

1           **Morphology of the earliest reconstructable tetrapod *Parmastega aelidae***

2  
3 Pavel A. Beznosov<sup>1</sup>, Jennifer A. Clack<sup>2</sup>, Ervīns Lukševičs<sup>3</sup>, Marcello Ruta<sup>4</sup>, Per Erik  
4 Ahlberg<sup>5</sup>

5  
6 <sup>1</sup> Institute of Geology, Komi Science Centre, Ural Branch of the Russian Academy of  
7 Sciences, Pervomaykaya st. 54, Syktyvkar, 167982, Russia

8 <sup>2</sup> University Museum of Zoology, University of Cambridge, Downing St., Cambridge  
9 CB2 3EJ, UK

10 <sup>3</sup> University of Latvia, Raiņa bulvāris 19, Rīga, LV-1586, Latvia

11 <sup>4</sup> School of Life Sciences, University of Lincoln, Joseph Banks Laboratories, Green  
12 Lane, Lincoln LN6 7DL, UK

13 <sup>5</sup> Department of Organismal Biology, Uppsala University, Norbyvägen 18A, SE-752  
14 36, Uppsala, Sweden

15  
16 **The taxonomic diversity of Devonian tetrapods has increased dramatically in**  
17 **recent decades, but much of this diversity consists of tantalising fragments. The**  
18 **interpretative framework for the earliest stages of tetrapod evolution is still**  
19 **dominated by the near-complete *Ichthyostega* and *Acanthostega*, with supporting**  
20 **roles for the less complete but partly reconstructable *Ventastega* and *Tulerpeton*.**  
21 **All four are of late Famennian age, 10 million years younger than the earliest**  
22 **tetrapod fragments and nearly 30 million years younger than the oldest**  
23 **footprints. Here we describe a tetrapod from the earliest Famennian of Russia,**  
24 ***Parmastega aelidae* gen. et sp. nov., represented by three-dimensional material**  
25 **that allows reconstruction of almost the entire skull and dermal shoulder girdle.**  
26 **Its raised orbits, lateral line canals and weakly ossified postcranial skeleton**  
27 **suggest a largely aquatic, surface-cruising animal. In Bayesian and parsimony-**  
28 **based phylogenetic analyses the majority of trees place *Parmastega* as sister**  
29 **group to all other tetrapods.**

30  
31 The rate of discovery of Devonian tetrapods accelerated greatly during the late 20th  
32 and early 21st centuries. The description of *Ichthyostega* in 1932 was followed by  
33 *Acanthostega* in 1952, *Metaxygnathus* in 1977 and *Tulerpeton* in 1984; all other  
34 genera (*Hynerpeton*, *Ventastega*, *Elginerpeton*, *Obruchevichthys*, *Densignathus*,

*Sinostega*, *Jakubsonia*, *Ymeria*, *Webererpeton*, *Tutusius*, *Umzantsia*) have been described or identified as tetrapods since 1994<sup>1-13</sup>. Un-named Devonian tetrapod material has been described from Belgium<sup>14,15</sup> and the United States<sup>16,17</sup>. However, by far the most complete and scientifically influential Devonian tetrapod material is still that of *Ichthyostega* and *Acanthostega* from East Greenland<sup>1,2,11,18-31</sup>, followed by *Ventastega* from Latvia<sup>6,31,32</sup> and *Tulerpeton* from Russia<sup>4,33,34</sup>. All other Devonian tetrapods are far less complete.

*Ichthyostega*, *Acanthostega*, *Ventastega* and *Tulerpeton* all date to the late Famennian, the last stage of the Devonian, when tetrapods had already been in existence for about 30 million years, judging by the trackway evidence<sup>35,36</sup>, and had colonised both equatorial and polar environments<sup>13</sup>. The four genera are quite disparate, hinting at long evolutionary histories; the differences between *Ichthyostega* and *Acanthostega* are particularly striking, including braincase morphologies so different it seems improbable that one could be derived from the other<sup>20</sup>.

The tetrapod material described here expands our understanding of the earliest stages of tetrapod evolution. It is securely dated to the earliest Famennian but is comparable to *Ventastega* in degree of completeness. Its source, the Sosnogorsk Formation of the southern part of Timan Ridge (Komi Republic, Russia)<sup>37</sup>, straddles the Frasnian-Famennian boundary with vertebrate remains occurring in the Famennian part (Extended Data Fig. 1). It is thus only marginally younger than the fragmentary genera *Elginerpeton*, *Obruchevichthys* and *Webererpeton*, which are the oldest tetrapod body fossils<sup>7,12</sup>. The quality of the material, which consists of numerous isolated bones and some articulated skull regions, is excellent. Multiple examples of the same bone all show the same distinctive features (Extended Data Fig. 2), indicating that only a single tetrapod species is present, meaning that all the data can be pooled into one interpretation (Extended Data Fig. 3). The Sosnogorsk fossils thus give us the first detailed picture of an animal from the earliest part of the known tetrapod body fossil record.

## **Systematic palaeontology**

Tetrapoda Jaekel, 1909

*Parmastega aelidae* gen. et sp. nov.

67 **Remark.** The term Tetrapoda is used here in its traditional, apomorphy-based sense  
68 of limbed vertebrates.

69 **Etymology.** The generic name derives from *parma*, a word in the Komi language  
70 describing the landscape of hills covered by coniferous forest, typical for South  
71 Timan, and Greek *stégi* meaning roof, understood here as skull roof. The specific  
72 name honours Associate Professor of Syktyvkar State University Dr. Aelida I. Popova  
73 (1929-2011), who first aroused PB's interest in natural sciences when he was a  
74 preschool boy.

75 **Holotype.** IG KSC 705/1, an articulated snout region (Fig. 1a-c).

76 **Referred material.** 106 individual bones or bone assemblies (Supplementary Table  
77 1).

78 **Locality and horizon.** Sosnovskiy Geological Monument, right bank of the Izhma  
79 River opposite Sosnogorsk, Komi Republic, Russia; Sosnogorsk Formation,  
80 lowermost Famennian (Extended Data Fig. 1).

81 **Diagnosis.** A stem tetrapod diagnosed by the following unique combination of  
82 characters: dermal ornament of preorbital region developed into transverse parallel  
83 'wave crests' with a spacing of a few millimetres; ornament present on dorsal blade of  
84 cleithrum and on anocleithrum; orbit strongly raised above skull roof, framed by an  
85 anterodorsal crest and a vertical anterior ridge carried on prefrontal; internasal  
86 fontanelle absent; median rostral paired; lacrimal excluded from orbit by prefrontal-  
87 jugal contact; intertemporal absent; pterygoids separated in midline by parasphenoid;  
88 interpterygoid vacuities absent; pterygoid dentition restricted to two lines of denticles,  
89 running anteriorly and anterolaterally from growth centre; ectopterygoid making large  
90 contribution to lateral wall of subtemporal fossa; middle part of otic capsule narrow,  
91 occupying approximately half of skull table width; posttemporal fossa wide,  
92 triangular; fang pair and row of marginal teeth on adsymphysial plate; middle part of  
93 prearticular with large muscle scar; interclavicle rounded with short posterior process.

94

## 95 **Description**

96 The material of *Parmastega* comprises the entire dermal skull apart from the  
97 preopercular and the posterior part of the quadratojugal, the entire ethmoid and dorsal  
98 part of the otoccipital braincase, the entire lower jaw, the dermal pectoral girdle  
99 (comprising, from dorsal to ventral, anocleithrum, cleithrum, clavicle and  
100 interclavicle) and the partly ossified scapulocoracoid (Figs 1-2). A total of 106

101 numbered specimens (Supplementary Table 1, 2), representing a minimum of 11  
102 individuals, show a wide size range (Extended Data Figs 2, 4) but were all found  
103 within a small area of the site (Extended Data Fig. 1). Most specimens are isolated  
104 bones, but an articulated ethmoid (Fig. 1a-c) and several skull tables (Fig. 1d-g) are  
105 also present. The bones are three-dimensionally preserved in limestone, with little or  
106 no distortion, and have been freed from the matrix using dilute acetic acid (see  
107 Methods). Bones from the same individual can sometimes be identified by matching  
108 size and sutural fit (Extended Data Fig. 3). This allows us to reconstruct the skull,  
109 lower jaw and pectoral girdle with a high degree of confidence, excepting only the  
110 posterior part of the suspensorium (Fig. 3). Assuming proportions similar to  
111 *Acanthostega*<sup>19</sup>, the maximum length of *Parmastega* was approximately 130 cm.

112 The skull shape is broadly similar to that of *Ventastega* and *Acanthostega*,  
113 although the orbits of *Parmastega* are raised higher above the skull table and the  
114 snout has a distinctly concave profile (Extended Data Fig. 4). The strongly raised  
115 orbits and relatively narrow snout are reminiscent of the elpistostegids *Elpistostege*  
116 and *Tiktaalik*<sup>38,39</sup>. However, the orbits are proportionately larger than in elpistostegids  
117 (Extended Data Fig. 5).

