a single female can lay multiple eggs per nest (Sick et al., 1994). Detailed studies on the mode of parasitism in *Sphecodes* are needed. Bogusch et al. (2006) recently documented interesting patterns of intra-specific host specialization in two species of *Sphecodes* (*S. ephippius* and *S. monilicornis*). While the species are generalists in the sense that members attack diverse host species, individuals appear to specialize on particular host species. Rigorous phylogenetic studies have not been conducted for this group. Because they are cleptoparasitic they tend to be rare and associating males and females has been difficult in some cases.

Halictini is the largest tribe of halictid bees and includes over 1600 described species (Fig. 2). A single genus (*Lasioglossum sensu lato*) accounts for over 70% of the species (Michener, 2000). *Lasioglossum* includes solitary, communal, eusocial, and socially parasitic species. *Lasioglossum* is particularly diverse in Australia where over 350 species occur (Cardale, 1993; Walker, 1986, 1995, 1997; Michener, 2000). Several species of Australian *Lasioglossum* are communal and some species are known in which males are dimorphic, with large-headed flightless males that remain within the nest and small-headed flight-capable males that can be collected on flowers (Knerer and Schwarz, 1976, 1978;

Kukuk and Schwarz, 1987, 1988; Kukuk and Crozier, 1990; Kukuk, 1992; Kukuk and Sage, 1994; McConnel-Garner and Kukuk, 1997). Some *Lasioglossum* are crepuscular and/or matinal fully nocturnal (e.g., *Sphecodogastra*) and narrow host-plant preferences are known in some *Lasioglossum* subgenera (e.g., *Sphecodogastra* on Onagraceae and *Hemihalictus* on *Pyropappus* [Asteraceae]). *Lasioglossum* in our sense includes groups traditionally treated as genera (e.g., *Homalictus*, *Echthralictus*, and *Urohalictus*). *Echthralictus* is a cleptoparasitic lineage closely related to, and presumably derived from, *Homalictus* (Michener, 2000). Our previous molecular data sets (Danforth and Ji, 2001; Danforth et al., 2004) have placed *Homalictus* (and presumably also *Echthralictus*) well within the larger genus *Lasioglossum*. Examination of the type species of *Urohalictus* indicates that it is similar to *L. (Parasphecodes)* and most likely a distinct subgenus of *Lasioglossum* (Danforth, pers. obs.). *Homalictus*, *Echthralictus* and *Urohalictus* all share the weakened outer wing venation that unites *Lasioglossum* and we believe the phylogenetic evidence would support treating these groups as subgenera of *Lasioglossum*. *Mexalictus* is an enigmatic genus of strong-veined halictine bees that occurs at high elevations from southeastern Arizona to northern Guatemala. These are rare bees that are collected in humid forests at elevations between ~2000 and ~3000 m (Eickwort, 1978; Packer, 1993b). Nothing is known of their social behavior or floral associations, although they appear to be solitary and polylectic. There are six described species. Most species are weakly metallic and resemble large, slender *Lasioglossum* (subgenus *Dialictus*). However, they lack the weakened outer wing veins of *Lasioglossum* and clearly represent a distinct strong-veined lineage (Eickwort, 1978). Among the most interesting halictine bees are those treated by Michener (2000) as the genus *Patellapis*. The genus is most diverse in southern Africa (with many subgenera including *Patellapis sensu stricto*, *Zonalictus*, *Chaetalictus*, *Lomatalictus*, and *Dictyohalictus*) and Madagascar (with endemic subgenera, such as *Madagalictus* and *Archihalictus*). One subgenus (*Pachyhalictus*)

**Table I.** Alternative classifications of “*Patellapis*” *sensu lato*.

Pauly 1999	Michener 2000
Genus <i>Patellapis</i> Friese	Genus <i>Patellapis</i>
Subgenus <i>Patellapis</i> Friese●	Subgenus <i>Archihalictus</i> <sup>1</sup> ●
Subgenus <i>Lomatalictus</i> Michener●	Subgenus <i>Chaetalictus</i> ●
Subgenus <i>Chaetalictus</i> Michener●	Subgenus <i>Dictyohalictus</i>
	Subgenus <i>Lomatalictus</i> ●
Genus <i>Madagalictus</i> Pauly ?	Subgenus <i>Pachyhalictus</i> ●
	Subgenus <i>Patellapis</i> ●
Genus <i>Pachyhalictus</i> Cockerell	Subgenus <i>Zonalictus</i> ●
Subgenus <i>Pachyhalictus</i> Cockerell●	
Subgenus <i>Dictyohalictus</i> Michener	
Subgenus <i>Archihalictus</i> Pauly●	

<sup>1</sup> including *Madagalictus* Pauly.

● Subgenera sampled in our study.

occurs in tropical Asia (Pauly, 2007) and northern Australia (Walker, 1993, 1996). Little is known about the social behavior in the over 160 species of *Patellapis s.l.* Timmermann and Kuhlmann (2008) recently provided the first detailed observations of nesting, foraging, and social behavior in this genus and found that female are polylectic foragers and that they form communal nests with up to eight females. Little is known about the phylogenetic affinities of *Patellapis* and the monophyly of the genus is not well supported by morphology. Conflicting classifications exist for this group (Tab. I). Pauly (1999, 2007) recognized three genera whereas Michener (2000) recognized a single, large genus (*Patellapis*) with multiple subgenera. It is worth noting that the oldest fossil halictid bee, *Electrolictus antiquus* from the Baltic amber (Engel, 2001), would be placed in the tribe Halictini *sensu stricto* and, in fact, may be very close to a *Patellapis*.

