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1 Ortervirales: A new viral order unifying five families of reverse-transcribing viruses

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widespread in animals, plants, algae and fungi (1, 2). This broad distribution suggests ancient origin(s) of these viruses, possibly concomitant with the emergence of eukaryotes (3). Reverse-transcribing viruses include prominent human pathogens, such as human immunodeficiency viruses 1 and 2 (HIV-1/2) and hepatitis B virus, as well as plant pathogens that cause considerable economic losses (4). The International Committee on Taxonomy of Viruses (ICTV) traditionally classified reversetranscribing viruses into five families: Caulimoviridae, Hepadnaviridae, Metaviridae, Pseudoviridae, and Retroviridae (5). In 2018, the ICTV recognized an additional family, Belpaoviridae, which contains the genus Semotivirus (previously included in Metaviridae (6)). The infection cycles, nucleic acid types, genome organizations, and virion morphologies of these viruses are very diverse. Indeed, reverse-transcribing viruses are distributed between two Baltimore Classes of viruses. Belpaoviruses, metaviruses, pseudoviruses — better known as Bel/Pao, Ty3/Gypsy, and Ty1/Copia retrotransposons, respectively (1, 7) — and retroviruses typically have single-stranded RNA genomes (Table 1) and frequently integrate into the host genomes as part of their replication cycles (Baltimore Class VI). In contrast, members of the families Caulimoviridae and Hepadnaviridae, often referred to as "pararetroviruses" (8), encapsidate circular double-stranded DNA genomes and do not actively integrate into host chromosomes (Baltimore Class VII). However, capture of pararetroviral DNA in host genomes, presumably by illegitimate recombination, is commonplace, particularly in plants, giving rise to the corresponding endogenous elements (9, 10). Mechanistic studies on the replication cycles of reverse-transcribing viruses of different families have revealed many similarities that have been reinforced by comparative genomics of the viral reverse transcriptases (RTs), the hallmark enzymes encoded by all reverse-transcribing viruses. Indeed, phylogenetic analyses support the monophyly of all viral RTs, to the exclusion of those encoded by non-viral retroelements from both eukaryotes and prokaryotes (11, 12). In addition to the evidence from the RT phylogeny, belpaoviruses, caulimoviruses, metaviruses, pseudoviruses, and

retroviruses share several conserved features that hepadnaviruses lack (Table 1). In particular, the

polymerase (Pol) polyproteins of belpaoviruses, metaviruses, pseudoviruses, and retroviruses possess

Reverse-transcribing viruses, which synthesize a copy of genomic DNA from an RNA template, are

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similar domain architectures. These Pol polyproteins contain an aspartate protease, which is responsible for the processing of viral polyproteins, and an integrase of the DDE recombinase superfamily. The genomes of these viruses also share long terminal repeats (LTRs) (13). Within certain clades, Pol polyproteins of retroviruses and metaviruses share additional features, such as a dUTPase domain (14-16) and the GPY/F subdomain of the integrase (17, 18). Caulimoviruses also possess a homologous aspartate protease domain in their Pol polyprotein (19), but lack an integrase and LTR. However, RT-based phylogenies consistently place these plant-infecting viruses as a sister clade to the metaviruses (Figure 1), suggesting that among "pararetroviruses", encapsidation of a DNA genome is a homoplasious character and therefore not a reliable criterion for classification. The basal branches of the RT tree are not resolved and are presented as a multifurcation in Figure 1. This topology is at least compatible with placing the Hepadnaviridae clade outside the viral group that includes belpaoviruses, caulimoviruses, metaviruses, pseudoviruses, and retroviruses.

Belpaoviruses, caulimoviruses, metaviruses, pseudoviruses, and retroviruses share not only homologous proteins involved in genome replication and polyprotein processing, but also the two principal protein components of the virions, namely, the capsid and nucleocapsid proteins/domains (20-22), although the nucleocapsid domain appears to be absent in spumaretroviruses (family Retroviridae; Table 1). By contrast, hepadnaviruses encode an unrelated capsid protein (23). These findings suggest that belpaoviruses, caulimoviruses, metaviruses, pseudoviruses, and retroviruses have evolved from a common viral ancestor, rather than from distinct capsid-less retrotransposons (20).

Finally, similarities between belpaoviruses, caulimoviruses, metaviruses, pseudoviruses, and retroviruses extend to the mechanism of replication priming. All these viruses utilize host tRNA molecules as primers for genome replication by reverse transcription (24), whereas hepadnaviruses use a specific protein priming mechanism mediated by the polymerase terminal protein domain (25).

