Supplementary Appendix

This appendix has been provided by the authors to give readers additional information about their work.

Supplement to: Novel Swine-Origin Influenza A (H1N1) Virus Investigation Team. Emergence of a novel swine-origin influenza A (H1N1) virus in humans. N Engl J Med 2009;360:2605-15. DOI: 10.1056/NEJMoa0903810.

(PDF updated May 22, 2009.)

Supplement: Laboratory

Viral RNA was extracted from clarified supernatant fluid using the Qiagen BioRobot M48 workstation with the MagAttract Viral RNA M48 kit (Qiagen, Valencia, CA). For sequence determination, degenerate primers were designed based upon the alignment of all human A (H1N1) sequences available and used in RT-PCR reactions to generate a series of overlapping template amplicons for all genome segments. An M13 sequence tag was added to the 5' end of each primer to be used for later sequencing. Primer sequences are in **Supplementary Table 1.**

RT–PCR was performed using the AccessQuick RT-PCR Kit (Promega, WI). Amplicons were prepared for sequencing by incubating them at 37 °C for 15 min and then at 80 °C for 15 min with ExoSAP-it (USB Corporation, Cleveland, OH) to inactivate remaining dNTPs and primers. Each amplicon was sequenced from each end using M13 primers (F primer: TGTAAAACGACGGCCAGT; R primer: CAGGAAACAGCTATGACC). Sequencing reaction products were resolved on an Applied Biosystems 3730 ABI sequencer. Phylogenetic analyses were performed using the Genetic Algorithm for Rapid Likelihood Inference (GARLI 0.96b7), based on General Time Reversible (GTR) + I + γ 4 substitution model.³ Results are shown in Supplementary figures 1-4.

Adamantane susceptibility was assessed by conventional sequencing and/or pyrosequencing assay⁴ using viral RNA extracted from original clinical specimens and/or virus isolates. All samples tested contained the S31N mutation in the M2 protein which has been shown to confer cross-resistance to the adamantane class of anti-influenza drugs.

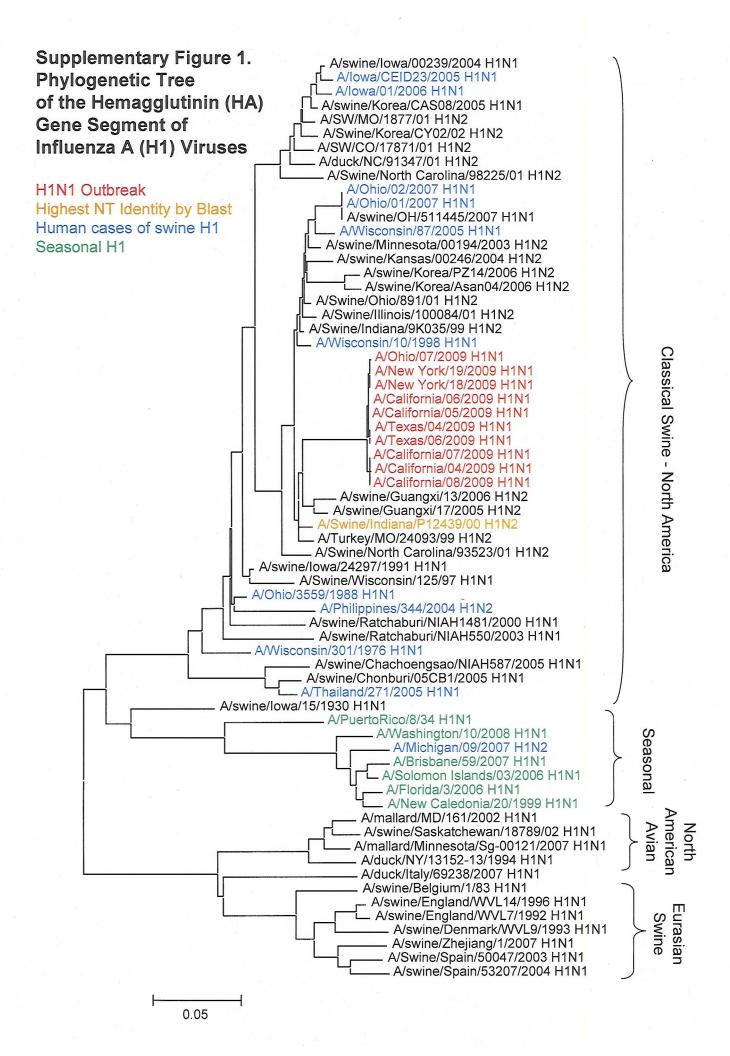
Nucleotide sequence analysis of both original clinical specimens and virus isolates revealed no predicted amino acid changes in NA previously shown to confer resistance to licensed NA inhibitors in the N1 NA subtype. Susceptibility of virus isolates to the NAIs oseltamivir and zanamivir was functionally assessed in the chemiluminescent neuraminidase inhibition assay using the NAStarTM Kit as previously described. All virus isolates (n=37) exhibited IC50 values (concentration needed to inhibit 50% of enzyme activity, nM) characteristic for the oseltamivir- and zanamivir-susceptible influenza viruses. Seasonal influenza A (H1N1) viruses were used as susceptible and resistant controls. The median IC50 value for oseltamivir was 0.54 nM, while the median for zanamivir was 0.59 nM.