118 The dermal bone pattern of the skull roof and cheeks is, with a single  
119 exception, characteristic of Devonian tetrapods. There is no postrostral mosaic or  
120 internasal fontanelle. The median rostral is paired, as in *Acanthostega*, *Ventastega* and  
121 *Elpistostege*, but unlike *Ichthyostega* and *Elginerpeton* where it is single<sup>7,18,26,32,38</sup>. A  
122 tectal bone forms the dorsal margin of the naris, which lies very close to the jaw  
123 margin and faces ventrally; the ventral margin of the naris is formed by the maxilla as  
124 there is no lateral rostral. The lacrimal is excluded from the orbit by a long suture  
125 between the jugal and prefrontal. The latter is elongate and carries two bony crests,  
126 one forming the anterior part of the 'eyebrow' and the other an oblique ridge in front  
127 of the orbit, both more strongly developed in large specimens (Fig. 1m, 3a-c). The  
128 frontals are elongate with a distinct transverse 'step' on the posterior part of the dorsal  
129 surface marking the transition from snout to skull table. Intertemporals are absent.  
130 The lateral margins of the supratemporal and tabular form a raised spiracular margin;  
131 the tabular horn has distinct dorsal and ventral components. A small part of the dorsal  
132 surface of the braincase is exposed posterior to the tabulars. The dermal ornament of  
133 the preorbital region includes areas of irregular transverse ripples (Fig. 1h, m;  
134 Extended Data Fig. 2), somewhat similar to the ornament of *Umzantsia*<sup>13</sup> but much



135 coarser; elsewhere it grades into conventional tetrapod 'starburst' ornament. Partly  
136 enclosed sensory line canals are well developed on the premaxilla, cheek bones and  
137 anterior part of the nasals, but are absent from the skull table (Fig. 1d).

138       Between the anterior suture for the jugal and the posterior suture for the  
139 preopercular, the ventral margin of the squamosal presents two distinct sutural  
140 margins that appear to be contacts for two bones (Fig. 1l). The posterior of these must  
141 be for the quadratojugal; given that the jugal lacks a posterior process, we tentatively  
142 infer that the anterior segment of the ventral margin contacts the maxilla (Fig. 3a). A  
143 squamosal-maxillary contact is characteristic for 'fish' members of the tetrapod stem  
144 group such as *Eusthenopteron*<sup>40</sup>; its presence in *Parmastega* is unique for tetrapods.

145       The palatal morphology of *Parmastega* is intermediate between those of  
146 elpistostegids and Devonian tetrapods. In the elpistostegids *Panderichtys* and  
147 *Tiktaalik*, the pterygoids are separated in the midline by a long denticulated  
148 parasphenoid<sup>41,42</sup>. The vomer has a transverse posterior margin, which in  
149 *Panderichthys* ends mesially in a short posterior process extending along the lateral  
150 margin of the parasphenoid<sup>41</sup>. This condition is broadly similar to that in  
151 *Eusthenopteron*<sup>40</sup>. By contrast, in *Ichthyostega*, *Acanthostega* and *Ventastega* the  
152 pterygoids meet in the midline, separating the parasphenoid from the vomers, and the  
153 most posterior point of the vomer is its posterolateral corner<sup>6,18,23</sup>. In *Parmastega* the  
154 parasphenoid separates the pterygoids, but is not denticulated anteriorly, and the  
155 vomeral morphology is intermediate (Fig. 1a, 3d). The pterygoid carries a longitudinal  
156 row or narrow band of denticles, and a shorter oblique band extending anterolaterally.  
157 Uniquely, the ectopterygoid extends posteriorly past its contact with the pterygoid to  
158 contribute to the lateral margin of the subtemporal fossa (Fig. 3d). This relationship is  
159 demonstrated by a sutural fit of three bones from one individual (Fig. 1p).

160       Two parts of the braincase are preserved: the ethmoid and part of the sphenoid  
161 in IG KSC 705/1 and the dorsal part of the otoccipital in IG KSC 705/17 (Fig. 1a,f-g).  
162 An ossified ethmoid is only shared with *Ichthyostega* among known Devonian  
163 tetrapods<sup>18</sup>. The otoccipital has a strongly developed prootic buttress, a narrow cranial  
164 cavity with small inner ears, and a posttemporal fossa bounded laterally by a crista  
165 parotica that extends onto the tabular horn. Its outline in ventral view resembles  
166 *Tiktaalik*<sup>42</sup> but is proportionately broader. Previously known Devonian tetrapod  
167 otoccipitals show two radically different morphologies. In *Acanthostega* and  
168 *Ventastega* the narrow posttemporal fossa is open laterally and the braincase occupies

almost the whole ventral surface of the skull table, whereas in *Ichthyostega* the narrow braincase is flanked by large cavities under the skull table that probably housed spiracular diverticula<sup>20,24,25,32</sup>. The otoccipital of *Parmastega* provides a plausible ancestral ground plan for both these morphologies (Extended Data Fig. 6).

The lower jaw is of typical tetrapod construction<sup>30</sup> but unusually slender and delicate (Fig. 2a-h, 3e). The only ossified parts of the Meckelian element are the articular and the symphysis. The prearticular carries very few denticles but bears a large ventral muscle scar on its middle part. Remarkably, the contact between the prearticular and the mesial lamina of the splenial is not a tight suture as in other known Devonian tetrapods<sup>30</sup> but a loose overlap that must have contained a ligamentous component and allowed a degree of flexibility. Fang pairs, positioned mesial to the tooth row, are present on the adsymphysial plate, dentary, and anterior and middle coronoids. Postsplenial and surangular pit lines are present. The dentary is splint-like and loosely attached.

The pectoral girdle is U-shaped in anterior view with the dorsal blades of the cleithra approximately parallel (Fig. 2i-o, 3a,c). The dorsal orientation of the anocleithrum, determined from well-preserved contact surfaces, makes the girdle surprisingly tall. Cleithrum and anocleithrum both carry dermal ornament, a characteristic otherwise absent in tetrapods except *Umzantsia*<sup>13</sup>. The clavicle is narrow and the interclavicle has a rounded corpus with a short posterior process (Fig. 2n,o); both bones somewhat resemble the corresponding elements in *Ichthyostega*<sup>18</sup>, whereas *Acanthostega* and *Ventastega* have broader clavicles and kite-shaped interclavicles<sup>29,32</sup>. The scapulocoracoid is ossified in two parts: a dorsal scapular part coossified with the cleithrum (Fig. 3i), and a posterior coracoid ossification that carries the glenoid (Fig. 3p). As in *Ichthyostega*, *Elginerpeton* and *Hynerpeton*, the subscapular fossa is deep with a narrow apex; in *Acanthostega* and *Ventastega*, by contrast, the fossa is shallow and broad<sup>5,18,29,32,43</sup>. Limbs, pelvis, vertebrae and ribs are not preserved.

## Phylogenetic analysis

The phylogenetic position of *Parmastega* was evaluated with maximum parsimony and Bayesian inference analyses applied to a data matrix of 26 taxa and 113 characters. A full account of the tree search settings and results is given in **Methods**;

character list and data matrix are provided in Supplementary Information files 2-4.  
Trees are shown in Extended Data Fig. 7.

The resolution of the strict consensus unweighted parsimony analysis was poor: all Devonian tetrapods including *Parmastega* formed a polytomy together with 'whatcheeriid-grade' Carboniferous taxa (Extended Data Fig. 7a). However, in 70% of the trees, *Parmastega* was the sister group to all other tetrapods. A range of different approaches (character reweighting by Rescaled Consistency Index and K values; calculation of agreement subtrees from consensus trees) was used to investigate the phylogenetic signal in the data set (Extended Data Fig. 7b-c,e-h). This revealed consistent patterns. If the position of *Parmastega* was resolved, it was always placed as the sister group to all other tetrapods; if *Ventastega* was resolved, it was placed immediately crownward to *Parmastega*. *Ichthyostega* was resolved crownward to *Acanthostega* in the Adams consensus of unweighted trees, but in the reweighted analyses *Acanthostega* was crownward to *Ichthyostega*. The Bayesian tree (Extended Data Fig. 7d) also recovered these positions for *Parmastega* and *Ventastega*, but failed to resolve *Ichthyostega* and *Acanthostega*.

## Discussion

In essence, *Parmastega* is morphologically intermediate between the elpistostegids *Tiktaalik*, *Elpistostege* and *Panderichthys* on the one hand, and previously known Devonian tetrapods on the other. However, the mosaic of primitive and derived characters is not evenly distributed across the anatomy. The morphology of the lower jaw and the pectoral girdle is entirely tetrapod-like, as are the external dermal bone pattern of the snout region, the absence of gular plates, and the relative size of the orbits, whereas elpistostegid-like characteristics persist in the construction of the palate and the dermal ornamentation of the cleithrum and anocleithrum. This pattern hints at a sequence of evolutionary steps. Although no appendage bones are known, the morphology of the pectoral girdle strongly suggests that *Parmastega* possessed limbs rather than paired fins. Particularly significant is the scapulocoracoid, which forms the proximal attachment for many forelimb muscles and undergoes substantial shape change from elpistostegids<sup>44,45</sup> to tetrapods<sup>5,18,29,32,34</sup>. The scapulocoracoid of *Parmastega* conforms fully to the tetrapod pattern. The shape and construction of the lower jaw, and the absence of gular plates, suggest that gill ventilation and prey capture worked in the same way as in more crownward Devonian tetrapods. The

reconfiguration of the palate and the loss of dermal ornament on the shoulder girdle evidently lagged behind these transformations.