Previous phylogenetic studies of Halictinae (e.g., Danforth et al., 2004) were based on limited sampling of the African genera (including *Patellapis* [and its many subgenera], *Thrinchostoma*, *Parathrinchostoma*, and Nomioidini). In 2001 we initiated several collecting trips to Africa and Madagascar to obtain and study a representative sample of African halictine bees. As a result of these trips we have been able to include all the major African genera and subgenera in an expanded

data set. We report below the analysis of this data set based on parsimony. Our results establish more clearly the affinities of the African halictine bees.

## 2. MATERIALS AND METHODS

### 2.1. Data sets

We generated a data set based on three single copy nuclear genes: *wingless*, LW rhodopsin and elongation factor-1 $\alpha$ . Primers and sequencing protocols were provided in Danforth et al. (2004) and follow standard methods detailed in Danforth (1999), Ascher et al. (2001), and Danforth and Ji (2001). For more detailed information on genes and primers see <http://www.entomology.cornell.edu/BeePhylogeny/>

We added 31 species to the original data set (Danforth et al., 2004), including several additional genera and species of Nomioidinae, Thrinchostomini, Halictini and Sphecodini (see supplementary Table published as online material). Most new species were collected in South Africa and Madagascar. Species identifications were made by BND, KW, CE and LP. The total data set consisted of 89 species in 34 genera representing all four halictid subfamilies, and all tribes of the subfamily Halictinae (see supplementary Table published as online material). Numerous subgenera were included for the larger genera, such as *Halictus*, *Lasioglossum*, and *Patellapis*. For *Patellapis* we included all genera/subgenera recognized by Michener (2000) and Pauly (1999, 2007) (Tab. I) excluding *Patellapis*

**Table II.** Descriptive results for each gene and nucleotide position.

	No. Sites	No. PI % sites	A/T	<i>P</i> value <sup>1</sup>
<b>Opsin</b>				
nt1	234	65	57.15	1.00 (ns)
nt2	234	32	56.61	1.00 (ns)
nt3	234	182	31.71	0.219 (ns)
<b>Wingless</b>				
nt1	137	24	41.72	1.00 (ns)
nt2	137	4	54.27	1.00 (ns)
nt3	137	113	<b>14.78</b>	<0.001
<b>EF-1<math>\alpha</math></b>				
nt1	376	37	44.27	1.00 (ns)
nt2	376	19	58.21	1.00 (ns)
nt3	375	302	<b>55.61</b>	<0.001
intron	296	174	61.08	1.00 (ns)
<b>overall</b>	2536	952	49.97	0.478 (ns)

<sup>1</sup> *P*-value corresponds to the probability of rejecting the null hypothesis of homogeneity among taxa in base composition. Values in bold indicate significant deviation in base composition. “ns” indicates non-significant deviations from the null hypothesis of equal base frequencies across all taxa.

(*Dictyohalictus*), a subgenus of 12 rarely-collected species distributed through western (Gabon, Ivory Coast, Congo), eastern (Kenya, Uganda), and southern (Zimbabwe, South Africa) Africa (Pauly, 2007). We included all three subgenera of *Thrinchostoma* as well as their cleptoparasite (*Parathrinchostoma*). Supplementary table (published as supplementary online material) shows the complete list of taxa, locality data, and Genbank accession numbers. Genbank accession numbers for the newly added taxa are EU203218-EU203307. Voucher specimens are deposited in the Cornell University Insect Collection. Our combined data set is available on supplementary online material.

## 2.2. Phylogenetic methods

Alignments for all genes were generated in the Lasergene DNA Star software package using Clustal W. Reading frames and intron/exon boundaries were determined by comparison with published coding sequences for the honey bee, *Apis mellifera* (opsin [U26026], EF-1 $\alpha$  [AF015267]) and for *Drosophila* (*wingless* [J03650]).

We performed parsimony analyses using Paup\* v. 4.0b10 (Swofford, 2002). Initially we performed equal weights parsimony analyses on each of the three data sets and then combined the data sets into

a single analysis. We saw no obvious incongruence among data sets. Branch support for the individual data sets as well as the combined data set was estimated using bootstrap analysis (Felsenstein, 1985). For parsimony searches we performed 1000 random sequence additions. For calculating bootstrap proportions we performed 500 replicates with 10 random sequence additions per replicate.

## 3. RESULTS

The combined three-gene data set consisted of a total of 2539 aligned nucleotide sites (Tab. II). We excluded introns within opsin because they did not appear to be alignable. Conserved intron regions within EF-1 $\alpha$  were included (“introns included” below) or excluded (“introns excluded” below). With introns included we obtained 952 parsimony informative sites. With introns excluded we obtained 778 parsimony informative sites. While the overall data set showed an unbiased base composition, some partitions of the data set showed substantial base compositional bias (Tab. II). *Wingless* third positions are G/C rich, while EF-1 $\alpha$  third positions are A/T rich. For all three genes combined there was no



significant base compositional heterogeneity among taxa (Tab. II). The data set is complete for all genes and for all taxa except for four missing EF-1 $\alpha$  sequences, one missing *wingless* sequence, and two missing opsin sequences (supplementary Table published as online material). This represents 3.2% of the total aligned data set in terms of base pairs.

Analysis of the three gene data set with and without conserved regions of the EF-1 $\alpha$  introns provided largely congruent results. Analyses with introns included resulted in 24 trees of 5839 steps. Analysis with introns excluded resulted in 60 trees of 4692 steps. Figure 3 shows the consensus tree based on the analysis with introns included. Bootstrap values above the nodes are those calculated with introns included; bootstrap values below the nodes are calculated with introns excluded. Our trees recover each of the subfamilies and tribes described above with a high level of support (bootstrap values >95 when introns are included). The branching pattern of the halictid tribes corroborates an earlier study (Danforth et al., 2004), but some nodes are weakly supported (Fig. 3). Relationships within Halictini support monophyly of *Patellapis* in the sense of Michener (2000). Interestingly, our results place *Patellapis* as sister to *Mexalictus*, a small genus of poorly known species endemic to high altitude regions of the southwestern US, northern Mexico and Central America. While this node is not strongly supported (bootstrap value of 60 with introns included) there are morphological similarities between *Patellapis* and *Mexalictus* (Michener, 2000). If correct, this result would suggest that *Mexalictus* represents a highly relictual genus widely separated from its closely related Asian/African sister group (*Patellapis*).