Taken together, the common complement of proteins required for genome replication, polyprotein processing, and virion formation, the topology of the RT phylogenetic tree, and mechanistic similarities in genome replication present strong evidence that belpaoviruses, caulimoviruses, metaviruses, pseudoviruses, and retroviruses share a common evolutionary origin. The hepadnaviruses, which typically branch out at the base of the viral RT clade (Figure 1), possess a

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unique capsid protein and employ a distinct replication mechanism, appear to be more distantly related to all these virus families. In recognition of these relationships, the ICTV has recently regrouped the families Belpaoviridae, Caulimoviridae, Metaviridae, Pseudoviridae and Retroviridae into an order Ortervirales (orter: an inversion of retro, which was derived from reverse transcription; virales: suffix for an order). This change in taxonomy acknowledges and formalizes the long-proposed evolutionary relationship among most groups of reverse-transcribing viruses (26). We note that although hepadnaviruses are not included in the order, they might be unified with other reverse-transcribing viruses at a higher taxonomic level in the future. Acknowledgments This work was supported in part through Battelle Memorial Institute's prime contract with the US National Institute of Allergy and Infectious Diseases (NIAID) under Contract No. HHSN272200700016I (JHK). EVK is supported by intramural funds of the US Department of Health and Human Services (to the National Library of Medicine). SS acknowledges support from SRI Funds from Mississippi Agriculture and Forestry Experiment Station of Mississippi State University. M. Krupovic, B. Harrach, S. Sabanadzovic, H. Sanfaçon, and J. H. Kuhn are members of the 2014–2017 International Committee on Taxonomy of Viruses (ICTV) Executive Committee. J. Blomberg, J. M. Coffin, H. Fan, R. Gifford, W. Johnson, D. Lindemann, J. Mayer, J. P. Stoye, and M. Tristem are members of the 2014–2017 ICTV Retroviridae Study Group. I. Dasgupta, A. D. Geering, R. Hull, J. F. Kreuze, B. Lockhart, E. Muller, N. Olszewski, H. R. Pappu, M. Pooggin, K. R. Richert-Pöggeler, J. E. Schoelz, S. Seal, L. Stavolone, and P.-Y. Teycheney are members of the 2014–2017 ICTV Caulimoviridae Study Group. R. Hull is retired from the John Innes Centre, Norwich, Norfolk,

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Figure legend

Figure 1. Maximum likelihood phylogeny of viral reverse transcriptases. The tree includes sequences of 290 viruses belonging to all ICTV-

Table 1. Features shared by reverse-transcribing viruses.

| Family Subfamily | | Retroviridae | | Metaviridae | Pseudoviridae | Belpaoviridae | Caulimoviridae | Hepadnaviridae |
|---------------------|-----------|-------------------|-------------------|-------------|---------------|---------------|----------------|----------------|
| | | Orthoretrovirinae | Spumaretrovirinae | | | | | - Articles |
| Pol | RT-RH | + | + | + | + | + | + | + |
| | Protease | + | + | + | + | + | + | - |
| | Integrase | + | + | + | + | + | - | - |
| Gag | CA/CP | + | + | + | + | + | (+) | - |
| | NC | + | - | + | + | + | + | - |
| LTR | | + | + | + | + | + | -s | -# |
| Priming | | tRNA | tRNA | tRNA | tRNA | tRNA | tRNA | TP |
| Genome type | | ssRNA | ssRNA/dsDNA* | ssRNA | ssRNA | ssRNA | dsDNA | dsDNA |

- * Members of the subfamily Spumaretrovirinae contain both ssRNA and dsDNA in extracellular particles and reverse transcription occurs during virus assembly and disassembly.
- \$ In the genus Petwirus (Caulimoviridae) an inactivated integrase-like domain and quasi (long) terminal repeats have been identified (27, 28), suggesting that certain ancestral elements have been lost during the evolution of caulimoviruses.
- # Upstream of the capsid protein gene, hepadnavirus genomes contain a sequence showing similarity to the U5 region of the retroviral LTR (29). Abbreviations: CA/CP, capsid protein; Gag, group-specific antigen; LTR, long terminal repeats; NC, nucleocapsid protein; RH, RNase H; RT, reverse transcriptase; Pol, polymerase polyprotein; TP, terminal protein.