Until now, one of the most puzzling aspects of Devonian tetrapod anatomy has been the specialised ear region of *Ichthyostega*, which differs so much from those of other early tetrapods that it has been challenging to establish detailed homologies<sup>18,20</sup>. *Parmastega* partly resolves this problem by presenting a braincase morphology that is intermediate between *Ichthyostega* on the one hand and *Acanthostega* and *Ventastega* on the other, providing a plausible hypothetical ancestor for both patterns (Extended Data Fig. 6a). However, these transformations cannot be mapped parsimoniously onto the phylogeny, indicating the presence of non-trivial homoplasy either in the braincases or in other parts of the skeleton (Extended Data Fig. 6b).

The three-dimensional preservation and apparent absence of post-mortem transport makes the *Parmastega* fossils palaeobiologically informative. The environment of preservation, which was almost certainly also the living environment of *Parmastega*, was a coastal lagoon with brackish water and a rich fish fauna including the placoderm *Bothriolepis* and various sarcopterygians<sup>46</sup>. The concentration of the tetrapod remains to a small area of the site (Extended Data Fig. 1) suggests that *Parmastega* may have been a schooling animal. The vertebrate-bearing bed, Bed 40 (the “fish dolomite”), is composed of two consecutive tempestites; possibly a school of *Parmastega* was killed by the first storm event and their skeletons partly disarticulated by the second. Schooling behavior is also implied by the mass occurrence of *Acanthostega* on Stensiö Bjerg, East Greenland<sup>47</sup>.

Raised orbits and a lack of lateral line canals on the skull table in *Parmastega* (Fig. 3a) suggests a surface-skimming position in the water, with emergent eyes, similar to crocodilians (Extended Data Fig. 8)<sup>47</sup>. The increase in orbit size across the fish-tetrapod transition has been linked to a shift from aquatic to aerial vision<sup>48</sup>; the relative orbit size of *Parmastega* falls well within the tetrapod range (Extended Data Fig. 5) and its eyes were thus probably adapted for use in air. Although all known Devonian tetrapods have dorsally positioned eyes, *Parmastega* shows the most extreme version of this condition (Extended Data Fig. 4). The nostrils of *Parmastega* face ventrally, indicating that the nose was not used for air-breathing while resting at the surface (Extended Data Fig. 8). We suggest that the dorsally placed spiracles took on this function, as previously argued for *Panderichthys*<sup>49</sup> and more crownward Devonian tetrapods<sup>20, 50</sup>. The lower jaw does not match the upper jaw in curvature,

270 either in lateral or ventral view (Extended Data Fig. 9a,d). This pattern is also seen in  
271 *Ventastega* (Extended Data Fig. 9b), *Acanthostega*<sup>31</sup> and *Ichthyostega*<sup>18</sup>.

272 Surprisingly, the *Parmastega* material contains no vertebrae, ribs, pelvic  
273 girdles or limb bones. The lack of evidence for post-mortem transport, the partially  
274 ossified nature of the scapulocoracoid even in the largest individuals, and the  
275 preservation of the delicate isolated coracoid ossifications (Fig. 2i-l,p), suggests that  
276 this absence is not a taphonomic artefact but reflects a very lightly ossified or even  
277 cartilaginous axial and appendicular skeleton. *Ventastega* may also have had a lightly  
278 ossified postcranial skeleton<sup>32</sup>. *Acanthostega* and *Ichthyostega* became fully ossified  
279 as adults<sup>2,18,19,21,27,29</sup>, but *Acanthostega* appears to have had a long juvenile stage with  
280 unossified endoskeleton<sup>47</sup>. Functionally, the poor ossification of *Parmastega* suggests  
281 little or no capacity for terrestrial locomotion. However, it contrasts strangely with the  
282 cranial morphology, which suggests that the eyes were habitually held above the  
283 surface of the water and thus implies some kind of engagement with the terrestrial  
284 environment. Even more puzzling is the fact that this poorly ossified postcranial  
285 skeleton is apomorphic: elpistostegids are well ossified, as are the majority of  
286 tetrapodomorph fishes<sup>39,40</sup>.

287 *Parmastega* gives us the earliest detailed glimpse of a tetrapod: an aquatic,  
288 surface-skimming predator, just over a metre in length, living in a lagoon on a tropical  
289 coastal plain. It is phylogenetically least crownward of the non-fragmentary tetrapods,  
290 but is not necessarily representative of primitive conditions for the group. The slightly  
291 earlier *Elginerpeton*, which was also probably aquatic and even larger than  
292 *Parmastega* (Extended Data Fig. 3), had well ossified girdles and limb bones as well  
293 as a distinctive head shape with a narrow snout<sup>7,30,43</sup>. Moreover, the trackway record  
294 shows that tetrapods originated at least 20 million years before *Parmastega*<sup>35,36</sup>, and  
295 the very existence of the trackways – which implies weight-bearing limbs, even if the  
296 prints were made in water – points to these forms having well ossified postcranial  
297 skeletons. Together with the evidence for significant morphological homoplasy  
298 among Devonian tetrapods, this hints at a tangled and still elusive early history for  
299 limbed vertebrates.

300

301 **Acknowledgements**

302 We thank Dr. Yuriy Gatovsky (Moscow), Dr. Andrey Zhuravlev (Syktyvkar) and Dr.  
303 Dmitry Ponomarev (Syktyvkar) for their support of the project, and the dig crews of  
304 the 2009-2012 excavations for all their hard work. Dr. Alexander Ivanov (St.  
305 Petersburg) identified the first tetrapod mandible from Sosnogorsk, in the Chernyshov  
306 Collection. PB acknowledges the support of National Geographic Society Grant 9099-  
307 12 and UNDP/GEF Project No. 00059042. EL acknowledges the support of Latvian  
308 Council of Science Grant Z-6153-110. PEA acknowledges the support of a  
309 Wallenberg Scholarship from the Knut and Alice Wallenberg Foundation.

310

#### 311 **Author contributions**

312 PB initiated and directed the excavation program at Sosnogorsk that produced the  
313 material for the study. EL and PEA participated in excavations. PB carried out all  
314 preparation, consolidation and photography of specimens. PEA made the  
315 reconstructions of the skull, lower jaw and shoulder girdle. MR performed the  
316 phylogenetic analyses. PEA made Figures 1-3 and Extended Data Figures 2 and 3-9;  
317 PB made Extended Data Figures 1 and 3, as well as Tables 1 and 2. All authors  
318 participated in the interpretation of the material and the writing of the paper.

319

320

321

#### 322 **References**

323 1. Säve-Söderbergh, G. Preliminary note on Devonian stegocephalians from East  
324 Greenland. *Meddel. Grønland* **94**, 1–107 (1932).

325

- 326 2. Jarvik, E. On the fish-like tail in the ichthyostegid stegocephalians with  
327 descriptions of a new stegocephalian and a new crossopterygian from the Upper  
328 Devonian of East Greenland. *Meddel. Grønland* **114**, 1–90 (1952).  
329
- 330 3. Campbell, K. S. W. & Bell, M. W. A primitive amphibian from the Late Devonian  
331 of New South Wales. *Alcheringa* **1**, 369–381 (1977).  
332
- 333 4. Lebedev, O. A. The first find of a Devonian tetrapod in USSR. *Doklady Acad.*  
334 *Nauk SSSR* **278**, 1470–1473 [in Russian] (1984).  
335
- 336 5. Daeschler, E. B., Thomson, K. S. & Amaral, W. W. A Devonian tetrapod from  
337 North America. *Science* **265**, 639–642 (1994).  
338
- 339 6. Ahlberg, P. E., Lukševičs, E. & Lebedev, O. The first tetrapod finds from  
340 the Devonian (Upper Famennian) of Latvia. *Phil. Trans. R. Soc. B* **343**, 303–328  
341 (1994).  
342
- 343 7. Ahlberg, P. E. *Elginerpeton pancheni* and the earliest tetrapod clade. *Nature* **373**,  
344 420–425 (1995).  
345
- 346 8. Daeschler, E. B. Early tetrapod jaws from the Late Devonian of Pennsylvania,  
347 USA. *J. Paleontol.* **74**, 301–308 (2000).  
348
- 349 9. Zhu, M., Ahlberg, P. E., Zhao, W. & Jia, L. First Devonian tetrapod from Asia.  
350 *Nature* **420**, 760–761 (2002).

351

352 10. Lebedev, O. A. A new tetrapod *Jakubsonia livnensis* from the Early Famennian  
353 (Devonian) of Russia and palaeoecological remarks on the Late Devonian tetrapod  
354 habitats. *Acta Univ. Latviensis* **679**, 79–98 (2004).

355

356 11. Clack, J. A., Ahlberg, P. E., Blom, H. & Finney, S. M. A new genus of Devonian  
357 tetrapod from North-East Greenland, with new information on the lower jaw of  
358 *Ichthyostega*. *Palaeontology* **55**, 73–86 (2012).