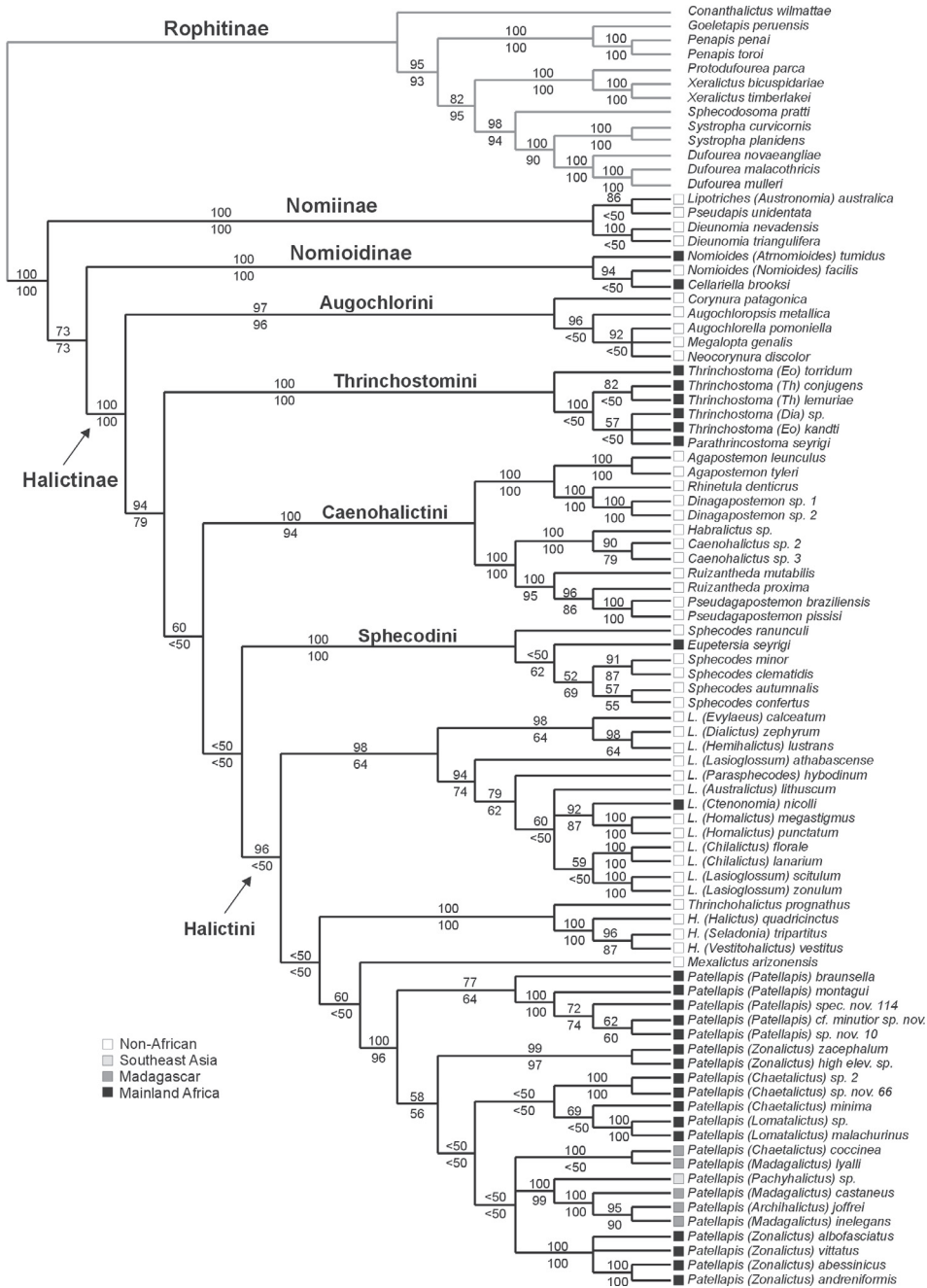
Relationships within the African taxa suggest that some genera may be paraphyletic. *Parathrincostruma*, for example, arises within *Thrincostruma*. Relationships within *Patellapis* suggest that some currently defined subgenera (*Zonalictus*, *Patellapis*, *Chaetalictus*) may be paraphyletic and in need of revision. Our topology suggests that the generic/subgeneric classification proposed by Michener (2000; Tab. I) more accurately reflects the topological relationships among

the subgenera than that of Pauly (1999, 2007). Pauly's genus *Pachyhalictus* (including *Pachyhalictus sensu stricto*, *Dictyohalictus* and *Archihalictus*) appears to be paraphyletic based on our analysis. *Patellapis* (*sensu* Pauly, 1999, 2007) is also apparently paraphyletic. Interestingly, the sole representative of the Asian *Patellapis* (*Pachyhalictus* sp.) is nested within a group of Madagascan species (Fig. 3). Finally, our results place *Eupetersia* within *Sphecodes*, supporting monophyly of Sphecodini, but paraphyly of *Sphecodes*. Figure 4 summarizes the relationships obtained based on this study. There remains some uncertainty about the tribal relationships, as well as the relationships among the three major branches of Halictini. An earlier study (Danforth et al., 2004) supported a grouping of *Lasioglossum* as sister to *Halictus* + *Thrincohalictus*. The current data set supports placement of *Halictus* + *Thrincohalictus* as sister to *Patellapis*.