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fragment 6	1169	tgt aaa acg acg gcc agt aag caa cca gra gat tgr ttc a	cag gaa aca gct atg acc ctg aga cca ytg aat ttt
fragment 7	1447	aaa acg acg gcc agt cca agy acm gag atg tca	cag gaa aca gct atg acc ttr ctc art tca ttg atg
fragment 8	1683	tgt aaa acg acg gcc agt caa tac cta yca rtg gat cat cag aa	2341 cag gaa aca gct atg acc tag tag aaa caa ggt cgt t
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fragment 2	233	aaa acg acg gcc agt caa ctc	cag gaa aca gct atg acc gtt caa gct ttt crc awa
fragment 3	389	aaa acg acg gcc	cag gaa aca gct atg acc ctg aac cay tca ggy tga
fragment 4	711	aaa acg acg gcc agt tga	cag gaa aca gct atg acc ttg aac atg ccc atc atc
fragment 5	974	aaa acg acg gcc agt aat caa aay	cag gaa aca got atg acc age tec atg etr aaa ttr
fragment 6	1139	aaa acq acq qcc agt caa ata ccy gca	atg acc cca agr tca ttg
fragment 7	1489	aaa acq acq qcc aqt atq aqy aaa aag aag tcy ta	1954 cag gaa aca gct atg acc tca aty tcy tta tgg gtg ac
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fragment 2	235	aaa acg gcc agt cca	cag gaa aca gct atg acc tga gaa agc ttg ccc tca atg
fragment 3	361	aaa acg acg gcc agt tat gay	ttc cat cca aag aat
fragment 4	702	aaa acq acq qcc aqt tgc	1292 cag gaa aca gct atg acc tcr cak gcc ttg ttg aac tca tt
fragment 5	894	aaa acg acg gc agt aaa ttr agc att gar	1662 cag gaa aca gct atg acc tcw agt cty ggg tca gtg ag
fragment 6	1204	aga acq acq acc agt tag acq att tra agc	aca gct atg acc aay
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fragment 2	318	tac aca aaa gac aay agc	cag gaa aca gct atg acc ggr cca tcg gtc att
fragment 3	536	aaa acg acg gcc agt ggt cag caa gcg cat	cag gaa aca gct atg acc cat aty tgt atg aaa acc
fragment 4	726	ggr car gcc tcr tac	cag gaa aca gct atg acc gct gct ycc rct
fragment 5	941	tgt aaa acg acg gcc agt tag gat aca tct gca gtg g	1452 cag gaa aca gct atg acc agt aga aac aag gag
Σ	forward		9
fragment 1	0	agt agc aaa agc agg tag	cag gaa aca gct atg acc gca atc tgy tca cak
fragment 2	223	acg gcc agt cac cgt gcc cag tga gcg	cag gaa aca got atg acc tca ytt gaa yug ytg
fragment 3	383	tgt aaa acg acg gcc agt tct gct ggw gca ctt gcc agt tg	102/ cag gaa aca got atg acc agt agil aac aag gra gt
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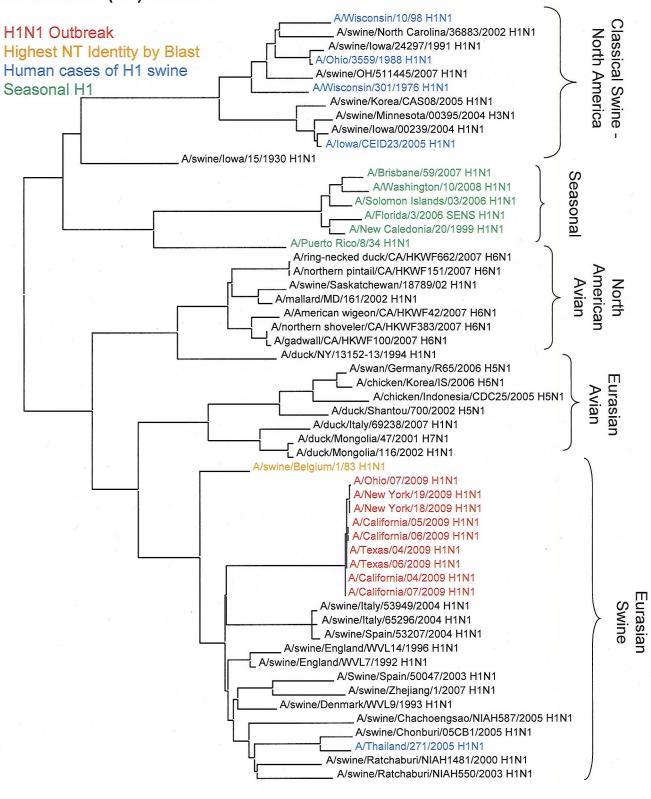
m13 forward tgt aaa acg acg gcc agt m13 reverse cag gaa aca gct atg acc