359

360 12. Clément, G. & Lebedev, O. A. Revision of the early tetrapod *Obruchevichthys*  
361 Vorobyeva, 1977 from the Frasnian (Upper Devonian) of the North-western East  
362 European Platform. *Paleontol. J.* **48**, 1082–1091 (2014).

363

364 13. Gess, R. & Ahlberg, P. E. A tetrapod fauna from within the Devonian Antarctic  
365 Circle. *Science* **360**, 1120–1124 (2018).

366

367 14. Clément, G., Ahlberg, P. E., Blicek, A., Blom, H., Clack, J. A., Poty, E., Thorez,  
368 J. & Janvier, P. Devonian tetrapod from Western Europe. *Nature* **427**, 412–413  
369 (2004).

370

371 15. Olive, S., Ahlberg, P. E., Pernègre, V., Poty, E., Steurbaut, E. & Clément, G. New  
372 discoveries of tetrapods (ichthyostegid-like and whatcheeriid-like) in the Famennian  
373 (Late Devonian) localities of Strud and Becco (Belgium). *Palaeontology* **59**, 827–840  
374 (2016).

375



- 376 16. Shubin, N. H., Daeschler, E. B. & Coates, M. I. The early evolution of the  
377 tetrapod humerus. *Science* **304**, 90–93 (2004).  
378
- 379 17. Daeschler, E. B., Clack, J. A. & Shubin, N. H. Late Devonian tetrapod remains  
380 from Red Hill, Pennsylvania, USA: how much diversity? *Acta Zool.* **90**(Suppl. 1),  
381 306–317 (2009).  
382
- 383 18. Jarvik, E. The Devonian tetrapod *Ichthyostega*. *Fossils & Strata* **40**, 1–206 (1996).  
384
- 385 19. Ahlberg, P. E., Clack, J. A. & Blom, H. The axial skeleton of the Devonian  
386 tetrapod *Ichthyostega*. *Nature* **437**, 137–140 (2005).  
387
- 388 20. Clack, J. A., Ahlberg, P. E., Finney, S. M., Dominguez Alonso, P., Robinson, J. &  
389 Ketcham, R. A. A uniquely specialized ear in a very early tetrapod. *Nature* **425**, 66–  
390 69 (2003).  
391
- 392 21. Callier, V., Clack, J. A. & Ahlberg, P. E. Contrasting developmental trajectories  
393 in the earliest known tetrapod forelimbs. *Science* **324**, 364–367 (2009).  
394
- 395 22. Clack, J. A. Discovery of the earliest tetrapod stapes. *Nature* **342**, 425–430  
396 (1989).  
397
- 398 23. Clack, J. A. *Acanthostega gunnari*, a Devonian tetrapod from Greenland: the  
399 snout, palate and ventral parts of the braincase. *Meddel. Grønland Geoscience* **31**, 1–  
400 24 (1994).

- 401
- 402 24. Clack, J. A. Earliest known tetrapod braincase and the evolution of the stapes and  
403 fenestra ovalis. *Nature* **369**, 392–394 (1994).
- 404
- 405 25. Clack, J. A. The neurocranium of *Acanthostega gunnari* Jarvik and the evolution  
406 of the otic region in tetrapods. *Zool. J. Linn. Soc.* **122**, 61–97 (1998).
- 407
- 408 26. Clack, J. A. A revised reconstruction of the dermal skull roof of *Acanthostega*  
409 *gunnari*, an early tetrapod from the Late Devonian. *Trans. R. Soc. Edinb. Earth Sci.*  
410 **93**, 163–165 (2002).
- 411
- 412 27. Coates, M. I. & Clack, J. A. Polydactyly in the earliest known tetrapod limbs.  
413 *Nature* **347**, 66–69 (1990).
- 414
- 415 28. Coates, M. I. & Clack, J. A. Fish-like gills and breathing in the earliest known  
416 tetrapod. *Nature* **352**, 234–236 (1991).
- 417
- 418 29. Coates, M. I. The Devonian tetrapod *Acanthostega gunnari* Jarvik: postcranial  
419 anatomy, basal tetrapod interrelationships and patterns of skeletal evolution. *Trans. R.*  
420 *Soc. Edinb. Earth Sci.* **87**, 363–421 (1996).
- 421
- 422 30. Ahlberg, P. E. & Clack, J. A. Lower jaws, lower tetrapods - a review based on the  
423 Devonian genus *Acanthostega*. *Trans. R. Soc. Edinb. Earth Sci.* **89**, 11–46 (1998).
- 424
- 425 31. Porro, L. B., Rayfield, E. J. & Clack, J. A. 2015. Descriptive anatomy and three-

dimensional reconstruction of the skull of the early tetrapod *Acanthostega gunnari*  
Jarvik, 1952. *PLoS ONE* **10**, e0118882 (2015).

32. Ahlberg, P. E., Clack, J. A., Lukševičs, E., Blom, H. & Zupins, I. *Ventastega*  
*curonica* and the origin of tetrapod morphology. *Nature* **453**, 1199–1204 (2008).

33. Lebedev, O. A. & Clack, J. A. Upper Devonian tetrapods from Andreyevka, Tula  
Region, Russia. *Palaeontology* **36**, 721–734 (1993).

34. Lebedev, O. A. & Coates, M. I. The postcranial skeleton of the Devonian tetrapod  
*Tulerpeton curtum*. *Zool. J. Linn. Soc.* **114**, 307–348 (1995).

35. Niedzwiedzki, G., Szrek, P., Narkiewicz, K., Narkiewicz, M. & Ahlberg, P. E.  
Tetrapod trackways from the early Middle Devonian period of Poland. *Nature* **463**,  
43–48 (2010).

36. Stössel, I., Williams, E. A. & Higgs, K. T. Ichnology and depositional  
environment of the Middle Devonian Valentia Island tetrapod trackways, south-west  
Ireland. *Palaeogeog., Palaeoclimatol., Palaeoecol.* **462**, 16–40 (2016).

37. Beznosov, P. Sosnogorsk Formation – a new local stratigraphic unit of the Upper  
Devonian from South Timan. *Geologiya i mineral'nye resursy evropeyskogo servero-*  
*vostoka Rossii: Materialy XV geologicheskogo s'yezda Respubliki Komi* **2**, 9–12 [in  
Russian] (2009).

- 451 38. Schultze, H.-P. & Arsenault, M. The panderichthyid fish *Elpistostege*: a close  
452 relative of tetrapods? *Palaeontology* **28**, 293–310 (1985).  
453
- 454 39. Daeschler, E. B., Shubin, N. H. & Jenkins, F. A. A Devonian tetrapod-like fish  
455 and the evolution of the tetrapod body plan. *Nature* **440**, 757–763 (2006).  
456
- 457 40. Jarvik, E. *Basic Structure and Evolution of Vertebrates* Vol. 1 (Academic Press,  
458 London, 1980).  
459
- 460 41. Ahlberg, P. E., Clack, J. A. & Lukševičs, E. Rapid braincase evolution between  
461 *Panderichthys* and the earliest tetrapods. *Nature* **381**, 61–64 (1996).  
462
- 463 42. Downs, J. P., Daeschler, E. B., Jenkins, F. A. Jr & Shubin, N. H. The cranial  
464 endoskeleton of *Tiktaalik roseae*. *Nature* **455**, 925–929 (2008).  
465
- 466 43. Ahlberg, P. E. Postcranial stem tetrapod remains from the Devonian of Scat Craig,  
467 Morayshire, Scotland. *Zool. J. Linn. Soc.* **122**, 99–141 (1998).  
468
- 469 44. Vorobyeva, E. I. The shoulder girdle of *Panderichthys rhombolepis* (Gross)  
470 (Crossopterygii); Upper Devonian; Latvia. *Geobios* **28**(Suppl. 2), 285–288 (1995).  
471
- 472 45. Shubin, N. H., Daeschler, E. B. & Jenkins, F. A. The pectoral fin of *Tiktaalik*  
473 *rosae* and the origin of the tetrapod limb. *Nature* **440**, 764–771 (2006).  
474
- 475 46. Lukševičs, E., Beznosov, P. & Stūris, V. A new assessment of the Late Devonian

476 antiarchan fish *Bothriolepis leptocheira* from South Timan (Russia) and the biotic  
 477 crisis near the Frasnian–Famennian boundary. *Acta Palaeontol. Pol.* **62**, 97–119  
 478 (2017).  
 479  
 480 47. Sanchez, S., Tafforeau, P., Clack, J. A. & Ahlberg, P. E. Life history of the stem  
 481 tetrapod *Acanthostega* revealed by synchrotron microtomography. *Nature* **537**, 408–  
 482 411 (2016).  
 483  
 484 48. MacIver, M. A., Schmitz, L., Mugan, U., Murphey, T. & Mobley, C. D. Massive  
 485 increase in visual range preceded the origin of terrestrial vertebrates. *Proc. Natl.*  
 486 *Acad. Sci. USA* **114**, E2375–84 (2017).  
 487  
 488 49. Brazeau, M. D. & Ahlberg, P. E. Tetrapod-like middle ear architecture in a  
 489 Devonian fish. *Nature* **439**, 318–21 (2006).  
 490  
 491 50. Clack, J. A. Discovery of the earliest known tetrapod stapes. *Nature* **342**, 425–427  
 492 (1989).  
 493