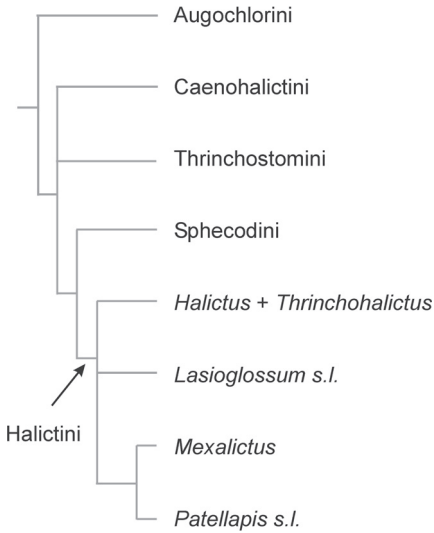
#### 4. DISCUSSION

Overall, our results provide the first analysis of halictine genera with the inclusion of all the major African taxa. Our study provides a basis for future taxonomic revisionary work on the genera of African halictine bees. Table III presents a revised classification based on these results.

Placement of *Mexalictus* as sister to the African/Asian genus *Patellapis*, while weakly supported, suggests a remarkable biogeographic disjunction between mountainous regions of western North America and southern Africa. Michener (2000, p. 340) noted the morphological similarity between the two genera and stated that *Mexalictus* and *Patellapis sensu stricto* "could both be archaic types surviving in limited areas." It seems likely that ancestral forms of this group were widely distributed throughout the northern Hemisphere and southward into Africa at some point in the past because dispersal from Africa to southwestern North America (or vice versa) seems an unlikely scenario. The relictual distribution of *Mexalictus* (restricted to humid forests at 2000–3000 m elevation) would likely have arisen as western North America became more arid during the Pleistocene.



**Figure 3.** Parsimony analysis of the three-gene data set. Bootstrap values are shown above the nodes for the exon+intron analysis; values below the nodes for the analysis of exons alone. Shaded boxes indicate the geographic location of the taxa sampled. Open boxes indicate non-African taxa; shaded boxes indicate taxa from Madagascar; lightly shaded boxes indicate taxa from southeast Asia; black boxes indicate mainland African taxa.



**Figure 4.** Summary cladogram showing relationships among tribes and select genera of Halictinae. Nodes that were not well supported (based on bootstrap analysis) are collapsed.

*Mexalictus arizonensis* would have to be considered a bee of conservation concern in the United States because of its limited distribution near the tops of a few mountains in southern Arizona. In terms of “phylodiversity” (Faith, 1992), *Mexalictus* represents an extremely important lineage in North and Central America.

Another important finding is the placement of *Parathrinchostoma* well within *Thrinchostoma* (Fig. 3). Many genera and subgenera of parasitic halictine bees make their host genus or subgenus paraphyletic. Examples include *Lasioglossum* (*Paralictus*) [parasite] and *Lasioglossum* (*Dialictus*) [host], *Echthralictus* and *Homalictus*, *Megalopta* (*Noctoraptor*) and *Megalopta* (*Megalopta*), *Megommation* (*Cleptommmation*) and other pollen-collecting subgenera of *Megommation*. In fact, Sphecodini is unusual among halictine cleptoparasites in that it represents an ancient parasitic lineage that has no close association with other non-parasitic groups of Halictinae. This may help explain its broad host usage. Our results place *Parathrinchostoma* within a group of species in the subgenera *Eoathrinchostoma* and *Diagonozus*, neither of which occurs in Madagascar. Michener (1978a, p. 313) com-

mented on the morphological similarity between *Parathrinchostoma* and *Eoathrinchostoma* and speculated that *Parathrinchostoma* may have arisen within mainland Africa and subsequently gone extinct, leaving the sole extant species on Madagascar. Given the placement of *Parathrinchostoma* it would now be appropriate to treat these two species as a subgenus of *Thrinchostoma* (Tab. III). It is clear from our results that the subgeneric limits within *Thrinchostoma* need to be revised.

Finally, our results provide strong support for monophyly of Michener’s (2000) *Patellapis sensu lato*. This group lacks clear morphological synapomorphies and some characters unite elements of *Patellapis* with other genera/subgenera of halictine bees (e.g., the unusual structure of the hind tibial and sternal scopa in the subgenus *Pachyhalictus* is reminiscent of *Homalictus*). The molecular data unambiguously support monophyly of this group (bootstrap support of 100 with introns included and 96 with introns excluded). However, most subgenera within *Patellapis* appear paraphyletic or polyphyletic. One obvious example is *P. (Zonalictus)* which appears as two unrelated lineages. The lineage including *P. (Zonalictus) zacephalum* occurs at high elevations in the eastern part of South Africa (Mpumalanga) while the lineage including *P. (Zonalictus) albofasciatus* occurs at lower elevations throughout southern and western South Africa. A broader survey of *P. (Zonalictus)* species, which occur north to the Arabian peninsula (Michener, 2000), will be needed to establish if these are indeed distinct clades. The one synapomorphy uniting *P. (Zonalictus)* is the white, yellow or pearly-banded margins of the abdominal terga. Bands such as these have arisen in unrelated bee groups (*Nomiocolletes* [Colletidae], *Nomiinae* [Halictidae], and *Amegilla* [Apidae]) and may prove to be convergent within *Patellapis*. We hope our results will provide impetus for taxonomic revisionary work at the species and subgeneric levels within *Patellapis*.

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**Table III.** Classification of the subfamily Halictinae. Tribal names are based on family group names listed in Michener (1986). Numbers of species are derived from Michener (2000), and other recent publications (e.g., Janjic and Packer 2003).