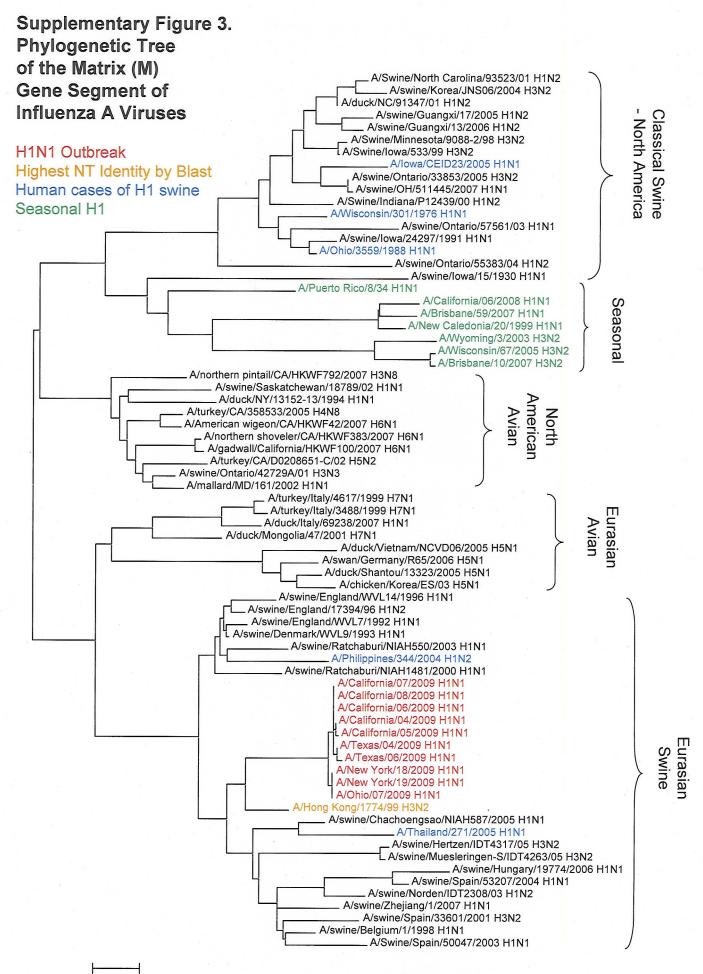
m13 forward m13 reverse



Supplementary Figure 2.
Phylogenetic Tree
of the Neuraminidase (NA)
Gene Segment of
Influenza A (N1) Viruses

0.02





Accession Numbers For Strains Included In Supplemental Figure 2:

	HA	NA	M
A/CALIFORNIA/04/2009	FJ966082	FJ966084	FJ966085
A/CALIFORNIA/05/2009	FJ966952	FJ966956	FJ966954
A/CALIFORNIA/06/2009	FJ966960	FJ971075	FJ966962
A/CALIFORNIA/07/2009	FJ969540	FJ984386	FJ969537
A/CALIFORNIA/08/2009	FJ971076		FJ969532
A/NEW YORK/18/2009	FJ984355	FJ984350	FJ984348
A/NEW YORK/19/2009	FJ984394	FJ984390	FJ984388
A/OHIO/07/2009	FJ984397	FJ969520	FJ984395
A/TEXAS/04/2009	FJ981612	FJ966981	FJ966980
A/TEXAS/06/2009	FJ984385	FJ984383	FJ984381