494 **Figure 1 | *Parmastega aelidae*: skull roof, cheek and palate.** **a-c**, IG KSC 705/1,  
 495 holotype of *Parmastega aelidae*; an articulated ethmosphenoid with associated  
 496 prefrontal in ventral (**a**), dorsal (**b**) and lateral (**c**) views. The 10 mm scale bar of this  
 497 specimen applies to the whole figure except **f-g**. **d-e**, 705/2, skull table in dorsal (**d**)  
 498 and ventral (**e**) views. **f-g**, 705/17, skull table and partial braincase in ventral view. **g**  
 499 is a false colour image identifying the components of the specimen. **h**, 705/18, right  
 500 frontal, dorsal view. **i**, 705/19, left postorbital, external view. **j**, 705/20, left jugal,  
 501 external view. **k**, 705/25, left lacrimal, lateral (top) and dorsal (bottom) views. **l**,  
 502 705/26, right squamosal, external view. **m**, 705/5, right prefrontal, external view. **n**,  
 503 705/4, left postfrontal, lateral (top) and dorsal (bottom) views. **o**, 705/28, right maxilla  
 504 in internal (top), ventral (middle) and external (bottom) views. **p**, 705/29 (left  
 505 dermopalatine), 705/30 (ectopterygoid) and 705/31 (pterygoid) in ventral view. **q**,  
 506 705/32, left dermopalatine in lateral (top) and ventral (bottom) views. cho, choana; fr,  
 507 frontal; m.ro, median rostrals; na, nasal; pa, parietal; pi, pineal foramen; pmx,  
 508 premaxilla; pp, postparietal; prf, prefrontal; psp, parasphenoid; socc, supraoccipital;  
 509 su, supratemporal; ta, tabular; te, tectal; vo, vomer.

510

511

512 **Figure 2 | *Parmastega aelidae*: lower jaw and pectoral girdle.** **a**, IG KSC 705/21,  
 513 right adsymphysial plate in mesial (bottom) and dorsal (top) views. **b**, 705/22, right  
 514 anterior coronoid in mesial (bottom) and dorsal (top) views. **c**, 705/33, right middle  
 515 coronoid in mesial (bottom) and dorsal (top) views. **d**, 705/36, left posterior coronoid  
 516 in mesial (bottom) and dorsal (top) views. **e**, 705/37, articulated left splenial and  
 517 adsymphysial plate in ventrolateral (top) and mesial (bottom) views. **f**, 705/34,  
 518 articulated left postsplenial, angular and surangular in lateral view. **g**, 705/76, left

519 prearticular in mesial view. **h**, 705/67, right dentary in lateral (top), dorsal (middle)  
 520 and mesial (bottom) views. **i-k**, 705/15, left cleithrum and partial scapulocoracoid in  
 521 mesial (**i**), anterior (**j**) and lateral (**k**) views. **l**, 705/95 (right cleithrum) and 705/98  
 522 (anocleithrum) in lateral view. **m**, 705/98, right anocleithrum in lateral view. **n-o**,  
 523 705/92 (right clavicle) and 705/89 (interclavicle) in anterior (**n**) and ventral (**o**) views.  
 524 **p**, 705/102, left coracoid in lateral view. **e-p** are shown to the same scale.

525

526

527 **Figure 3 | *Parmastega aelidae*: reconstructions.** **a**, skull, lower jaw and pectoral  
 528 girdle of *Parmastega* in right lateral view. **b**, skull in dorsal view. **c**, skull and pectoral  
 529 girdle in anterior view. **d**, skull in ventral view. **e**, right lower jaw ramus in mesial  
 530 view. adsym, adsymphysial plate; an, anocleithrum; ang, angular; ant.cor, anterior  
 531 coronoid; art, articular; cho, choana; cla, clavicle; clei, cleithrum; cor, coracoid; de,  
 532 dentary; dpal, dermopalatine; ect, ectopterygoid; fr, frontal; gle, glenoid; ju, jugal; la,  
 533 lacrimal; mid.cor, middle coronoid; m.ro, median rostrals; mx, maxilla; na, nasal; no,  
 534 nostril; orb, orbit; ot.br, otoccipital braincase; pa, parietal; pi, pineal foramen; pmx,  
 535 premaxilla; po, postorbital; pof, postfrontal; pospl, postsplenial; post.cor, posterior  
 536 coronoid; pp, postparietal; prf, prefrontal; psp, parasphenoid; pter, pterygoid; qj,  
 537 quadratojugal; scap, scapula; socc, supraoccipital; spl, splenial; sq, squamosal; su,  
 538 supratemporal; suf, subtemporal fossa; sur, surangular; ta, tabular; te, tectal; vo,  
 539 vomer. Vertical hatching indicates missing element with unknown outline, horizontal  
 540 hatching damaged object with known outline. Scale of reconstruction determined by  
 541 largest individual. **a-d** are shown to the same scale.

542

## 543   **Methods**

544

### 545   **Preparation and illustration of specimens**

546   The specimens were collected from the Sosnovskiy Geological Monument, right bank  
547   of the river Izhma opposite Sosnogorsk Town, Komi Republic, Russia, during a series  
548   of field seasons from 2002 to 2012. The bulk of the material was collected during the  
549   large-scale excavation in 2009-2012, when approximately 50 m<sup>2</sup> of the bone-bearing  
550   “fish dolomite” bed was dug out and then broken into small blocks using hammers,  
551   chisels, angle grinder, drill and portable jackhammer. Blocks containing parts of the  
552   same bone fragments glued together. The bones were freed from the limestone matrix  
553   using dilute (7-10 %) acetic acid alternating with drying and covering by consolidants  
554   PVB (before 2010) and Paraloid® B-72 (after 2010). The reconstructions of the skull  
555   and lower jaw were assembled by hand on the basis of photographs of individual  
556   bones, taken at appropriate angles. The pectoral girdle reconstruction was produced  
557   by sticking together the right anocleithrum, cleithrum, clavicle and interclavicle of  
558   one individual, making a three-dimensional virtual model of the assembly using  
559   photogrammetry (Agisoft PhotoScan), and importing this model into 3-matic  
560   (Materialise) where it was duplicated, mirrored and assembled into a complete girdle.  
561   The drawings of the girdle in Fig. 4 were traced directly from lateral and anterior  
562   projections of the model.

563

### 564   **Phylogenetic analysis**

565   The phylogenetic position of *Parmastega* was evaluated with maximum parsimony  
566   and Bayesian inference analyses applied to a data matrix of 26 taxa and 113  
567   characters (Supplementary Files 1-3), based on a recent matrix published by Chen et



568 al.<sup>51</sup> with the addition of four new characters (nos. 7, 27, 28, 29). Prior to all analyses,  
569 we explored the occurrence of possible “taxonomic equivalents”<sup>52</sup> by subjecting the  
570 matrix to safe taxonomic reduction using the *Claddis* package<sup>53</sup> in the R environment  
571 for statistical computing and graphics (<https://cran.r-project.org>). No taxon was  
572 identified as being suitable for safe deletion.

573 For all parsimony analyses, we used PAUP\* version 4.0a (build 164)<sup>54</sup> with  
574 the following search settings. The “collapse branch” option was enforced for branches  
575 possibly attaining a minimum length of zero. Tree searches employed a heuristic  
576 option with tree bisection-reconnection branch swapping algorithm, saving no more  
577 than a single tree of length greater than/equal to 1 step in each replicate, and using a  
578 maximum of 5000 random step-wise taxon addition replicates while holding a single  
579 tree in memory at each step. Following this initial round of tree searches, an  
580 additional branch-swapping round was conducted on all trees saved in memory, this  
581 time with the option of saving multiple trees in effect. This second round of tree  
582 searches was repeated 10 times. No shorter or additional trees were found at the end  
583 of this second round in any of the parsimony analyses. Three analyses were carried  
584 out under maximum parsimony, each with the settings specified above.

585 In the first analysis, all characters were treated as unordered and of equal unit  
586 weight. We obtained 23 shortest trees at 278 steps, with an ensemble consistency  
587 index (C.I.) of 0.5 (0.4908 excluding 5 parsimony-uninformative characters), an  
588 ensemble retention index (R.I.) of 0.6911, and an ensemble rescaled consistency  
589 index (R.I.) of 0.3456. A permutation-tail probability test<sup>55</sup> using 1000 replicates  
590 showed that the length of the shortest trees differed significantly from random ( $p \sim$   
591 0.001). The strict consensus (Fig. 5a) was poorly resolved. The Adams consensus  
592 (Fig. 5b) had greater resolution, placing *Parmastega* and *Elginerpeton* as the joint

593 (unresolved) sister groups to all other tetrapods. The agreement subtree (a pruned  
594 topology including only those taxa for which all most parsimonious trees agree upon  
595 mutual relationships) included 18 out of the 26 original taxa (Extended Data Fig. 7a;  
596 deleted: *Acanthostega*; *Dendrerpeton*; *Densignathus*; *Elginerpeton*; *Greererpeton*;  
597 *Metaxygnathus*; *Ossinodus*; *Tantallognathus*). Node support value was evaluated via  
598 bootstrapping<sup>56</sup> and jackknifing<sup>57</sup> in PAUP\*, in each case using 50% character  
599 resampling, and 50,000 random resampling replicates with the fast step-wise addition.  
600 In both cases, very few nodes receive support, namely post-*Panderichthys* taxa, post-  
601 elpistostegalian taxa, baphetids, and a clade of *Eoherpeton* plus *Proterogyrinus*.