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Tribe Augochlorini	Beebe 1925 (25 genera, ~250 spp.)
	See Michener 2000 for list of genera <sup>1</sup>
Tribe Thrinchostomini	Sakagami 1974 (58 spp.)
	<i>Thrinchostoma</i> Saussure (>58 spp.) †
Tribe Caenohalictini	Michener 1954 (148 spp.)
	<i>Agapostemon</i> Guérin-Méneville (43 spp.)
	<i>Agapostemonoides</i> Roberts and Brooks (2 spp.) <sup>2</sup>
	<i>Caenohalictus</i> Cameron (45 spp.)
	<i>Dinagapostemon</i> Moure and Hurd (8 spp.)
	<i>Habralictus</i> Moure (22 spp.)
	<i>Paragapostemon</i> Vachal (1 spp.)
	<i>Pseudagapostemon</i> Schrottky (25 spp.)
	<i>Rhinotula</i> Friese (2 spp.)
	<i>Ruizantheda</i> Moure (4 spp.)
Tribe Sphecodini	Schenck 1869 (288 spp.)
	<i>Eupetersia</i> Blüthgen (29 spp.) †
	<i>Microsphecodes</i> Eickwort and Stage (7 spp.) †
	<i>Ptilocleptis</i> Michener (3 spp.) †
	<i>Sphecodes</i> Latreille (249 spp.) †
Tribe Halictini	Thomson 1869 (>1690 spp.)
	<i>Glossodialictus</i> Pauly (1 spp.)
	<i>Halictus</i> Latreille (217 spp.) †•
	<i>Lasioglossum</i> Curtis (>1200 spp., numerous subgenera) <sup>3</sup> †•
	<i>Mexalictus</i> Eickwort (6 spp.)
	<i>Patellapis</i> Friese (161 spp., numerous subgenera)
	<i>Thrincohalictus</i> Blüthgen (1 spp.)

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<sup>1</sup> See Eickwort (1969b) and Engel (2000) for generic treatments of the Augochlorini.

<sup>2</sup> *Agapostemonoides* was recognized as a subgenus of *Agapostemon* by Michener. Recent cladistic studies (Janjic and Packer, 2003) demonstrated that this is a valid genus.

<sup>3</sup> *Lasioglossum* in our sense includes *Homalictus*, *Echthralictus*, and *Urohalictus*.

• Indicates genera with both solitary and eusocial species; † indicates taxa with cleptoparasitic species.

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**Phylogénie des Halictidae avec une étude plus particulière sur les Halictinae africaines endémiques.**

**Apoidea / évolution / biogéographie / abeille / Afrique**

**Zusammenfassung – Phylogenie der Halictidae unter besonderer Berücksichtigung endemischer Halictinae aus Afrika.** In der Bienenfamilie Halictidae sind über 3500 Arten aus allen Kontinenten beschrieben. Diese Familie vereint die verschiedensten Bientaxa, darunter solitäre, soziale, kleptoparasitische und sozial-parasitische Arten. Viele Arten sind bezüglich ihrer Trachtpflanzen Generalisten, andere Spezialisten. Unsere Untersuchungen konzentrieren sich auf die Unterfamilie Halictinae, zu der die allermeisten Arten gehören. Wir analysierten die evolutionsbiologischen Beziehungen bei den verschiedenen Unterfamilien, Tribus und Gattungen der Halictiden unter besonderer Berücksichtigung der endemischen afrikanischen Halictinae. Verglichen mit Vertretern der Halictiden aus anderen Teilen der Welt hat man den afrikanischen Gattungen und Untergattungen bisher wenig Beachtung geschenkt. Wir wissen bisher sehr wenig über ihr Sozial- und Nistverhalten und wie diese Bienen in die Phylogenie der Halictiden einzuordnen sind. Wir analysierten die DNA-Sequenzen von drei Genen (long-wavelength opsin, *wingless* und elongation factor-1 $\alpha$ ) und einen Datensatz von insgesamt 2536 Nukleotide unter Verwendung von Alignment-Analysen und Parsimony-Methoden. Anhand unserer Ergebnisse identifizierten wir zwei distinkte endemische afrikanische Abstammungslinien. Erstens den Tribus *Thrinchostomini* (einschließlich der Gattungen *Thrinchostoma* und *Parathrinchostoma*), der früh im Stammbaum abzweigt. Zweitens eine gut abgesicherte monophyletische Gruppe (*Patellapis sensu lato*), die offensichtlich nahe verwandt mit dem Genus *Mexalictus* ist, der ausschließlich in hochgelegenen Gebieten vom westlichen Nordamerika bis nach Zentralamerika vorkommt. Dieses ungewöhnliche biogeographische Muster könnte darauf hinweisen, dass eine ursprünglich über die gesamte nördliche Hemisphäre verbreitete Gruppe ausgestorben ist. Wir überprüfen die Literatur zu den Halictiden bezüglich ihrer Beziehung zu Trachtpflanzen, ihrer Nistbiologie und dem Vorkommen von Kleptoparasitismus. Unsere Untersuchungen erlauben erstmals eine Beurteilung der Verwandtschaftsverhältnisse innerhalb der Halictinae anhand umfangreicher Proben aller afrikanischen Gattungen und Untergattungen.