602 In the second analysis, characters were re-weighted by the largest values of  
603 their rescaled consistency indexes from the initial analysis. PAUP\* yielded a single  
604 tree (Fig. 5c) 112.3561 steps long, with C.I. = 0.6804 (0.6655 excluding  
605 uninformative characters), R.I. = 0.8297, and R.C. = 0.5645. This tree was 3 steps  
606 longer than the trees from the unweighted analysis and did not represent a  
607 significantly better fit for the data, in terms of tree length, than the unweighted trees,  
608 based upon Templeton, Kishino-Hasegawa, and Winning-sites tests in PAUP\*. The  
609 weighted analysis confirmed the status of *Parmastega* as the most basal tetrapod.

610 In the third analysis, we used implied weighting<sup>58</sup>, experimenting with  
611 different integer values of Goloboff's constant of concavity K. We ran analyses with 1  
612  $\leq K \leq 10$  (e.g. ref. 59). For each K value, we saved all trees generated at the end of the  
613 analysis. The separate tree files obtained from all K-weighted analyses were stored in  
614 PAUP\* after filtering out duplicated tree topologies. This process resulted in 5 K-  
615 weighted trees, which were summarised with a strict consensus (Extended Data Fig.  
616 7b), an agreement subtree (Extended Data Fig. 7c), and an Adams consensus

617 (Extended Data Fig. 7d). The agreement subtree included 22 taxa (deleted:  
618 *Densignathus*; *Elginerpeton*; *Metaxygnathus*; *Ossinodus*).

619 For the Bayesian inference analysis, we employed MrBayes v. 3.2.6 (ref 60),  
620 with the following settings: variable coding; gamma-distributed rate model; 10<sup>7</sup>  
621 generations and four chains; discarding the first 25% of sampled trees. Convergence  
622 diagnostic was evaluated through inspection of the Potential Scale Reduction Factor  
623 values<sup>61</sup> output by MrBayes. These values approached or were identical to 1,  
624 indicating successfully convergent runs (Supplementary File 4). Credibility values for  
625 nodes in the Bayesian results (Fig. 5c) were moderate to strong for most nodes.

626

## 627 **References**

628 51. Chen, D., Alavi, Y., Brazeau, M. D., Blom, H., Millward, D. & Ahlberg, P. E. A  
629 partial lower jaw of a tetrapod from "Romer's Gap". *Earth Env. Sci. Trans. R. Soc.*  
630 *Edinb.* **108**, 55-65 (2018).

631

632 52. Wilkinson, M. Majority-rule reduced consensus trees and their use in  
633 bootstrapping. *Mol. Biol. Evol.* **13**, 437–444 (1996).

634

635 53. Lloyd, G. T. Estimating morphological diversity and tempo with discrete  
636 character-taxon matrices: implementation, challenges, progress, and future directions.  
637 *Biol. J. Linn. Soc.* **118**, 131–151 (2016).

638

639 54. Swofford, D. L. PAUP\*. Phylogenetic Analysis Using Parsimony (\*and Other  
640 Methods). Version 4. Sinauer Associates, Sunderland, Massachusetts (2003).

641

- 642 55. Wilkinson, M., Peres-Neto, P. R., Foster, P. G. & Moncrieff, C. B. Type 1 error  
643 rates of the parsimony permutation tail probability test. *Syst. Biol.* **51**, 524–527  
644 (2002).
- 645 56. Felsenstein, J. Confidence limits on phylogenies: An approach using the  
646 bootstrap. *Evolution* **39**, 783–791 (1985).
- 647 57. Farris, J. S., Albert, V. A., Källersjö, M., Lipscomb, D. & Kluge, A. G. Parsimony  
648 jackknifing outperforms neighbor-joining. *Cladistics* **12**, 99–124 (1996).
- 649 58. Goloboff, P. A. Estimating character weights during tree search. *Cladistics* **9**, 83–  
650 91 (1993).
- 651
- 652 59. Congreve, C. R. & Lamsdell, J. C. Implied weighting and its utility in  
653 palaeontological datasets: a study using modelled phylogenetic matrices.  
654 *Palaeontology* **59**, 447–462 (2016).
- 655
- 656 60. Ronquist, F. & Huelsenbeck, J. P. MRBAYES 3: Bayesian phylogenetic inference  
657 under mixed models. *Bioinformatics* **19**, 1572–1574 (2003).
- 658
- 659 61. Gelman, A. & Rubin, D. B. Inference from iterative simulation using multiple  
660 sequences. *Statistical science*, **7**(4), pp.457–472 (1992).
- 661
- 662 62. Robinson, J. *The evolution of the early tetrapod middle ear and associated*  
663 *structures*. PhD thesis, University College London, pp 435 (2006).
- 664
- 665 63. Ahlberg, P. E., Friedman, M. & Blom, H. New light on the earliest known

666 tetrapod jaw. *Journal of Vertebrate Paleontology* **25**, 720-724 (2005).

667

668 64. Blom, H. Taxonomic revision of the Late Devonian tetrapod *Ichthyostega* from

669 East Greenland. *Palaeontology* **48**, 111-134 (2005).

670

#### 671 **Author Information**

672 The authors declare that there are no competing interests. Correspondence to Per Erik

673 Ahlberg [per.ahlberg@ebc.uu.se](mailto:per.ahlberg@ebc.uu.se)

674

#### 675 **Data Availability**

676 In total, 132 specimens comprising 183 skeletal elements have been collected during

677 the entire period of excavations (2002-2012). 106 specimens, all of them figured in

678 Supplementary Table 1, have been deposited in the collection of the Institute of

679 Geology, Komi Science Centre, Ural Branch of the Russian Academy of Sciences,

680 Syktyvkar, Russia under the collection number IG KSC 705/ and are available for

681 examination. Other specimens have been reserved for sharing with other museums.

682 The LSID for *Parmastega* is urn:lsid:zoobank.org:act:76B5BB03-42FE-4F46-A284-

683 F95E973CEE96.

684

685

686

687 **Extended Data Figure 1 | The distribution of *Parmastega* at the Sosnogorsk fossil**  
 688 **site. a-b**, Maps of increasing resolution showing the location of Sosnogorsk within  
 689 northwest Russia. The box around Ukhta and Sosnogorsk in **a** indicates the region  
 690 shown in **b**. In **b**, the brown belt extending from north to south indicates the outcrop of  
 691 Famennian ( $D_3fm$ ) deposits in the region, and the yellow arrow points to the  
 692 Sosnogorsk fossil site (Sosnovskiy Geological Monument). **c**, Stratigraphic column  
 693 through the Sosnogorsk Formation and part of the overlying marine Izhma Formation.  
 694 Note the possible position of the Frasnian-Famennian boundary ( $D_3f / D_3fm$ ) in the  
 695 lower part of the Sosnogorsk Formation. The vertebrate-bearing part of the formation  
 696 is shown in detail on the right, with the tetrapod-bearing level indicated with a red  
 697 vertical bar. **d**, general view of outcrop #20 (Sosnovskiy Geological Monument) from  
 698 the opposite bank of the Izhma River. 1 - limestone, 2 - dolomite, 3 - clay, 4 - nodular  
 699 limestone, 5 - scree, 6 - landslide.  $D_3sn$  - Sosnogorsk Formation,  $D_3iž$  - Izhma  
 700 Formation. Distance A'-B' indicates the area of main excavation in 2010-2012. **e**,  
 701 main excavation. Distance A-B indicates the area where all tetrapod bones were found  
 702 during the excavation in 2012. The photo was taken on 2 August 2012. **f**, sketch-map  
 703 of the main excavation, 2012, showing the distribution of tetrapod bones within the  
 704 bed. The cluster numbers are indicated in orange.

705  
 706 **Extended Data Figure 2 | Frontal bones of *Parmastega*.** The figure shows all the  
 707 complete and near-complete frontals of *Parmastega* (8 out of 9 known frontals), to  
 708 scale, oriented with anterior at the top and aligned on the centre of radiation  
 709 (horizontal line). Right frontals have been reversed so that all bones have the  
 710 appearance of left frontals. From left to right the specimens are IG KSC 705/3

711 (reversed), 705/40, 705/44 (reversed), 705/43, 705/45, 705/18 (reversed), 705/42 and  
712 705/41. Scale bar, 10mm.

713

714 **Extended Data Figure 3 | Bone associations.** **a, b**, diagrammatic images showing, in  
715 orange, associated bones of two individual skulls. **a**, the holotype, IG KSC 705/1. **b**,  
716 the largest individual, IG KSC 705/2 - 705/14 and 705/99. Note that in the lateral  
717 view of **b**, the preserved frontal and nasal are shown even though they are in fact on  
718 the other side of the skull. **c**, diagrammatic representation of the number of specimens  
719 of different bones in the sample.