#### Evolution / Biogeographie / Bienen / Apoidea

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# **Online Material**

**Table.** Taxa used in this study with locality data and Genbank accession numbers.

Taxon	Abbrev	No.	Subfamily	Tribe	Locality data	Opsin (~1200bp)	Wingless (450bp)	EF1 (~1600bp)
<b>Rophitinae</b>								
<i>Conanthalictus wilmattae</i>	Cowi	351	Rophitinae	Conanthalictini	USA: California [J. Neff]	AY 227934	AY 222553	AF435378
<i>Goletapis peruensis</i>	Gope	569	Rophitinae	Penapini	Peru: Lima Dept. [J.G. Rozen, Jr.]	AY 227923*	AY 222560	AF435386
<i>Penapis penai</i>	Pnpe	572	Rophitinae	Penapini	Chile: Huasco Prov. [J.G. Rozen, Jr.]	AY 227921	AY 222558	AF435401
<i>Penapis torvi</i>	Pnto	586	Rophitinae	Penapini	Chile: Region III [L. Packer]	AY 227922	AY 222559	AF435402
<i>Dufourea malacothricis</i>	Duma	235	Rophitinae	Rophitini	USA: Michigan [P. Lincoln]	AY 227917	AY 222554	AF435382
<i>Dufourea mulleri</i>	Dumu	233	Rophitinae	Rophitini	USA: Michigan [P. Lincoln]	AY 227918	AY 222555	AF435383
<i>Dufourea novaengliae</i>	Duno	354	Rophitinae	Rophitini	USA: New York, Cayuga Co.	AY 227919	AY 222556	AF435384
<i>Sphecodosoma pratti</i>	Sspr	355	Rophitinae	Rophitini	USA: Texas, Mason Co. [J. Neff]	AY 227924	AY 222561	AF435410
<i>Systropha curvicornis</i>	Sycu	350	Rophitinae	Rophitini	Austria: Vienna [M. Ayasse]	AY 227925	AY 222562	AF435411
<i>Systropha planidens</i>	Sypl	349	Rophitinae	Rophitini	Austria: Vienna [M. Ayasse]	AY 227926	AY 222563	AF435412
<i>Protodufourea parca</i>	Pfsp	237	Rophitinae	Xeralictini	USA: Arizona, Pima Co. [L. Packer]	AY 227920	AY 222557	AF435399
<i>Xeralictus bicuspidae</i>	Xrbi	566	Rophitinae	Xeralictini	USA: California, San Diego Co. [R. Snelling]	AY 227927	AY 222564	AF435413
<i>Xeralictus timberlakei</i>	Xrti	564	Rophitinae	Xeralictini	USA: California, Riverside Co. [R. Snelling]	AY 227928	AY 222565	AF435414
<b>Nomiinae</b>								
<i>Dieunomia nevadensis</i>	None	207	Nomiinae		USA: Arizona, Cochise Co.	AY 227931	AY 222568	AF435396
<i>Dieunomia triangulifera</i>	Notr	286	Nomiinae		USA: Kansas, Douglas Co.	AY 227932	AY 222569	AF435397
<i>Lipotriches (Austro.) australica</i>	Noau	754	Nomiinae		Australia: South Australia, Cowell	AY 227930	AY 222567	AF435395
<i>Pseudapis unidentata</i>	Psun	241	Nomiinae		Spain: Almeria Prov.	AY 227933	AY 222570	AF435404
<b>Nomioidinae</b>								
<i>Cellariella brooksi</i>	Cebr	1205	Nomioidinae		Madagascar: 8 km N. Tulear, 19 Oct. 2004	EU203280	EU203218	EU203249
<i>Nomioides (Atronomioides) tumidus</i>	Nosp	1209	Nomioidinae		Madagascar: 19 km N. Tulear, 17 Oct. 2004	EU203281	EU203219	EU203250
<i>Nomioides (Nomioides) facilis</i>	Nmsp	243	Nomioidinae		Spain: Granada Prov.	AY 227929	AY 222566	AF435394
<b>Halictinae</b>								
<i>Augochlorella pomoniella</i>	Aupo	592	Halictinae	Augochlorini	USA: California, Inyo Co. [J. Ascher]	AY 227935	AY 222572	AF435373
<i>Augochloropsis metallica</i>	Aume	334/5	Halictinae	Augochlorini	USA: New York, Tompkins Co.	AY 227934	AY 222571	AF140315
<i>Corynura patagonica</i>	Copt	581	Halictinae	Augochlorini	Chile: Region IX, near Temuco [L. Packer]	AY 227936	AY 222573	na
<i>Megalopta genalis</i>	Mgge	247	Halictinae	Augochlorini	Republic of Panama: Panama Prov. [M. Engel]	AY 227937	AY 222574	AF140316
<i>Neocorynura discolor</i>	Ncdi	249	Halictinae	Augochlorini	Colombia [M. Engel]	AY 227938	AY 222575	AF140317
<i>Agapostemon leunculus</i>	Agle	336	Halictinae	Caenohalictini	Republic of Panama: Panama Prov. [W. Wcislo]	AY 227939	AY 222576	AF435371
<i>Agapostemon tyleri</i>	Agty	230	Halictinae	Caenohalictini	USA: Arizona, Cochise Co.	AY 227940	AY 222577	AF140320
<i>Caenohalictus sp. 2</i>	Cnsp	578	Halictinae	Caenohalictini	Chile: Region I, Socoroma [L. Packer]	AY 227941	AY 222578	AF435376
<i>Caenohalictus sp. 3</i>	Cnsp	788	Halictinae	Caenohalictini	Brazil: Minas Gerais [D. Yanega]	AY 227942	AY 222579	AF435377

Table. Continued

Taxon	Abbrev	No.	Subfamily	Tribe	Locality data	Opsin	Wingless	EF1
						(~1200bp)	(450bp)	(~1600bp)
<i>Dinagapostemon sp. 1</i>	Dnsp	573	Halictinae	Caenohalictini	Republic of Panama: Chiriqui Prov. [L. Packer]	AY 227943	AY 222580	AF435380
<i>Dinagapostemon sp. 2</i>	Dnsp	790	Halictinae	Caenohalictini	Republic of Panama: Cocle Prov. [L. Packer]	AY 227944	AY 222581	AF435381
<i>Habralictus sp.</i>	Hasp	786	Halictinae	Caenohalictini	Brazil: Minas Gerais [D. Yanega]	AY 227945	AY 222582	AF435387
<i>Pseudagapostemon braziliensis</i>	Psbr	347	Halictinae	Caenohalictini	Brazil: Minas Gerais [D. Yanega]	AY 227946	AY 222583	AF140323
<i>Pseudagapostemon pissisi</i>	Pspis	576	Halictinae	Caenohalictini	Chile: Region VIII (nr. Temuco) [L. Packer]	AY 227947	AY 222584	AF435403
<i>Rhinotula denticrus</i>	Rhdn	799	Halictinae	Caenohalictini	Republic of Panama: Panama Prov. [D. Roubik]	AY 227948	AY 222585	AF435405
<i>Ruizantheda mutabilis</i>	Rumt	574	Halictinae	Caenohalictini	Chile: Region VIII (nr. Temuco) [L. Packer]	AY 227949	AY 222586	AF435406
<i>Ruizantheda proxima</i>	Rupr	575	Halictinae	Caenohalictini	Chile: Region VIII (nr. Temuco) [L. Packer]	AY 227950	AY 222587	AF435407
<i>Halictus (Halictus) quadricinctus</i>	Haqu	89	Halictinae	Halictini	France: Dordogne [C. Plateaux-Quenu]	AY 227956	AY 222592	AF140334
<i>Halictus (Seladonia) tripartitus</i>	Hatr	93	Halictinae	Halictini	USA: Arizona, Cochise Co.	AY 227957	AY 222593	AF140310
<i>Halictus (Vestitohalictus) vestitus</i>	Vève	373	Halictinae	Halictini	Spain: Granada Prov.	AY 227958*	AY 222594	AF140313
<i>Lasioglossum (Australictus) lithuscum</i>	Auli	711	Halictinae	Halictini	Australia: Victoria	AY 227962	AY 222598	AF435372
<i>Lasioglossum (Chilalictus) florale</i>	Chfl	320	Halictinae	Halictini	Australia: South Australia [S. Reyes]	AY 227966	AY 222602	AF264792
<i>Lasioglossum (Chilalictus) lanarium</i>	Chla	316	Halictinae	Halictini	Australia: Victoria [S. Reyes]	AY 227967	AY 222603	AF264793
<i>Lasioglossum (Ctenonomia) nicolli</i>	Ctni	1192	Halictinae	Halictini	Madagascar: Parc National Isalo	EU203307	EU203248	EU203279
<i>Lasioglossum (Dialictus) zephyrum</i>	Dizp	74	Halictinae	Halictini	USA: New York, Seneca Co.	AF448918*	AY 222607	AF435379
<i>Lasioglossum (Evylaeus) calceatum</i>	Evca	105	Halictinae	Halictini	France: Dordogne [C. Plateaux-Quenu]	AF448877*	AY 222608	AF435385
<i>Lasioglossum (Hemihalictus) lustrans</i>	Helu	186	Halictinae	Halictini	USA: Texas, Bastrop Co. [J. Neff]	AF448904*	AY 222609	AF435388
<i>Lasioglossum (Homalictus) megastigmus</i>	Homg	360	Halictinae	Halictini	Australia: Western Australia, Bluff Knoll [S. Reyes]	AY 227964	AY 222600	AF264839
<i>Lasioglossum (Homalictus) punctatus</i>	Hopu	245	Halictinae	Halictini	Australia: South Australia, Adelaide [S. Reyes]	AY 227965	AY 222601	AF435389
<i>Lasioglossum (Lasioglossum) athabascense</i>	Laat	556	Halictinae	Halictini	USA: New York, Tompkins Co.	AF448867	AY 222604	AF435390
<i>Lasioglossum (Lasioglossum) scitulum</i>	Lasc	550	Halictinae	Halictini	Japan: Shimane Prefecture [R. Miyanaga]	AY 227968	AY 222605	AF435391
<i>Lasioglossum (Lasioglossum) zonulum</i>	Lazo	284	Halictinae	Halictini	USA: New York, Tompkins Co.	AY 227969	AY 222606	AF264855
<i>Lasioglossum (Parusphecodes) hybodinum</i>	Pahy	299	Halictinae	Halictini	Australia: South Australia [S. Reyes]	AY 227963	AY 222599	AF264857
<i>Mexalictus arizonensis</i>	Mxaz	98	Halictinae	Halictini	USA: Arizona, Santa Cruz Co. [L. Packer]	AY 227959	AY 222595	AF140322
<i>Putellapis (Archihalictus) joffrei</i>	Arjo	1188	Halictinae	Halictini	Madagascar: Parc National Ranomafana	EU203301	EU203242	EU203273
<i>Putellapis (Chaetalictus) coccinea</i>	Chco	1189	Halictinae	Halictini	Madagascar: Parc National Ranomafana	EU203302	EU203243	EU203274
<i>Putellapis (Chaetalictus) minima</i>	Chsp	911	Halictinae	Halictini	South Africa: Western Cape Prov., Citrusdal	EU203289	EU203231	EU203261
<i>Putellapis (Chaetalictus) sp. 2</i>	Chsp	603	Halictinae	Halictini	South Africa: Kwazulu-Natal [L. Packer]	AY 227952	AY 222588	AF435374
<i>Putellapis (Chaetalictus) spec. nov. 66</i>	Chsp	956	Halictinae	Halictini	South Africa: Mpumalanga, Graskop	EU203290	EU203230	EU203262
<i>Putellapis (Lomatalictus) sp.</i>	Losp	604	Halictinae	Halictini	South Africa: Gauteng [C. Eardley]	AY 227953	AY 222589	AF435393
<i>Putellapis (Lomatalictus) malachurinus</i>	Lole	954	Halictinae	Halictini	South Africa: Mpumalanga, Graskop	EU203291	EU203232	EU203263