720

721 **Extended Data Figure 4 | Size and shape of Devonian tetrapods.** Silhouette  
722 reconstructions, drawn to the same scale, of the heads of the known reconstructable  
723 Devonian tetrapods. The lower jaw of *Elginerpeton*, the largest known Devonian  
724 tetrapod (for which the skull cannot be reconstructed), is also included. All  
725 reconstructions except *Acanthostega* are assembled from more than one specimen;  
726 specimen numbers indicate the specimen used to determine the scale. The right-hand  
727 column shows the largest known individuals. The left-hand column shows the  
728 smallest individuals of *Parmastega* (all from Sosnogorsk) and *Ichthyostega* (based on  
729 the entire East Greenland collection, reviewed in ref. 64). Note similarity of size range  
730 despite very different nature of samples. *Ventastega* and *Acanthostega* show narrow  
731 size ranges, which are not illustrated. Reconstructions modified from the following  
732 sources: *Ichthyostega*, ref 19; *Acanthostega*, ref 31; *Ventastega*, ref 32; *Elginerpeton*,  
733 ref. 63.

734

**Extended Data Figure 5 | relative orbit size.** Plot of orbit length vs. skull length for a range of tetrapodomorph fishes, elpistostegids, Devonian tetrapods and post-Devonian tetrapods. Data taken from ref. 47, except *Parmastega*, which is based on the largest known individual (see Extended Data Fig. 3). Post-Devonian tetrapods from ref. 47 not included in our phylogenetic analysis are not shown. *Ac*, *Acanthostega*; *Ba b*, *Baphetes bohemicus*; *Ba k*, *B. kirkbyi*; *Ba l*, *B. lintonensis*; *Bal*, *Balanerpeton*; *Be*, *Beelarongia*; *Br*, *Bruehnopteron*; *Cab*, *Cabonnichthys*; *Can*, *Canowindra*; *Cl*, *Cladarosymblema*; *Cra*, *Crassigyrinus*; *Den*, *Dendrerpeton*; *Ed*, *Edenopteron*; *Elp*, *Elpistostege*; *Eoh*, *Eoherpeton*; *Eu*, *Eusthenopteron*; *Gog*, *Gogoniasus*; *Goo*, *Gooloogongia*; *Gre*, *Greererpeton*; *Gy*, *Gyroptychius*; *He*, *Heddleichthys*; *Ich*, *Ichthyostega*; *Ko*, *Koharalepis*; *Man*, *Mandageria*; *Mar*, *Marsdenichthys*; *Meg*, *Megalocephalus*; *Oss*, *Ossinodus*; *Ost*, *Osteolepis*; *Pal*, *Palatinichthys*; *Pan*, *Panderichthys*; *Par*, *Parmastega*; *Ped*, *Pederpes*; *Pro*, *Proterogyrinus*; *Scr*, *Screbinodus*; *Sil*, *Silvanerpeton*; *Tik*, *Tiktaalik*; *Tin*, *Tinirau*; *Ven*, *Ventastega*; *Wha*, *Whatcheeria*.

**Extended Data Figure 6 | Otoccipital morphologies of Devonian tetrapods. a,** Comparative diagram of the otoccipital regions of *Parmastega*, *Ichthyostega* (new reconstruction, based on data from ref. 18, 20), *Ventastega* (modified from ref. 32) and *Acanthostega* (modified from ref. 20, semicircular canals modified from ref. 50) in ventral view. Note that the basioccipital-exoccipital complex is only preserved in *Ichthyostega* and *Acanthostega*; in these genera the inner ear is shown only on one side. Drawings are scaled to the same length from pineal region to posterior margin of otic capsule. The inner ear is represented by the grooves for the anterior and posterior oblique semicircular canals, except in *Ichthyostega* where it is represented by the



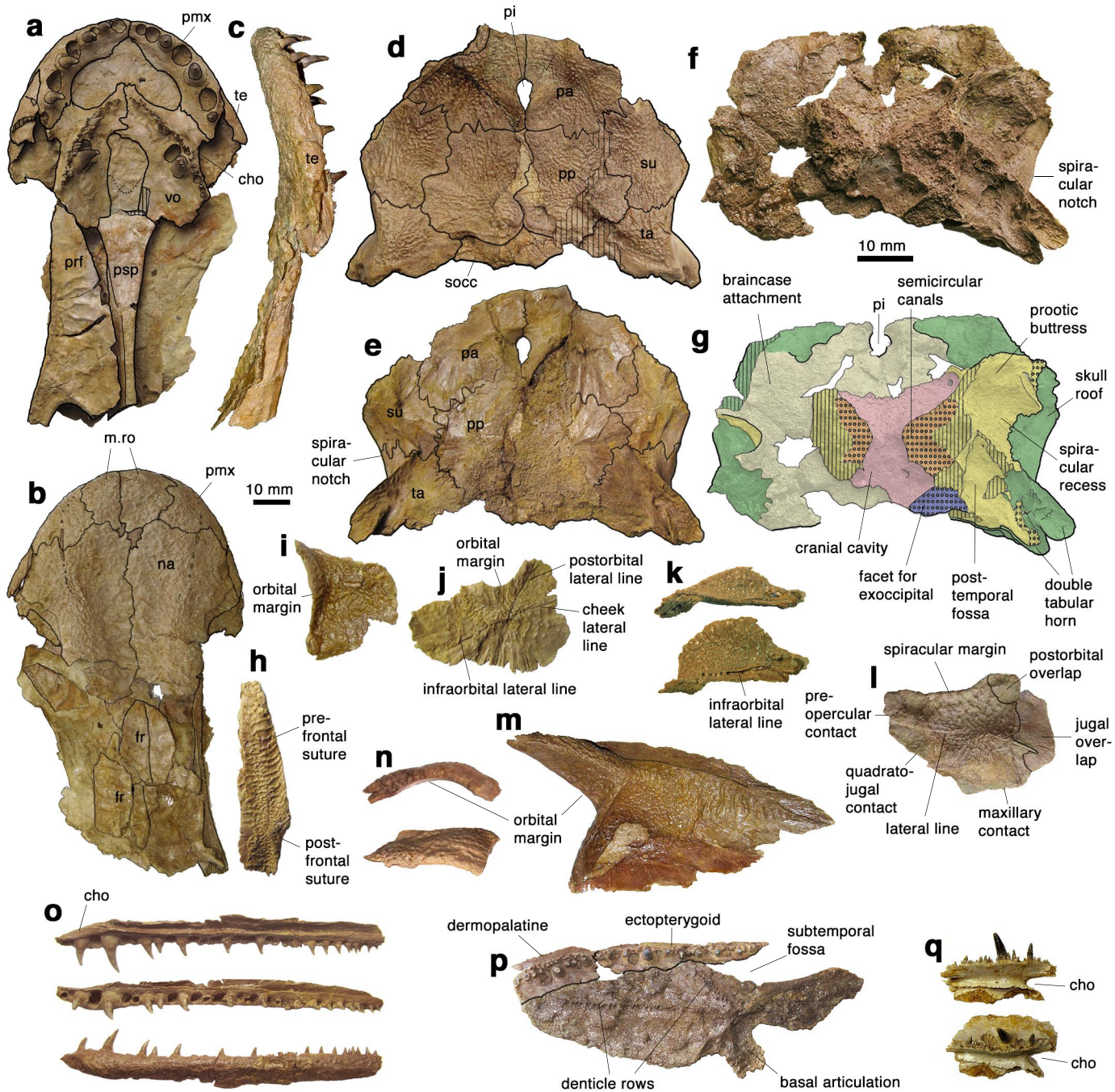
sacculus (modified from ref. 20). The braincases are arranged by morphological similarity, so that a minimum number of transformations are required along each branch. **b**, Consensus phylogeny from the analyses presented in this paper. The phylogenetic topology does not match the similarity dendrogram.

**Extended Data Figure 7 | Phylogenetic analysis.** **a**, unweighted strict consensus tree. **b**, unweighted Adams consensus tree. **c**, single tree resulting from reweighting characters by Rescaled Consistency Index. **d**, Bayesian tree, with credibility values at nodes. **e**, Maximum agreement subtree of unweighted parsimony analysis. **f**, Strict consensus of K-weighted trees. **g**, Maximum agreement subtree of K-weighted parsimony analysis. **h**, Adams consensus of all trees from all K-weighted analyses.

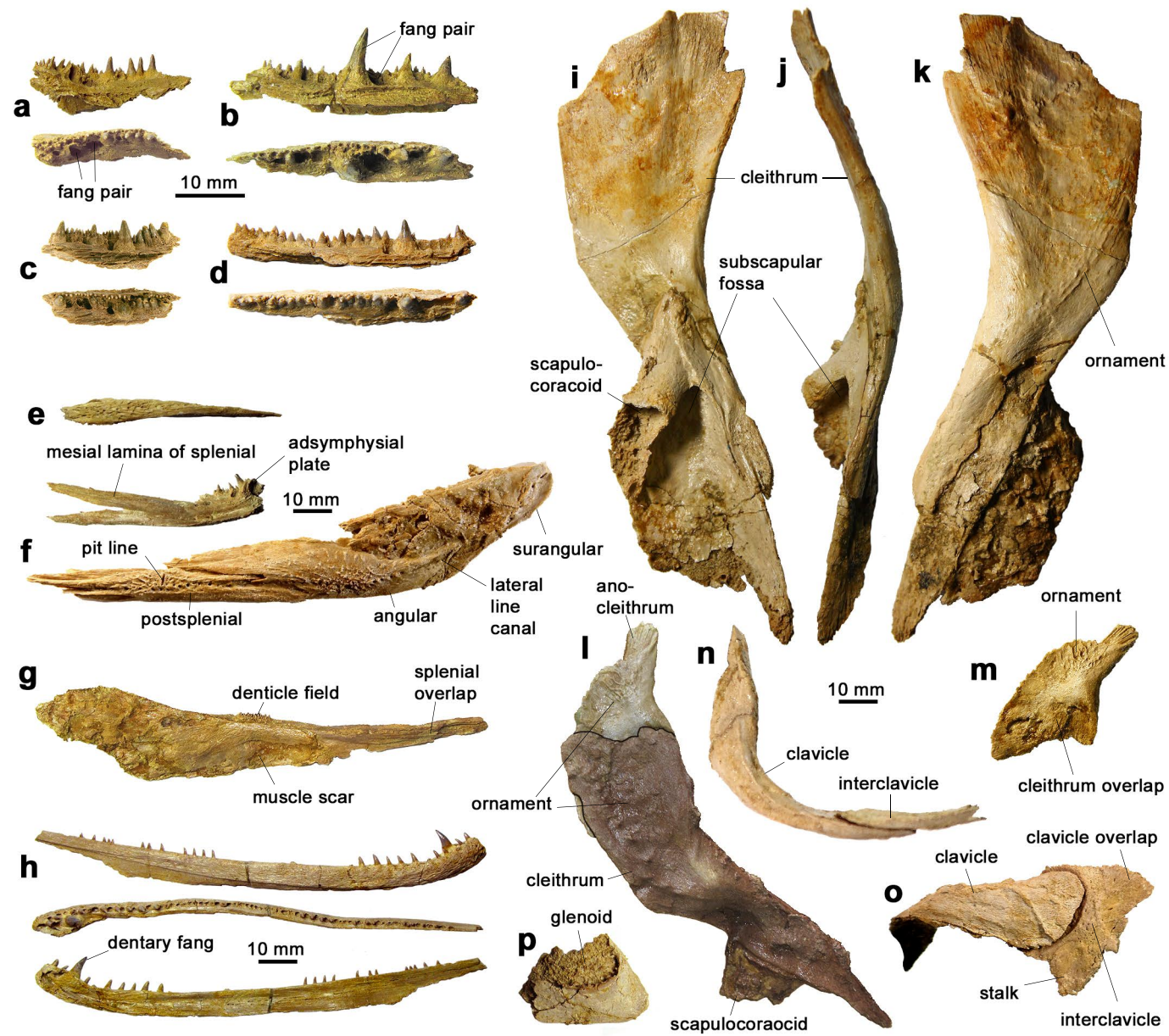
**Extended Data Figure 8 | *Parmastega* and caiman.** Comparison in left lateral view of spectacled caiman (*Caiman crocodilus*) on the left and *Parmastega* on the right, drawn to the same size, showing inferred similar cruising posture at the surface. Note the different positions of the nostrils. The caiman image is based on a CT scan in the Digimorph Archive ([http://www.digimorph.org/specimens/Caiman\\_crocodilus/](http://www.digimorph.org/specimens/Caiman_crocodilus/)).

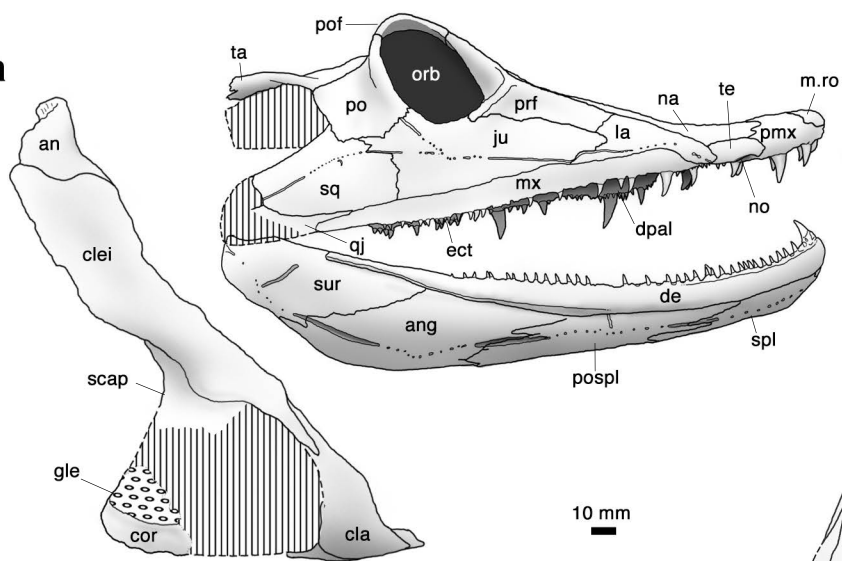
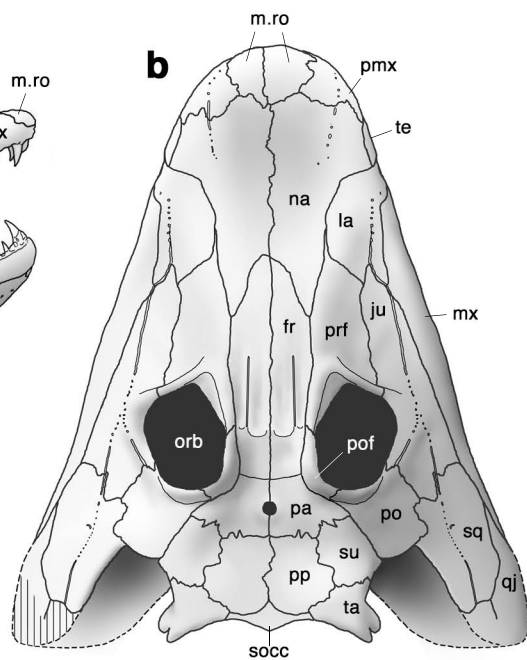
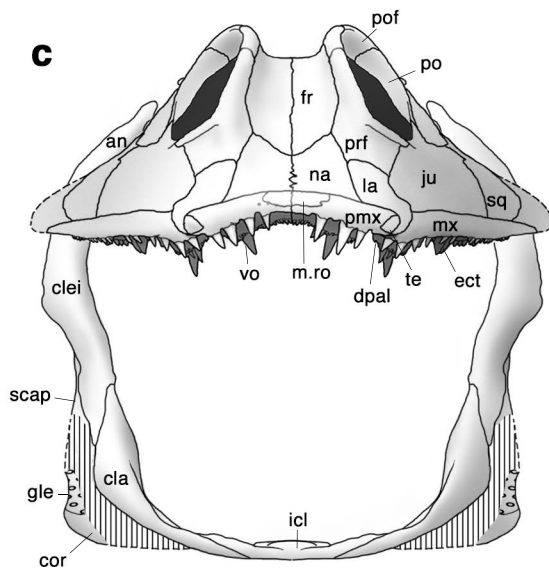
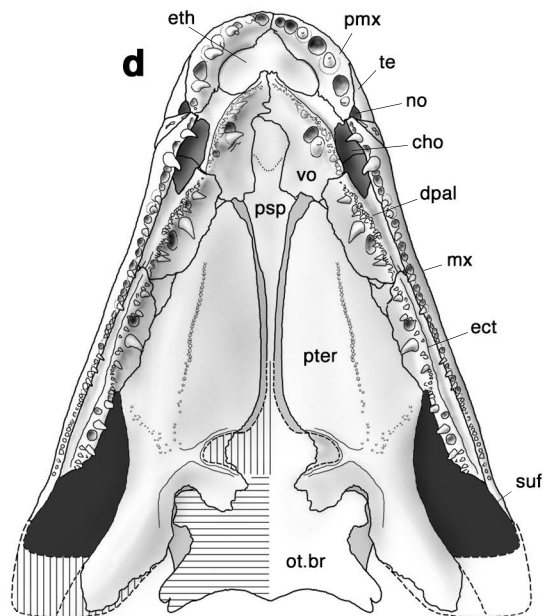
**Extended Data Figure 9 | fit of dentary against upper jaw.** **a**, dentary of *Parmastega* (IG KSC 705-67) fitted against palatal reconstruction to show the difference in curvature between the spade-shaped snout and the relatively straight dentary. **b**, lateral view of skull reconstruction of *Parmastega* with closed mouth, showing mismatch in curvature between upper and lower jaws. **c**, composite reconstruction of *Ventastega*, superimposing lower jaw rami (from ref. 30) on skull reconstruction (from ref. 32), showing shape relationship similar to **a**. Not to scale.









**a****b****c****d****e**