Table. Continued

Taxon	Abbrev	No.	Subfamily	Tribe	Locality data	Opsin	Wingless	EF1
						(~1200bp)	(450bp)	(~1600bp)
<i>Patellapis (Madagalictus) castaneus</i>	Mdca	1194	Halictinae	Halictini	Madagascar: Parc National Ranomafana	EU203303	EU203244	EU203275
<i>Patellapis (Madagalictus) inelegans</i>	Mdin	1195	Halictinae	Halictini	Madagascar: Parc National Ranomafana	EU203304	EU203245	EU203276
<i>Patellapis (Madagalictus) lyalli</i>	Mdly	1204	Halictinae	Halictini	Madagascar: Parc National Ranomafana	EU203305	EU203246	EU203277
<i>Patellapis (Pachyhalictus) sp.</i>	Phsp	357	Halictinae	Halictini	Vietnam: Ban Don [L. Packer]	AY 227954	AY 222590	AF435400
<i>Patellapis (Patellapis) braunsella</i>	Pabr	1196	Halictinae	Halictini	South Africa: Western Cape Prov., Nieuwoudtville	EU203306	EU203247	EU203278
<i>Patellapis (Patellapis) cf. minutior spec. nov.</i>	Pasp1	915	Halictinae	Halictini	South Africa: Western Cape Prov., Citrusdal	EU203293	EU203234	EU203265
<i>Patellapis (Patellapis) montagui</i>	Pamn	914	Halictinae	Halictini	South Africa: Western Cape Prov., Clanwilliam	EU203292	EU203233	EU203264
<i>Patellapis (Patellapis) spec. nov. 10</i>	Pasp2	918	Halictinae	Halictini	South Africa: Western Cape Prov., Vánrhynsdorp	EU203294	EU203235	EU203266
<i>Patellapis (Patellapis) spec. nov. 114</i>	Chrv	909	Halictinae	Halictini	South Africa: Northern Cape Prov., Port Nolloth	EU203288	EU203229	EU203260
<i>Patellapis (Zonalictus) high elevation sp.</i>	Zohe	957	Halictinae	Halictini	South Africa: Mpumalanga, Sabie	EU203300	EU203241	EU203272
<i>Patellapis (Zonalictus) abessinicus</i>	Zoab	950	Halictinae	Halictini	South Africa: Free State, Harrismith	EU203295	EU203236	EU203267
<i>Patellapis (Zonalictus) albofasciatus</i>	Zosp	949	Halictinae	Halictini	South Africa: Free State, Koppies	EU203297	EU203238	EU203269
<i>Patellapis (Zonalictus) andreniformis</i>	Zoan	919	Halictinae	Halictini	South Africa: Western Cape Prov., Citrusdal	EU203296	EU203237	EU203268
<i>Patellapis (Zonalictus) vittatus</i>	Zovt	921	Halictinae	Halictini	South Africa: Western Cape Prov., Citrusdal	EU203298	EU203239	EU203270
<i>Patellapis (Zonalictus) zacephalum</i>	Zoeh	955	Halictinae	Halictini	South Africa: Mpumalanga: Graskop	EU203299	EU203240	EU203271
<i>Thrincohalictus prognathus</i>	Thpr	434	Halictinae	Halictini	Israel: Golan Heights [L. Packer]	AY 227955	AY 222591	AF140326
<i>Eupetersia (Eupetersia) seyrigi</i>	Eusy	1243	Halictinae	Sphecodini	Madagascar: Parc National Ranomafana	EU203287	EU203228	EU203259
<i>Sphecodes autumnalis</i>	Spau	1019	Halictinae	Sphecodini	NY: Cortland Co., Cortlandville [Ascher]	na	EU203225	EU203256
<i>Sphecodes clematidis</i>	Spcl	1020	Halictinae	Sphecodini	NY: Tompkins Co., Salmon Creek [Ascher]	na	EU203226	EU203257
<i>Sphecodes confertus</i>	Spcn	1017	Halictinae	Sphecodini	NY: Tompkins Co., Salmon Creek [Ascher]	na	EU203227	EU203258
<i>Sphecodes minor</i>	Spmi	21	Halictinae	Sphecodini	USA: New York, Tompkins Co.	AY 227960	AY 222596	AF140324
<i>Sphecodes ranunculi</i>	Spra	337	Halictinae	Sphecodini	Canada: Nova Scotia [L. Packer]	AY 227961	AY 222597	AF140325
<i>Parathrinchostoma seyrigi</i>	Ptsk	1248	Halictinae	Thrinchostomini	Madagascar: Parc National Ranomafana [Pauly]	EU203287	EU203224	EU203255
<i>Thrinchostoma (Diagonozus) sp.</i>	Trsp	464	Halictinae	Thrinchostomini	Borneo: Sarawak [S. Heydon]	AY 227951	na	na
<i>Thrinchostoma (Eothrinchostoma) kandi</i>	Thkd	952	Halictinae	Thrinchostomini	South Africa: Kwazulu-Natal, Hluhluwe	EU203282	EU203220	EU203251
<i>Thrinchostoma (Eothrinchostoma) torridum</i>	Thto	953	Halictinae	Thrinchostomini	South Africa: Kwazulu-Natal, Hluhluwe	EU203283	EU203221	EU203252
<i>Thrinchostoma (Thrinchostoma) conjugens</i>	Thco	1199	Halictinae	Thrinchostomini	Madagascar: Parc National Ranomafana	EU203284	EU203222	EU203253
<i>Thrinchostoma (Thrinchostoma) lemuriæ</i>	Thle	1200	Halictinae	Thrinchostomini	Madagascar: Parc National Ranomafana	EU203285	EU203223	EU203254

\* = OpsFor/Rev only (others are For/Rev4)